



This is an author produced version of a paper published in *Animal*.

This paper has been peer-reviewed but may not include the final publisher proof-corrections or pagination.

Citation for the published paper:

P. A. Harris, A. D. Ellis, M. J. Fradinho, A. Jansson, V. Julliand, N. Luthersson, A. S. Santos and I. Vervuert. (2017) Review: Feeding conserved forage to horses: recent advances and recommendations. *Animal*.

Volume: 11, Number: 6, pp 958-967.

<http://dx.doi.org/10.1017/S1751731116002469>.

Access to the published version may require journal subscription.

Published with permission from: Cambridge University Press.

Epsilon Open Archive <http://epsilon.slu.se>

1 **Review: Review of feeding conserved forage to horses: recent advances and**
2 **recommendations**

3 P.A. Harris ¹, A.D. Ellis ², M.J. Fradinho ³, A. Jansson ⁴, V. Julliand ⁵, N. Luthersson ⁶,
4 A.S. Santos ⁷ and I. Vervuert ⁸.

5 ¹ *Equine Studies Group, WALTHAM Centre for Pet Nutrition, Leics LE14 4RT, UK*

6 ² *UNEQUI, Research~Education~Innovation, Southwell, Nottinghamshire, NG25 0DS,*
7 *UK*

8 ³ *CIISA, Faculdade de Medicina Veterinária, Universidade de Lisboa, Av.*
9 *Universidade Técnica, 1300-477 Lisboa, Portugal*

10 ⁴ *Dept of Anatomy, Physiology and Biochemistry, Swedish University of Agricultural*
11 *Sciences, 750 07 Uppsala, Sweden*

12 ⁵ *AgroSup Dijon, 21079 Dijon Cedex, France*

13 ⁶ *Hestedoktoren I/S, Bukkerupvej 195, 4360 Kr. Eskilstrup, Denmark*

14 ⁷ *Department of Veterinary Medicine, Escola Universitária Vasco da Gama, 3020-*
15 *210 Coimbra / CITAB-UTAD – Center for Research and Technology of Agro-*
16 *Environmental and Biological Sciences, Vila Real, PA Box 1013, 5001- 801 Vila*
17 *Real, Portugal*

18 ⁸ *Institute of Animal Nutrition, Nutrition Diseases and Dietetics, Faculty of Veterinary*
19 *Medicine, University of Leipzig, An den Tierkliniken 9, D-04103 Leipzig, Germany.*

20 **Corresponding Author:** Pat Harris. E-mail: pat.harris@effem.com

21 **Short Title:** Feeding conserved forage to horses

22 **Abstract:** The horse is a non-ruminant herbivore adapted to eating plant-fibre or forage
23 based diets. Some horses are stabled for most or the majority of the day with limited or no
24 access to fresh pasture and are fed preserved forage typically as hay or haylage and
25 sometimes silage. This raises questions with respect to the quality and suitability of these
26 preserved forages (considering production, nutritional content, digestibility as well as
27 hygiene) and required quantities. Especially for performance horses, forage is often
28 replaced with energy dense feedstuffs which can result in a reduction in the proportion of the
29 diet that is forage based. This may adversely affect the health, welfare, behaviour and even
30 performance of the horse. In the past 20 years a large body of research work has
31 contributed to a better and deeper understanding of equine forage needs and the
32 physiological and behavioural consequences if these are not met. Recent nutrient
33 requirement systems have incorporated some, but not all, of this new knowledge into their
34 recommendations. This review paper amalgamates recommendations based on the latest
35 understanding in forage feeding for horses, defining forage types and preservation methods,
36 hygienic quality, feed intake behaviour, typical nutrient composition, digestion and
37 digestibility as well as health and performance implications. Based on this, consensual
38 applied recommendations for feeding preserved forages are provided.

39

40

41 **Key words:** forage, health, hygiene, behaviour, requirements

42

43 **Implications:** Improved consistency in the terminology used for preserved forages
44 and the adoption of consensual recommended minimal forage intake levels.

45 **Introduction**

46 The horse is a non-ruminant grazing herbivore adapted to eating plant-fibre or forage
47 based diets. A wide range of plant species can be hydrolyzed and/or fermented within
48 its specialized gastro-intestinal tract (GIT), thanks to the presence of a mixed microbial
49 symbiotic population throughout the whole GIT, but particularly in the hindgut, which
50 facilitate fibre digestion (de Fombelle *et al.*, 2003; Dougal *et al.*, 2013). Gastrointestinal
51 tract (GIT) digestion provides energy mainly through the production of short-chain fatty
52 acids (SCFA), mostly acetate, propionate and butyrate, also often described as volatile
53 fatty acids (VFA's) (Merritt and Julliand, 2013). The whole digestive system is well
54 adapted to a trickle (almost continuous intake of small amounts) feeding intake pattern,
55 with the horse naturally foraging for around 10 – 15 hours a day (Ellis, 2010).
56 Historically the energy demands of working horses were too high, and time too short,
57 to be met by forage alone, especially as forage quality was often poor, grazing limited
58 and it was difficult to distribute and transport bulky forages. Therefore, oats, barley,
59 beans and root vegetables were used as a major component of the diet (Stewart,
60 1838). The inclusion of energy rich feedstuffs (particularly cereals and vegetable oils)
61 in the diets of many horses, especially performance horses, continues today despite
62 the availability of better quality forages (Richards *et al.*, 2006, Lindburg 2013). An
63 increase in starch intake, often in combination with a reduction in the amount of forage
64 provided, can have health, welfare and performance consequences.

65 Due mainly to environmental conditions and the lack of availability/undesirability of
66 grazing, many horses are fed preserved forages (hay, haylage and sometimes silage),
67 in particular preserved grass, either at specific times of the year or all year round and
68 therefore preserved forages provide all or part of their forage intake. During the 2012
69 meeting of the European Workshop on Equine Nutrition (EWEN) it was agreed that

70 there was a need for a consensus paper on the topic of preserved forage feeding to
71 horses. The aim of this review paper is therefore to summarize recent findings and to
72 provide consensual applied recommendations for feeding preserved forage.

73

74 **Forage types and preservation methods**

75 Roughages are high fibre feeds (Morrisson, 1956) generally obtained as a crop
76 residue or a by-product (Ellis *et al.*, 1988) e.g. straw and cereal hulls. Forages are also
77 high fiber feeds obtained by cutting and preserving the whole plant (except roots).
78 Although they define different products, the words forage and roughage are typically
79 used interchangeably. From a behavioural aspect the term 'foraging' encompasses all
80 feed intake activities of horses both on pasture and in housing situations.

