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Chemical and Physical Mechanisms of Fungal Bioweathering of Rock Phosphate

Gilberto de Oliveira Mendes^{a,b} , Jaleh Bahri-Esfahani^{a,c}, Laszlo Csetenyi^d, Stephen Hillier^{e,f} , Timothy S. George^c , and Geoffrey Michael Gadd^{a,g} 

^aGeomicrobiology Group, School of Life Sciences, University of Dundee, Dundee, UK; ^bLaboratório de Microbiologia e Fitopatologia, Instituto de Ciências Agrárias, Universidade Federal de Uberlândia, Monte Carmelo, Brazil; ^cEcological Sciences, The James Hutton Institute, Dundee, UK; ^dConcrete Technology Group, Department of Civil Engineering, University of Dundee, Dundee, UK; ^eEnvironmental and Biochemical Sciences, The James Hutton Institute, Aberdeen, UK; ^fDepartment of Soil and Environment, Swedish University of Agricultural Sciences (SLU), Uppsala, Sweden; ^gState Key Laboratory of Heavy Oil Processing, Beijing Key Laboratory of Oil and Gas Pollution Control, College of Chemical Engineering and Environment, China University of Petroleum, Beijing, China

ABSTRACT

This research has investigated fungal transformations of rock phosphate (RP) by geoactive fungi, with particular emphasis on *Aspergillus niger*. Direct hyphal interaction with RP particles induced morphological and mineralogical changes, as revealed by scanning electron microscopy (SEM) and X-ray diffraction (XRD) analysis. The formation of the oxalate mineral calcium oxalate monohydrate (whewellite, CaC₂O₄·H₂O) on RP surfaces showed that mycogenic oxalic acid was driving the chemical dissolution of apatite, with consequent phosphate release and secondary mineral formation. This was supported by abiotic testing of common fungal excreted organic acids which confirmed that oxalic acid was the only effective RP transforming agent and therefore responsible for the morphological and mineralogical changes observed in RP when exposed to fungal colonization. Cryogenic SEM provided evidence of fungal penetration and tunneling through RP particles demonstrating that physical interactions are also important for RP bioweathering, as well as biochemical mechanisms. These findings emphasize the important role of fungi in P cycling, with active participation in the transformation of mineral phosphates through physicochemical mechanisms and secondary oxalate biomineral formation.

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Aspergillus niger; calcium oxalate; fungi; oxalic acid; rock phosphate; whewellite

Introduction

Phosphates are most commonly found in sparingly-soluble mineral forms within terrestrial and marine environments and the breakdown of these minerals is required to provide bioavailable phosphate to natural ecosystems, particularly by organisms colonizing new environments such as volcanic flows, glacial deposits, and newly eroded or deposited land surfaces (Gadd 2007, 2017a; Pollard 2018). However, in many agricultural systems phosphorus (P) is a limiting nutritional factor for crop yield, so is frequently added to soil to compensate for inadequate levels of plant-available phosphate (Cordell et al. 2009; Roy et al. 2016). Such addition to preexisting total P levels in soil can have environmental repercussions, including effects on soil biodiversity, plant adaptation, and freshwater eutrophication (Cordell and Neset 2014; Rockström et al. 2009; Withers et al. 2015). Most of the P added to soil as fertilizer is derived from rock phosphate (RP), a natural multi-mineral substrate formed under high pressure over an extended geological time period (Oelkers and Valsami-Jones 2008). Much of this is industrially processed into fertilizer before soil application, but in developing countries or organic farming systems where

fertilizer use is not possible or prohibited, then rock phosphates are often added as the primary source of additional P (Rajan et al. 1996; Zapata and Roy 2004).

Many microorganisms and plants are capable of bioweathering RP, and other insoluble phosphate-containing minerals, to liberate free phosphate (Arcand and Schneider 2006; Gadd 2007, 2017a; Kang et al. 2019, 2020; Richardson and Simpson 2011; Sullivan and Gadd 2019; Suyamud et al. 2020), and fungi are particularly important in this regard. Free-living and symbiotic fungi are intimately involved in influencing phosphate availability in the rhizosphere as well as playing a role in soil bioremediation (Adeyemi and Gadd 2005; Fomina et al. 2007; Hagerberg et al. 2003; Sullivan and Gadd 2019; Whitelaw 1999). Many species of fungi can release soluble phosphate and metal species from minerals, predominantly mediated by the excretion of organic acids and protons (Gadd 1999, 2007; Gadd et al. 2014; Jones 1998; Jones et al. 2003; Mendes et al. 2014; Suyamud et al. 2020; Whitelaw 1999). This can result in the formation of secondary biogenic minerals, for example, calcium oxalate (Burford et al. 2003; Fomina et al. 2010; Gadd 2017b; Gadd et al. 2014). Magnesium oxalate (glushinskite) resulted during