81 Cut grass air dried/wilted in the field or a barn is termed *hay*, whereas *silage* is forage
82 preserved moist and airtight, and thus fermented (McDonald *et al.*, 1991) The term
83 haylage was originally used to describe silage with a DM content of around 50 %
84 (Gordon *et al.*, 1961) and although haylage (and hay/silage) can be harvested at any
85 stage of plant maturity today *haylages* are typically grown and cut at later growth
86 stages similarly to hay, but baled before becoming dry resulting in DMs typically >50
87 but <70% although occasionally up to 85% (see Fig. 1 and supplementary Table S1).
88 Hay baled with too high a moisture content due to unsuitable weather conditions at
89 cutting time and especially during wilting, insufficient turning especially if in rows
90 (rather than being wilted widespread which helps speed up wilting, thereby helping to
91 reduce protein degradation and loss of nutrients) etc. allows the development of fungal
92 spores/bacteria and increases the risk of mycotoxin development. To stop undesired
93 microbial growth, roughage/forage therefore has to be preserved under controlled
94 conditions (Fig. 2). Hay and straw should be preserved during conditions of low water

95 activity in the crop (i.e. preserved at a DM content ideally above 85%) and given that
96 the bales will be exposed to air, relative humidity must be lower than required for mould
97 growth at the given temperature (Fig. 2) and therefore ideally <70%. Forages may also
98 be preserved utilizing airtight conditions, with or without lactic acid fermentation. In
99 silage the water soluble carbohydrates in the forage are fermented by anaerobic
100 lactic acid bacteria (naturally occurring on the crop or added as inoculants (Fig. 1). For
101 sufficient lactic acid production to occur, the crop DM content must ideally be around
102 30% or less and certainly <50 % (Fig. 3). If the acids produced sufficiently increase
103 hydrogen ion concentrations, undesirable microbial growth will be inhibited. To confirm
104 proper ensiling of forages (without butyrate) with a DM 15-50%, pH can be used as an
105 indicator. Weissbach, 1996 and Spörndly *et al.*, 2003 suggest that the pH should be
106 less than: $0.0257 \times \text{DM}\% + 3.71$ (Weissbach, 1996) but according to Field and Wilman,
107 1996 a higher pH ($0.0028 \times \text{DM}(\text{g}/\text{kg}) + 4.209$ (equation estimated from their figure)
108 might be allowed if silage is preserved in bales and not in bunkers. Preservation at
109 higher DM contents relies on maintenance of airtight packaging not a low pH (Mihin,
110 1940). In haylage/silage bale production, at least 4 layers of film are needed to
111 promote good preservation (Keles *et al.*, 2009; McEniry *et al.*, 2011) but adding layers
112 (6- 10) increases the CO₂ content (Müller, 2005) and for bales to cope with handling,
113 transportation, birds and long-term storage 8 layers or more are recommended
114 (Jacobsson, 2002; Spörndly pers. com.). In haylages with very high DM contents there
115 may be an increased risk of perforation (allowing air entrance and mould growth) by
116 stiff and sharp stems. To facilitate safer preservation and storage such haylages
117 require more layers of film than forages with a lower DM content. Some professional
118 horse haylage producers may use 12-20 layers (Jansson A, pers. com.). More
119 research is needed into the impact of the number of layers used. Mature and rough

120 crop may be difficult to preserve correctly since more air can be trapped in such bales,
121 increasing the risk of localized mould growth. Higher density baling, late tossing, lower
122 dry matter (75%), as well as rain occurring after cutting strongly increased mould
123 counts in hay (Seguin *et al.*, 2010) but baling for haylage when too dry (86-88% DM)
124 also increased mould counts (Martinson *et al.*, 2011).. Bales of forage with very low
125 DM contents (<30 %) are also at risk of losing airtightness due to their heavy weight,
126 plant structure collapse during fermentation, formation of effluent etc. all of which result
127 in increased pressure on the film. In forage with DM content < 40 % *clostridial*
128 fermentation can also occur (McDonald *et al.*, 2002) and the prevalence of *clostridial*
129 fermentation seems to be higher in bales with unchopped vs chopped forage (Pauly,
130 1999). In well-made haylage, whilst there will still be some microbial activity, reduction
131 in WSC and production of by-products of fermentation this will be much more limited
132 compared with that for silage (Müller *et al.*, 2007; Muhonen *et al.*, 2009).

133 Figures 1, 2 and 3 here

134 **Nutritional composition**

135 The nutritive value of forage to the horse is determined predominantly by its nutrient
136 content and digestibility, which in turn may be influenced by level of intake and feed
137 interactions. Nutrient content depends on many plant related factors as well as the
138 environment (Buxton, 1996). Herbage maturity at harvest, however, is a key influencer
139 (Virkajärvi *et al.*, 2012). As the plant matures, the fibre fraction increases and crude
140 protein decreases, leading to a decline in digestibility plus overall energy availability
141 (Ragnarsson and Lindberg, 2008; 2010; Müller, 2012) (Fig. 4). Nutrient content is also
142 closely related to plant species and morphology, with legumes typically higher in
143 protein and calcium than grasses. The proportion of leaf to stem will also affect

144 nutritive value, even between cultivars of the same species (Van Soest, 1994;
145 Bélanger and McQueen, 1997).

146 Figure 4 Here

147 Forage quality is also influenced by management factors especially fertilizer
148 applications, harvest techniques and storage conditions (Van Soest, 1994; Cookson
149 *et al.*, 2000; Rotz, 2003). Hay nutrient losses are normally higher during harvest (e.g.
150 plant leaves left in the field due to mechanical handling of the dried crop) in contrast
151 with silage where losses are larger during storage, mostly due to fermentation
152 activities, air infiltration and silage effluents (McGechan, 1989, 1990; Dürr, 2004 ;
153 Müller, 2012). Environmental factors affecting herbage development (including
154 temperature, precipitation, solar radiation and soil nutrient availability) obviously can
155 vary with geographical location resulting in a range of climatic and soil conditions.

156 Not surprisingly forage chemical composition and consequent nutritional value
157 therefore shows a great variation between regions, crop years, harvest time, seasons
158 and even within farming places (see Supplementary Table S1). The variability
159 highlights the importance of forage analysis when designing forage based rations,
160 especially when feeding brood mares, growing and performance horses (Jansson *et*
161 *al.*, 2012) or those with particular clinical conditions (such as laminitis, HYPP etc.).
162 Ideally mineral content should also be analysed. Legume forages tend to have higher
163 protein and calcium contents than grass forages and this needs to be taken into
164 consideration when formulating the total ration (which should preferably be based on
165 nutrient analysis).

166

167 **Digestion and Digestibility**

168 Digestion can be defined as the process in the gastrointestinal tract by which forage
169 constituents are converted by physical and chemical breakdown into substances
170 (mainly SCFA) that can be absorbed and assimilated by the body (Argenzio *et al.*,
171 1974). Water soluble, as well as enzymatically digestible carbohydrates (WSC, starch
172 non-starch polysaccharides, soluble fibre) are digested from the stomach onwards
173 along the whole GIT as long as they are not trapped within insoluble fibre. Insoluble
174 fibre digestion depends exclusively on microorganisms that have the ability to
175 hydrolyze the predominant β 1-4 linked polysaccharides of cell walls. It occurs primarily
176 in the horse's hindgut with a limited amount of fermentation in the foregut (de Fombelle
177 *et al.*, 2003; Jouany *et al.*, 2009). The extent of any forage digestion depends on both
178 the fibrolytic microbial activity of the hindgut ecosystem and the total time during which
179 parietal components are exposed to this activity (Merritt and Julliand, 2013). This
180 duration is defined as the mean retention time (MRT) of digesta (Miyaji *et al.*, 2008b).
181

182 Several intrinsic and extrinsic factors can affect forage digestion and digestibility
183 (digestibility = total nutrient/energy ingested minus nutrient/energy excreted and
184 expressed as % of total ingested). The Influence of intrinsic factors such as horse
185 breed, individuality and age has been studied on forage digestion and digestibility. It
186 has been anecdotally suggested, for example, that 'easy keepers' may have higher
187 digestion efficiency than other horses. However, in a controlled study (Ragnarsson
188 and Jansson, 2011) where two haylages (cut at early and late maturity stage) were
189 fed to both Icelandic horses (easy keepers) and Standardbred horses no such effect
190 could be observed (Table 1). Recent studies also showed that DM, organic matter and
191 neutral detergent fibre fraction apparent digestibility were not different between
192 weaned foals (6 and 12 months) and adults (14 years) (Ringler *et al.*, 2009; Earing *et*

193 *al.*, 2013), which complemented and confirmed previous data reporting no variation of
194 total apparent digestibility of DM between weanlings (5 months), and those aged 8
195 and 12 months old (Cymbaluk *et al.*, 1989). At two months of age, the SCFAs profile
196 of foal's faeces remained constant and cellulolytic bacterial concentration was
197 comparable with adult values suggesting that the fibre-degrading capacity in foals was
198 established by two months of age (Faubladier *et al.*, 2013).

199

200 Extrinsic factors such as botanical characteristics are known to influence growth and
201 metabolism of plants, and therefore their nutritive value as forages. In horses, the
202 average DM digestibility decreased significantly in forages having higher NDF and
203 lower crude protein contents (Edouard *et al.*, 2008). More specifically, alfalfa has
204 higher DM and CP digestibility coefficients than tall fescue, caucasian bluestem
205 (Crozier *et al.*, 1997), coastal Bermuda grass (Sturgeon *et al.*, 2000; Potts *et al.*, 2010)
206 and Matua grass (Sturgeon *et al.*, 2000). Alfalfa also had higher OM digestibility than
207 grass hays (LaCasha *et al.*, 1999) and more specifically coastal Bermuda grass (Potts
208 *et al.*, 2010). Digestion and energy and crude protein content of forage is greatly
209 affected by the stage of maturity of plants (Fig 4; Table 2).

210 The type of forage preservation, however, appears to have limited impact on apparent
211 digestibility. When hay and haylage originating from the same crop (same
212 harvest/batch) were compared, digestibility did not differ (Bergero and Peiretti, 2011)
213 although when hay and silage from the same crop was compared, digestibility was
214 slightly higher in silage compared to hay (Muhonen *et al.*, 2008b). Feed changes from
215 hay to silage or haylage (even from the same harvest/batch) may, however, affect the
216 microflora as well as the DM of the hindgut contents (Muhonen *et al.* 2008, Muhonen
217 2009) Similarly changes between forage batches with different CP contents should

218 be made slowly, as colonic pH, for example, has been shown not to be stable within
219 3 weeks after such a change (Muhonen *et al.*, 2008a) although further studies are
220 needed to assess the importance of these alterations.. Until we do know more it is
221 recommended that changes between forage batches should therefore be made
222 carefully to minimise the risk of disturbances. In support of these recommendations
223 the incidence of certain types of colic has been shown to increase especially in the
224 first 7 days (but up to 28days) after a change in forage feeding (Hillyer et al 2002).

225

226 Insert Table 1 and 2

227

228 **Hygienic quality**

229 This is a key issue, as forages fed to horses have often been reported to be of poor
230 hygienic quality (Wichert *et al.*, 2008) which can lead to significant health problems for
231 example, mould spore exposure, especially from *Aspergillus fumigatus* has been
232 implicated in the aetiology of recurrent airway obstruction in horses (Pirie 2014).
233 Although the term “poor hygienic quality” is not well defined, it may include biological
234 contaminants (e.g. pests, microorganism and their related toxins), chemical
235 contaminants (e.g. fertilizer, heavy metals), and physical contaminants such as soil.
236 In that context, feed hygiene, as stated by EU legislation (Regulation (EC) No
237 183/2005), includes all aspects that must be considered in order to produce, sell and
238 feed a safe feedstuff that will not result in any harmful effects on the animal, and
239 therefore applies to forage as well. EU legislation (EC No 32/2002) defines maximum
240 acceptable levels for a variety of contaminants in feedstuffs including for heavy metals,
241 aflatoxin B1, rye ergot and substances such as pesticides.

242

243 Several parameters can be used to evaluate hygienic quality including feel (e.g. dry,
244 clammy), smell (e.g. typical, mouldy), colour (e.g. green, bleached), macroscopic
245 findings (e.g. presence of sand, soil, dead animals) and/or microbial evaluation
246 (Wichert *et al.*, 2008, Kamphues 2013, Wolf *et al.*, 2014). Knowing the DM content is
247 key, as discussed above, as microorganisms need water for survival and multiplication
248 (Kamphues 2013). A macroscopic evaluation should routinely be undertaken for
249 obvious moulds, as well as the presence of sand/soil and other potential contaminants
250 especially poisonous plants. Senecio spp., and *Taxus baccata* have been suggested
251 to be the main poisonous plants for horses (Berny *et al.*, 2010) although regionally
252 other poisonous plants may be important e.g. vitamin D-intoxication via Golden oat
253 grass in parts of Germany (Bockisch *et al.*, 2015).

254 Much attention is given to the microbiological analysis of feedstuffs but there are
255 significant effects of the analytical methods used. Pre-treatment, incubation
256 temperature, type of incubation substrate and incubation duration can all significantly
257 influence what is cultivated (Raymond *et al.*, 2000; Müller *et al.*, 2011). For example,
258 the growth of thermophilic bacteria such as actinomyces is supported by an incubation
259 temperature of 55°C for three days, whereas the growth of mesophilic aerobic bacteria
260 is supported by a lower incubation temperature (25°C), but a longer incubation time (7
261 days, Raymond *et al.*, 2000). In some European countries, the characterization and
262 benchmarking for bacteria, moulds and yeasts in feedstuffs has been standardized
263 (Tables 3 and 4) to provide upper acceptable levels of contamination which are
264 thought not to have any adverse effects on horse health. For this purpose
265 microorganisms are classified as being either epiphytic (i.e. normal contaminants that
266 are present even under optimal conditions) or spoilage inducing. Importantly, both
267 types may impact health (e.g. respiratory problems by moulds). However, it should be

268 emphasised that whilst these benchmark levels can be used to describe spoilage, the
269 consequences on equine health through exceeding the benchmark levels requires
270 more in-depth evaluation.

271

272 Undesirable bacteria, moulds and yeasts multiply under adverse conditions such as
273 rainfall, high humidity, and high ambient temperatures or due to vectors like mites. As
274 outlined above a DM content of >85% reduces microbial activity

275

276 Mycotoxins such as ergot alkaloids, , zearalenone, deoxynivalenol, fumonisin B1 and
277 B2, ochratoxin A or aflatoxin B1 are secondary metabolites produced by fungi that are
278 capable of causing toxicity (see review Riet-Correa *et al.*, 2013). However, little is
279 known about the impact of mycotoxins on equine health status. Several mycotoxins
280 are designated as undesirable substances with maximum levels in human food,
281 whereas only aflatoxin B1 and rye-ergot are currently designated as undesirable
282 substances with maximum levels for animal feedstuffs. Further analyses, therefore,
283 may be required including detailed mycotoxin determinations e.g. clinical signs of
284 leukoencephalomalacia requires fumonisin analysis especially when corn silage with
285 corn cobs is being fed to the affected horses. Whilst it is impossible to fully eliminate
286 mycotoxins, it is important to reduce contamination by optimizing harvesting and
287 storage conditions.

288 Botulism occurs following exposure to toxins (8 different serotypes) produced by
289 anaerobic spore-forming bacterium *Clostridium botulinum* and other botulinum toxin-
290 producing clostridia (Galey 2001). In adult horses, food-borne botulism is acquired by
291 the ingestion of preformed toxins (mainly type B, C or A). Feeding big bale silage or
292 haylage have been often associated with botulism outbreaks in horses, mules or

293 cattle (e.g. Ricketts *et al.*, 1984; Divers *et al.*, 1986; Wollanke 2004; Myllykoski *et al.*,
294 2009), although there have been outbreaks linked with hay feeding (Wichtel and
295 Whitlock 1991; Johnson *et al.*, 2010) . Botulinum toxin production typically occurs due
296 to contamination with animals, soil or poultry slurry. Equivocal results have been
297 reported regarding the potential risk of silage or haylage contamination with
298 *Clostridium botulinum* spores through using the wastage from biogas anaerobic
299 digesters for fertilization (Müller *et al.*, 2013; Neuhaus *et al.*, 2015).

300

301 *Methods of sampling*

302 A good sampling procedure is crucial so that any analysis (e.g. nutrients, microbial
303 counts) represents the mean value for the whole batch. For the official feed control EU
304 legislation (EC No 691/2013) suggests for example that 5 up to 40 individual samples
305 should be aggregated to form an initial overall sample of between 1 (low specific
306 gravity, e.g. hay or straw) and 4 kg as fed (e.g. silage). .

307 *Table 3 and 4 here*

308 ***Forage Intake Behaviour and Welfare considerations***

309 Free-ranging horses perform 10-15 individual feed-bouts/day and forage for 12.5±2.5
310 hours per day with recordings as high as 18 hours (Ellis, 2010, Supplementary Table
311 S2). 'Non-foraging' bouts are rarely longer than 3 hours duration (2±1.3) (Souris *et al.*,
312 2005; Van Dierendonck *et al.*, 1996; Hallam *et al.*, 2012, Ellis *et al.*, 2015;). Even when
313 stabled, horses tend to spend a minimum time (8.5-12 hrs/day) on foraging related
314 behaviours, at times 'topping up' their food intake behaviour by ingesting wood-
315 shavings and performing coprophagy for up to 3hrs per day (Ellis *et al.*, 2006, Ellis

316 2010). Curtis *et al.* (2011) suggested the potential for wood-shaving intakes of up to
317 3.5kg in a few individuals fed DM restricted diets as part of a weight loss programme
318 and this is in line with considerable amounts of woodshavings reported in the stomach
319 of horses on a low forage diet (Boswinkel *et al* 2007) . Intake times for chopped
320 lucerne/alfalfa are about twice as fast as those for long hay (Ellis, 2010). Chopping
321 very late cut hay to 3.5 cm particle length did not shorten intake times in 8 horses but
322 adding chopped forage (~2.5cm or 4cm) to pellets at 30% inclusion rate doubled
323 concentrate feed intake times (Ellis and Hill, 2002; Ellis *et al.*, 2005). Replacing hay
324 with ground-hay pellets reduces chewing time/kg by 75% (Ellis *et al.*, 2010) which may
325 partially explain the increased intakes (to 4-5% of BW) seen by Argo *et al.* (2002) and
326 Henneke and Callaham (2009) when they fed only pellets compared to a chaff-feed or
327 hay. Elia *et al.* (2010) found that foraging through wood-shaving bedding increased by
328 3.5 hrs/day, when feeding just hay pellets compared to feeding hay. This strong
329 motivation to spend a minimum of 8 (stabled) to 12 (grazing) hours on intake
330 behaviours, irrelevant of the energy density or composition of feedstuffs is supported
331 by many studies on voluntary intake behaviour (Supplementary Table S2) and night
332 time observations of stabled horses (Ellis *et al.*, 2015). In addition stabled horses rest
333 for 3-4 hours maximum before re-commencing foraging-related behavior (Ellis *et al.*,
334 2015).

335 Furthermore, in a race horse population of 2900 animals, those fed <6.8kg/day of
336 forage showed a significant increase in abnormal behaviours including oral
337 stereotypies and weaving (McGreevy *et al.*, 1995). Lack of foraging opportunity has
338 been directly linked to the onset of oral stereotypies in foals (Nicol *et al.*, 2002) and to
339 possible stereotypic pre-cursor behaviour (increased water play and drinking,
340 locomotion) in 3 out of 5 feed-restricted ponies (Dugdale *et al.*, 2010). Appetitive

341 behaviours have a positive feedback on motivation through the brain's pleasure
342 centres which become active as the horse pursues its goal (McBride and Hemmings,
343 2005). If the motivation to forage/chew is not fulfilled, other behaviours may replace
344 the original goal achievement behaviour as highlighted in several mammals (Hughes
345 and Duncan, 1988).

346 **Health considerations**

347 Horses as previously stated are adapted to a slow and almost continual intake of a
348 diet rich in structural fibre and low in rapidly hydrolysable carbohydrates. For many
349 commonly encountered equine health issues, forage feeding management is an
350 important factor in helping to reduce risk and maintain health.

351 Particle length influences motility and transit time within the GI tract, as larger particles
352 move more slowly (Drogoul *et al.*, 2000) and forage also influences gastric emptying
353 rate and/or the passage rate through the small intestine (Jensen *et al.*, 2012). A more
354 stable (Willing *et al.*, 2009) and diverse microbial population, with a larger core, is
355 found when a forage only diet is fed especially compared to a sugar and starch rich
356 ration (Dougal *et al.*, 2014). Stabling and feeding preserved forage results in a change
357 in gut motility and higher DM faeces despite an increased water intake (Williams *et al.*,
358 2011, 2015). Horses have a lower pH in the proximal stomach during early morning
359 (1:00–9:00 AM), when stomach-fill tends to be lower especially in stabled animals
360 (Husted *et al.*, 2008). It is therefore not surprising that several nutritional risk factors
361 for equine gastric ulcer syndrome (EGUS), colic and diarrhea have been identified and
362 many of them are related to limited, reduced or a changed intake of forage (see table
363 5).

364 Recurrent airway obstruction (RAO) is the most common cause of chronic coughing
365 in horses in temperate countries, with up to 14% prevalence in the UK (Hotchkiss *et*
366 *al.*, 2007). This lower airway inflammatory disease results in a range of clinical signs
367 from exercise intolerance/poor performance, to severe expiratory dyspnea (Pirie *et al.*,
368 2002; Pirie *et al.*, 2003; McGorum and Pirie 2008). Exposure to airborne organic dust
369 (mostly endotoxins) via stabling and feeding of hay/straw with a high mould count
370 plays a primary role (Couetil and Ward 2003) as discussed above.

371 The management strategies for horses at increased risk of several conditions, such
372 as certain forms of laminitis (including those with the Equine Metabolic Syndrome and
373 Pituitary Pars Intermedia Dysfunction) as well as various muscular disorders including
374 the Equine Rhabdomyolysis Syndrome, includes reducing the intake of non-structural
375 carbohydrates (NSC) (MacLeay *et al.*, 2000; McKenzie *et al.*, 2003; Valentine *et al.*,
376 2001; Hunt *et al.*, 2008). Such horses are often put on restricted or no pasture access
377 (especially when the levels of fructans, starch and sugar are high or there is a high
378 herbage yield) and are fed a diet based on preserved forage or forage replacers (e.g.
379 commercial fibre based products) with a known and low content of NSC (<10-12%
380 DM) to minimize post-feeding glycemic and insulinaemic responses (Borgia *et al.*,
381 2011; Geor and Harris 2013; Harris *et al.*, 2013). Recent observations also indicate
382 that forage CP content might be of importance for the insulin response (Ringmark and
383 Jansson 2013).

384 Table 5 here

385 **Performance considerations**

386 Historically, the use of forage in diets of performance horses has been limited
387 (Jansson and Harris 2013). Recent studies (Connysson *et al.*, 2006; Muhonen *et al.*,

388 2008b; Connysson *et al.*, 2010; Essén-Gustavsson *et al.*, 2010; Jansson and Lindberg
389 2012; Ringmark *et al.*, 2012; Ringmark *et al.*, 2015) show that forage-only diets can
390 meet the high energy requirements of horses in very heavy training (equal to or more
391 than twice maintenance requirements) if the energy density is high enough. The
392 energy density of such forage must correspond to at least 10.5 MJ ME/kg DM
393 (Jansson and Lindberg 2012; Ringmark 2014). These studies indicate no adverse
394 effects on performance with exception for slightly lower (ca -10 %) muscle glycogen
395 contents within 3 days after high intensity work compared to a high starch diet
396 (Jansson and Lindberg 2012). However, very high muscle glycogen contents and
397 indications of rapid glycogen recovery have been documented on forage-only diets
398 providing crude protein intakes above current feeding recommendations and forage
399 WSC contents of 8.5-13.5 % of DM (Essén-Gustavsson *et al.*, 2010; Ringmark, 2014).
400 In the study by Jansson and Lindberg (2012) plasma lactate response was decreased,
401 venous pH and blood glucose was increased during exercise on the forage-only diet
402 compared to the traditional high starch diet, indicating a metabolic pattern that might
403 improve performance. A recent study (Ringmark 2014) has also shown that it is
404 possible to get Standardbred yearlings into racing condition at the age of 3, and also
405 to win races, on a high-energy forage diet supplemented only with minerals and
406 vitamins. Horses also maintained plasma volume longer during 12 h of feed
407 deprivation on a forage only-diet compared to a high starch diet (Connysson *et al.*,
408 2010). Altogether these studies suggest that high energy preserved forage can be an
409 alternative to high starch feeds in exercising horses and that such diets might promote
410 both health and performance.

411 ***Author's Recommendations for best practice***

412 Based on the above and other published work (as well as personal views where
413 stated).

414

415 A. The general use of the following terms is recommended

416

417 I. *haylage* for forages stored airtight and with DM content \geq 50%.

418 II. *silage* for forages stored airtight with DM contents below 50%

419 III. *hay* for forage preserved at a DM content ideally above 85%

420

421 B. To fully understand the value of a forage, and its impact as part of the horse's
422 diet, nutrient analysis is recommended and an estimation of the energy content
423 should be made. This becomes particularly important when feeding horses with health
424 disorders (e.g. low BCS, obesity, laminitis, PSSM).

425

426 C. Hygienic quality of forages must be a key consideration as poor hygienic
427 quality can lead to significant health problems. As a minimum, regular visual and
428 olfactory inspection should be undertaken and poor quality forage appropriately
429 disposed of. Benchmarks for microorganism contamination should be considered.

430

431 D. Changes in forage should be done gradually especially if the nutrient
432 composition (energy, protein, WSC) is unknown or known to differ considerably due
433 to changes in plant-species, growth stage and preservation technique. In such
434 instances at least 2 and possibly more than 3 weeks adaptation period may be
435 required.

436

437 In terms of forage requirements for horses it is recommended that:

438 1. The basis of any horse's diet should be fresh or preserved forage
439 (recommended particle length >2.5 cm) offered ideally ad libitum or spread
440 throughout the day to avoid prolonged periods (i.e. >4-5hrs) in a stable without
441 foraging opportunity. However, in older horses with chewing difficulties, using ground
442 high fibre products in a soaked mash form may be beneficial.

443 2. A horse requires sufficient forage fed in a form that supports species specific
444 feed intake behaviour ideally for a minimum of 8 hrs and preferably 10 hours/24hrs.

445 3. The amount of preserved forage in the ration should be calculated on a g DM
446 /kg BW rather than % of ration or an as-fed basis due to the great variation in DM
447 content of different forages.

448 4. The lower limit of daily forage intake should be 15g DM/kg BW in addition to
449 complementary (concentrate/compound) feeding, with an absolute minimum
450 recommendation of 12.5g DM/kg BW. Previously recommended minimums of forage
451 at 8-10g DM per kg BW/day are not acceptable according to the latest understanding
452 of equine ethological needs and health considerations.

453 5. Any lower amounts should only be fed solus (i.e. no other fibre provision), under
454 exceptional clinical circumstances (e.g. as low energy providing hays for animals
455 undergoing severe restriction for weight loss purposes, post-surgery etc.), under
456 veterinary supervision and with an appropriate forage vitamin/mineral/protein
457 balancer.

458 6. Straw, if required, should be introduced into the diet very slowly, which helps
459 the horse to adapt its chewing behavior and reduce the risk of impaction, although this
460 remains a significant risk with certain individuals. The risk of gastric ulceration also

461 may increase when straw is the main roughage. Some of the authors (PH, AE, VJ, NL)
462 *personally recommend* that not more than 30% of the forage DM ration should be
463 straw (other than for donkeys). Others (AJ, IV) may recommend higher proportions
464 providing the straw is of a good hygienic standard and the overall ration is balanced
465 for protein vitamins, minerals and trace elements.

466 7. For horses requiring increased energy intake due to reproduction status or
467 work level, less mature forages that have a higher energy content, should be
468 considered. Forages cut at an earlier plant maturity stage are less likely to require
469 protein supplementation.

470 8. Intakes above those recommended here may be fed or consumed by horses,
471 as long as a healthy energy balance (body condition) is maintained. If body weight
472 increases even when feeding the minimum recommended intake, a forage with a
473 lower energy content should be fed rather than reducing the dry matter forage intake.

474

475

476 **Acknowledgements:** We thank all committee members, participants, sponsors and
477 organizers of the European Workshop in Equine Nutrition (EWEN) for their input and
478 support. In addition, special thanks are expressed to Manfred Coenen, Sarah
479 Ralston, Thomas Pauly, Rolf Spörndly, Cecilia Müller, Markku Saastamoinen, Dag
480 Austbø, Anne-Helene Tauson, Sveinn Ragnarsson, Joaquin Clotet, Samy Julliard,
481 Nicoletta Miraglia, Pier Giorgio Peiretti, Teresa Dentinho, Luis Ferreira, Rui Bessa
482 and Andreas Olt.

483

484

485 **References from 2010 onwards (The list of references from 2009 and before is given in**
486 **Supplementary Material S1).**

- 487 Andrews FM, Larson C and Harris P 2015. Nutritional management of gastric ulceration.
488 Equine Veterinary education (in press).
- 489 Bergero D and Peiretti PG 2011. Intake and Apparent Digestibility of Permanent Meadow Hay
490 and Haylage in Ponies. Journal of Equine Veterinary Science 31, 67-71.
- 491 Berny [P](#), [Caloni F](#), [Croubels S](#), [Sachana M](#), [Vandenbroucke V](#), [Davanzo F](#) and [Guitart R](#)
492 2010. Animal poisoning in Europe. Part 2: Companion animals. Veterinary Journal 183,
493 255-9.
- 494 [Bockisch F](#), [Aboling S](#), [Coenen M](#), and Vervuert [J](#) 2015. Yellow oat grass intoxication in
495 horses: Pitfalls by producing hay from extensive landscapes? A case report.
496 Tierärztliche Praxis Grosstiere Nutztiere 43, 296-304.
- 497 Borgia L, Valberg S, McCue M, Watts K. and Pagan J 2011. Glycaemic and insulinaemic
498 responses to feeding hay with different non-structural carbohydrate content in control and
499 polysaccharide storage myopathy-affected horses. Journal of animal physiology and
500 animal nutrition 95, 798-807.
- 501 Connysson M, Essén-Gustavsson B, Lindberg J E and Jansson A 2010. Effects of feed
502 deprivation on Standardbred horses in training fed a forage-only diet and a 50:50 forage-
503 oats diet. Equine Veterinary Journal Suppl. 38, 335-340.
- 504 Curtis G, Barfoot C, Dugdale A, Harris P and Argo C 2011. Voluntary ingestion of wood
505 shavings by obese horses under dietary restriction. British Journal of Nutrition 106, S178–
506 S182.
- 507 Dougal K, de la Fuente G, Harris PA, Girdwood SE, Pinloche E, Geor RJ, Nielsen BD, Schott
508 II HC, Elzinga S and Newbold CJ 2014. Characterisation of the faecal bacterial community
509 in adult and elderly horses fed a high fibre, high oil or high starch diet using
510 pyrosequencing. *PloS one*, 9 p.e87424.

511 Dougal K, de la Fuente G, Harris PA, Girdwood SE, Pinloche E and Newbold CJ 2013.
512 Identification of a core bacterial community within the large intestine of the horse. *PloS one*
513 8, p.e77660.

514 Dugdale AHA, Curtis GC, Cripps P, Harris PA and Argo C 2010. Effect of dietary restriction
515 on body condition, composition and welfare of overweight and obese pony mares. *Equine*
516 *Veterinary Journal* 42, 600-610.

517 Durham A 2013. Intestinal disease In *Equine Clinical and Applied nutrition*. (ed. RJ Geor, PA
518 Harris and M Coenen), pp 568-581. Elsevier, Amsterdam, the Netherlands.

519 Earing JE, Lawrence LM, Hayes SH, Brummer M and Vanzant E. 2013. Digestive capacity in
520 weanling and mature horses. *Journal of Animal Science* 91, 2151-2157.

521 Elia JB, Hollis N, Houpt KA 2010. Motivation for hay: Effects of a pelleted diet on behavior and
522 physiology of horses. *Physiology and Behavior* 101, 623–627.

523 Ellis AD 2010. Biological basis of behaviour and Feed Intake in horses. In *The impact of*
524 *Nutrition on the Health and Welfare of Horses* (ed. AD Ellis, A Longland, M Coenen and
525 N Miraglia), pp. 53-74. EAAP Publication No. 128, Wageningen Academic Publishers,
526 Wageningen, the Netherlands.

527 Ellis AD, Redgate S, Zinchenko S, Owen H, Barfoot C and Harris P 2015. The effect of
528 presenting forage in multi-layered haynets and at multiple sites on night time budgets of
529 stabled horses, *Applied Animal Behaviour Science* 171, 108–116.

530 Essén-Gustavsson B, Connysson M and Jansson A 2010. Effects of crude protein intake from
531 forage-only diets on muscle amino acids and glycogen levels in horses in training. *Equine*
532 *Veterinary Journal* 42, 341-346.

533 Faubladiere C, Julliand V, Danel J and Philippeau C 2013. Bacterial carbohydrate-degrading
534 capacity in foal faeces: changes from birth to pre-weaning and the impact of maternal
535 supplementation with fermented feed products. *British Journal of Nutrition* 110, 1040-1052.

536 Fedtke A, Pfaff M, Volquardsen J, Venner M and Vervuert I (2015): Effects of alfalfa chaff on
537 gastric mucosa in weanling foals. *Pferdeheilkunde* 31, 596.

538 Geor RJ and Harris PA 2013. Laminitis. In Equine Applied and Clinical Nutrition, Health,
539 Welfare and Performance. Saunders, Elsevier (ed. RJ Geor, PA Harris and M Coenen), pp.
540 469-486. Elsevier, Amsterdam, the Netherlands.

541 Hallam S, Campbell EP, Qazamel M, Owen H and Ellis AD 2012. Effects of traditional versus
542 novel feeding management on 24 hour time budget of stabled horses. In Forages and
543 Grazing in Horse Nutrition (ed. M Saastamoinen, MJ Fradinho, AS Santos, N Miraglia), pp.
544 319-321. EAAP Publication No. 132, Wageningen Academic Publishers, Wageningen, the
545 Netherlands.

546 Harris PA, Coenen M and Geor RJ 2013. Controversial areas in equine nutrition and feeding
547 management : the editors' views. In Equine Clinical and Applied nutrition (ed. RJ Geor,
548 PA Harris and M Coenen), pp 455-468. Elsevier, Amsterdam, the Netherlands.

549 Jansson A and Harris P 2013. A bibliometric review on nutrition of the exercising horse from
550 1970 to 2010. Comparative Exercise Physiology 9, 169-180.

551 Jansson A and Lindberg JE 2012. A forage-only diet alters the metabolic response of horses
552 in training. Animal 6, 1939–1946.

553 Jansson A, Saastamoinen M and Lindberg JE 2012. Forage feeding systems. In Forages and
554 grazing in horse nutrition (ed. MT Saastamoinen, MJ Fradinho, AS Santos and N Miraglia),
555 pp. 289-304. EAAP publication No. 132, Wageningen Academic Publishers, Wageningen,
556 The Netherlands.

557 Jensen RB, Austbø D and Tauson AH 2012. Feeding forage before or after oats affects
558 caecum pH profiles in the horse. In: Forages and grazing in horse nutrition (ed. MT
559 Saastamoinen, MJ Fradinho, AS Santos and N Miraglia), pp 327-330. EAAP publication
560 No. 132, Wageningen Academic Publishers, Wageningen, The Netherlands.

561 Johnson [AL](#), [McAdams SC](#) and [Whitlock RH](#) 2010. Type A botulism in horses in the United
562 States: a review of the past ten years (1998-2008). Journal of Veterinary Diagnostic
563 Investigation 22, 165-73.

564 Kamphues J 2013. Feed hygiene and related disorders in horses. In: Equine applied and
565 clinical nutrition (ed. RJ Geor, PA Harris and M Coenen), pp 367-380. Elsevier,
566 Amsterdam, the Netherlands.

567

568 Lindburg JE 2013 Feedstuffs for horses In: Equine applied and clinical nutrition (ed. RJ Geor, PA
569 Harris and M Coenen), pp 319 -331. Elsevier, Amsterdam, the Netherlands. Martinson K,
570 Coblenz, W and Sheaffer C 2011. The Effect of Harvest Moisture and Bale Wrapping on
571 Forage Quality, Temperature, and Mold in Orchardgrass Hay. Journal of Equine Veterinary
572 Science 31, 711-716.

573 McEniry J, Forristal PD and O'Kiely P 2011. Factors influencing the conservation
574 characteristics of baled and precision-chop grass silages. Irish Journal of Agricultural and
575 Food Research 50, 175-188.

576 Merritt AM and Julliard V 2013 Gastrointestinal physiology. In Equine Clinical and Applied
577 nutrition (ed. RJ Geor, PA Harris, and M Coenen), pp 3-32. Elsevier, Amsterdam, the
578 Netherlands.

579 Müller CE, Hultén C, and Gröndahl G 2011. Assessment of hygienic quality of haylage fed to
580 healthy horses. Grass and Forage Science 66, 453-463

581

582 Müller, C.E. 2012. Equine digestion of diets based on haylage harvested at different plant
583 maturities. Animal Feed Science and Technology 177, 65-74.

584 Müller CE, Johansson M, Salomonsson AC and Albiñ A 2013. Effect of anaerobic digestion
585 residue vs. livestock manure and inorganic fertilizer on the hygienic quality of silage and
586 haylage in bales. Grass and Forage Science 69, 74-89

587 Neuhaus [J](#), [Schrödl W](#), [Shehata AA](#) and [Krüger M](#) 2015. Detection of Clostridium botulinum
588 in liquid manure and biogas plant wastes. Folia Microbiologica 60, 451-6.

589 Potts L, Hinkson J, Graham B, Löest C and Turner J 2010 Nitrogen Retention and Nutrient
590 Digestibility in Geldings Fed Grass Hay, Alfalfa Hay, or Alfalfa Cubes. *Journal of Equine*
591 *Veterinary Science* 30, 330-334.

592 Ragnarsson S and Jansson A 2011. A comparison of grass haylage digestibility and metabolic
593 plasma profile in Icelandic and Standardbred horses. *Journal of animal physiology and*
594 *animal nutrition* 95, 273-279.

595 Ragnarsson S and Lindberg JE 2010. Nutritional value of mixed grass haylage in Icelandic
596 horses. *Livestock Science* 131, 83-87.

597 Riet-Correa F, Rivero R, Odriozola E, Adrien Mde L, Medeiros RM, Schild AL 2013.
598 Mycotoxicoses of ruminants and horses. *Journal Veterinary Diagnostic Investigation* 25,
599 692-708.

600 .

601 Ringmark S, Roepstorff L, Essén-Gustavsson B, Revold T, Lindholm A, Hedenström U,
602 Rundgren M, Ögren G and Jansson A 2012. Growth, training response and health in
603 Standardbred yearlings fed a forage-only diet. *Animal* 7, 746-753.

604 Ringmark S, Lindholm A, Hedenstrom U, Lindinger M, Dahlborn K, Kwart C, and Jansson
605 A 2015. Reduced high intensity training distance had no effect on VLa4 but attenuated
606 heart rate response in 2-3-year-old Standardbred horses. *Acta Veterinaria Scandinavica*
607 57, 1.

608 Ringmark S, Jansson A 2013. Insulin response to feeding forage with varying crude protein
609 and amino acid content in horses at rest and after exercise. *Comparative Exercise*
610 *Physiology* 9, 209-217.

611 Ringmark S 2014. A forage-only diet and reduced high intensity training distance in
612 Standardbred horses. PhD thesis, Swedish University of Agricultural Sciences, Uppsala,
613 Sweden.

614 Sarkijarvi S, Sormunen-Cristian R, Heikkila T, Rinne M and Saastamoinen M 2012. Effect
615 of grass species and cutting time on *in vivo* digestibility of silage by horses and sheep.
616 Livestock Science 144, 230-239.

617 Séguin V, Lemauviel-Lavenant S; Garon D, Bouchart V, Gallard Y, Blanchet B, Diquelou
618 S, Personeni E, Gauduchon P, Ourry A 2010. Effect of agricultural and environmental
619 factors on the hay characteristics involved in equine respiratory disease. Agriculture,
620 Ecosystems and Environment 135, 206-215.

621 Virkajärvi P, Saarijärvi K, Rinne M and Saastamoinen M 2012. Grass physiology and its
622 relation to nutritive value in feeding horses. In Forages and grazing in horse nutrition (ed.
623 MT Saastamoinen, MJ Fradinho, AS Santos and N Miraglia), pp. 17-43. EAAP publication
624 No. 132. Wageningen Academic Publishers, Wageningen, The Netherlands.

625 Williams S, Horner J, Orton E, Green M, McMullen S, Mobasher A and Freeman SL 2015,
626 Water intake, faecal output and intestinal motility in horses moved from pasture to a stabled
627 management regime with controlled exercise. Equine Veterinary Journal 47, 96–100.

628 Williams S, Tucker CA, Green M J. and Freeman SL 2011. Investigation of the effect of
629 pasture and stable management on large intestinal motility in the horse, measured using
630 transcutaneous ultrasonography. Equine Veterinary Journal, 43, 93–97.

631 Wolf P, Siesenop U, Verspohl J and Kamphues J 2014. Hygienic quality of feedstuffs for small
632 mammals sent to the consultation service. Tierärztliche Praxis Kleintiere 42, 101-106.

633

634

635 Table 1. *Coefficients of total tract apparent digestibility in Icelandic and Standardbred*
636 *horses fed the same early and late cut forages (after Ragnarsson and Jansson, 2011)*

	Early cut Icelandic	Early cut Standardbred	Late cut Icelandic	Late cut Standardbred	SE
OM	0.536 ^a	0.565 ^b	0.431	0.427	0.006
CP	0.636	0.660	0.478	0.479	0.012
NDF	0.517	0.536	0.322	0.320	0.008
ADF	-	-	-	-	-
Energy	0.517 ^a	0.540 ^b	0.407	0.400	0.007

637 ^{a, b}. Values in the same row without common superscripts differ ($p < 0.05$) and indicate breed
638 differences.

639 OM : Organic matter, CP : Crude Protein; NDF : Neutral detergent fibre; ADF : Acid Detergent
640 fibre

641

642

643

644

645 Table 2. *Coefficients of total tract apparent digestibility of haylages cut (first cut) at different*
646 *stages of maturity (after Ragnarsson and Lindberg 2008; Ragnarsson and Lindberg, 2009)*

647

	Timothy haylage					Mixed grass haylage				
	Cut 1	Cut 2	Cut 3	Cut 4	SE	Cut 1	Cut 2	Cut 3	Cut 4	SE
DM	0.716 ^a	0.626 ^b	0.513 ^c	0.457 ^d	0.011	0.691 ^a	0.616 ^b	0.619 ^b	0.556 ^c	0.010
OM	0.747 ^a	0.647 ^b	0.527 ^c	0.485 ^c	0.010	0.710 ^a	0.630 ^b	0.640 ^b	0.578 ^c	0.010
CP	0.809 ^a	0.735 ^b	0.642 ^c	0.639 ^c	0.014	0.765 ^a	0.740 ^{ab}	0.708 ^{ab}	0.688 ^b	0.015
NDF	0.770 ^a	0.646 ^b	0.516 ^c	0.440 ^d	0.009	0.717 ^a	0.584 ^{bc}	0.594 ^b	0.520 ^c	0.013
ADF	0.746 ^a	0.629 ^b	0.480 ^c	0.400 ^d	0.008	0.685 ^a	0.545 ^b	0.555 ^b	0.485 ^b	0.017
Energy	0.733 ^a	0.633 ^b	0.515 ^c	0.468 ^c	0.006	0.670 ^a	0.600 ^b	0.595 ^b	0.560 ^b	0.011

648 ^{a, b, c}, Values in the same row without common superscript differ (p<0.05).

649 DM : Dry matter; OM : Organic matter, CP : Crude Protein; NDF : Neutral detergent fibre; ADF
650 : Acid Detergent fibre

651

652

653

654 Table 3: *Classification of microorganisms in feedstuffs according to Kamphues (2013)*

Microorganism	Classification	Category	Species examples
Aerobic bacteria	Epiphytic	1	<i>Flavobacterium</i>
			<i>Pseudomonas</i>
			<i>Xanthomonas</i>
	Spoilage	2	<i>Erwinia</i>
			<i>Bacillus spp</i>
			<i>Staphylococcus</i>
Aerobic moulds	Epiphytic	3	<i>Micrococcus</i>
			<i>Streptomyces spp.</i>
			<i>Verticillium</i>
	Spoilage	4	<i>Acremonium</i>
			<i>Fusarium</i>
			<i>Aurebasidium</i>
Yeasts	Spoilage	5	<i>Aspergillus</i>
			<i>Penicillium</i>
			<i>Scopulariopsis</i>
		6	<i>Wallemia</i>
		6	<i>Mucor spp.</i>
		7	All species

655

656

657

658

659

660 Table 4: *Benchmark for microorganisms in feedstuffs (cfu per g feed) according to Kamphues*
661 *(2013)*

Category ¹	Aerobic bacteria x 10 ⁶ cfu, g			Moulds x 10 ³ cfu, g			Yeasts x 10 ³ , g
	1	2	3	4	5	6	7
Hay	30	2	0.15	200	100	3	150
Straw	100	2	0.15	200	100	5	400
Grass silage	0.2	0.2	0.01	5	5	5	200
Corn silage	0.4	0.2	0.03	5	5	5	1000

662 ¹*Categories; 1: Epiphytic bacteria; 2 and 3: Spoilage indicating bacteria; 4: Epiphytic moulds; 5*
663 *and 6: Spoilage indicating moulds; 7: yeasts all species*

664

665

666

667 Table 5 Some suggested forage related risk factors for gastric ulcers and colic (see also Andrews
 668 *et al.*, 2015, Durham, 2013)

Disease	Risk factor	Possible (non-exclusive) explanation	Ref
Gastric ulcers squamous	Frequency of forage feeding. Intervals > 6h may increase risk of ulcers	Decreased production of saliva, slower passage rate and reduced buffering capacity in the stomach	Luthersson <i>et al.</i> , 2009
Gastric ulcers squamous	Straw as the main forage source in horses/ponies may increase risk of ulcers	Straw may provide low levels of additional buffering support (low in protein, low in calcium). Straw may create mucosal irritation, and may affect the nature of the fibrous mat within the stomach. In addition, the potential for poor hygienic quality (including increased risk of mycotoxins) may play a role.	Luthersson <i>et al.</i> , 2009 Wichert <i>et al.</i> , 2008
Gastric ulcer - glandular	Feeding alfalfa chaff increased the number of glandular ulcers compared to	The glandular lesions could be a result of mechanical injury caused by the very small particles/physical	Fedtke <i>et al.</i> , 2015

hay or alfalfa pellets properties of the alfalfa
 in weanlings or adult chaff
 horses

Colic – in Reduced intake of Any change in forage Cohen *et al.*, 2000
 general grass*, limited intake intake will cause changes Hassel *et al.*, 2004
 of forage. Importantly in the microflora in the Hillyer *et al.*, 2002
 a change in the type hindgut. This may cause Hudson *et al.*, 2001
 of forage fed. * NB increased risk of Eg. Change of hay within
 Horses with dysfermentation, change in two weeks without
 duodenitis-proximal motility, and several adaptation (Tinker *et al.*,
 jejunitis were were physiological changes in 1997; Cohen *et al.*,
 significantly more likely the GI tract. A change in 1999). Change between
 to have grazed pasture hay within previous 2 two silages with different
 than the control weeks may increase risk of crude protein content
 population (Cohen et colic between 4.9 to 10 (Muhonen *et al.*, 2008).
 al., 2006) times. Change from hay to
 haylage or silage from the
 same botanical origin
 (Muhonen *et al.*, 2009).

Colic - Feeding costal Indigestible fibers have Little and Blikslager 2002
 impaction Bermuda hay reduced water holding and
 releasing capacity, can
 cause changes in the
 microflora and can cause
 altered motility

670 **Figures**

671 Figure 1. A very schematic overview giving some definitions and characteristics of hay, haylage
672 and silage (WSC=water soluble carbohydrates) made without additives. It is important to note
673 that DM content (and water activity) will determine the potential for fermentation so are linked with
674 lactate production. .In general as lactate starts to be produced the WSC and pH will start to
675 reduce, although other factors can influence pH (including buffer capacity). The WSC of any
676 preserved crop will be lower than the fresh forage it originated from due to respiration (plant and
677 microbial) post harvesting but the WSC content of haylage compared with the hay from the same
678 fresh pasture will depend on whether there have been any lactic acid (or VFA production) If there
679 is virtually no lactic acid (or VFA) production, then there will be no decrease in total WSC in
680 haylage compared to hay (Muhonen et al 2009).

681

682 Figure 2. A schematic figure on how minimum required air humidity (relative humidity, RH) for
683 mould growth varies with ambient temperature (adapted from Lehmann, 1971).

684 Figure 3. The relationship between airtight stored grass forage dry matter (DM) and lactate
685 content. At DM contents higher than 50 % (dotted line) the lactate content is very low. Data from
686 Wilkinson *et al.*, 1976 (3 observations, 16- 45%DM); Müller, 2007 (10 observations 29-68%DM);
687 Muhonen, 2008 (8 observations, 36-81%DM); Ragnarsson, 2009 (8 observations, 36-81%) and
688 Sarkijärvi *et al.*, 2012 (6 observations, 36-55%DM). All observations are from grass forage
689 preserved in wrapped bales except for Wilkinson *et al.*, 1976 which were in silos. Forages were
690 preserved without additives except 6 observations, with lactic acid inoculant, from Muhonen, 2008
691 and formic acid inoculant in the Sarkijärvi *et al.*, 2012 observations. Data fits ($R^2=0.84$) the
692 equation: $y = 0,0406x^2 - 5,4306x + 179.3$ and the equation is relevant in the interval 16-70 % DM.

693 Figure 4. A schematic illustration (not to scale)of the maturation of plants and the effect on dry
694 matter content (NB in Southern European countries the increase in DM is more dramatic than for
695 Northern Europe as illustrated by the two lines), digestible plus metabolizable (ME) and Net (NE)
696 energy plus digestible protein content on a wet matter (WM) basis. The extent of such changes
697 may be affected by environmental conditions as well as management practices.

698