CONTACT Geoffrey Michael Gadd  g.m.gadd@dundee.ac.uk  Geomicrobiology Group, School of Life Sciences, University of Dundee, Dundee, DD1 5EH, Scotland, UK

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solubilization and phosphate release from the insoluble phosphate mineral struvite (magnesium ammonium phosphate – $\text{MgNH}_4\text{PO}_4 \cdot 6\text{H}_2\text{O}$) by *Aspergillus niger* (Suyamud et al. 2020), La and Ce oxalates resulted from the transformation of monazite sand, a naturally occurring lanthanide (Ln) phosphate mineral $[\text{Ln}_x(\text{PO}_4)_y]$ (Kang et al. 2020), while lead oxalate was formed through fungal bioweathering of pyromorphite ($\text{Pb}_5(\text{PO}_4)_3\text{Cl}$), a lead-containing phosphate mineral (Ceci et al. 2015; Fomina et al. 2004; Sayer et al. 1999). Other mechanisms of phosphate solubilization include acidification of the microenvironment by respiration-induced carbonic acid production (Burgstaller and Schinner 1993) and/or the production of other metal-complexing metabolites (Hoffland et al. 2004; Kucey et al. 1989).

Phosphate-solubilizing fungi are therefore able to promote the development of environmentally desirable metal-phosphate species and metal complexes which can reduce metal toxicity in polluted soils and improve plant yields by increasing P availability (Burgstaller and Schinner 1993; Fomina et al. 2004; Gadd 2017b; Goldstein et al. 1993; Mendes et al. 2015, 2017; Sullivan and Gadd 2019; Vassilev et al. 2013; Whitelaw 1999). Characterization of fungal interactions with a variety of P-containing minerals/metal compounds has been carried out (Fomina et al. 2004, 2005; Gadd 2017b; Jacobs et al. 2002; Liang and Gadd 2017; Mendes et al. 2014; Suyamud et al. 2020). Many studies have investigated the geochemical importance of fungi to nutrient mobilization, element cycling, and plant growth through bioweathering, including ectomycorrhizas (Hoffland et al. 2004; Landeweert et al. 2001), arbuscular mycorrhizas (Duponnois et al. 2005; Gosling et al. 2013), and rock-inhabiting microbial communities (Gleeson et al. 2010; Gorbushina 2007). Conversely, free-living saprotrophic fungi have received less attention concerning P mineral breakdown, despite the ability of many species to produce geoactive metabolites, such as citric and oxalic acid (Gadd 1999, 2017a; Gadd et al. 2014; Suyamud et al. 2020). Furthermore, studies on fungal RP solubilization have been predominantly conducted under liquid fermentation conditions, where phosphate is released from RP as a result of medium acidification by the tested fungi (Arcand and Schneider 2006; Illmer et al. 1995; Illmer and Schinner 1995; Mendes et al. 2014; Schneider et al. 2010; Vassilev et al. 2014). Another proposed strategy to take advantage of phosphate-solubilizing fungi is their application as soil inoculants (Alori et al. 2017; Sammauria et al. 2020; Vassilev et al. 2017; Vassilev and Mendes 2018). However, soil conditions are very different from liquid cultures, especially regarding soil pH buffering capacity (Nelson and Su 2010), and, therefore, fungal RP solubilization in the soil would require additional mechanisms. Although some solid-state fermentation studies on fungal RP solubilization have been carried out (Mendes et al. 2013, 2015; Vassileva et al. 2010; Vassilev and Mendes 2018), direct interactions between fungal hyphae and RP particles or the physical and biochemical bioweathering mechanisms involved have not been investigated. Therefore, the aim of this research was to investigate both chemical and physical bioweathering of RP, using a range of

free-living saprotrophic fungi with known geoactive properties, to clarify the mechanisms involved and their significance in mediating phosphate release and secondary mineral formation.

Materials and methods

Organisms and media

Aspergillus niger ATCC 1015, *Serpula himantioides*, and *Trametes versicolor*, from the Geomicrobiology Group culture collection, were maintained on malt extract agar (MEA, lab M, Bury, UK) at 25 °C in the dark. All test fungi are known to produce organic acids and secondary minerals and all have the potential to solubilize various minerals or insoluble metal compounds (Adeyemi and Gadd 2005; Ferrier et al. 2019; Fomina and Gadd 2008; Gadd 2016; Gharieb et al. 1998).

Rock phosphate

The rock phosphate used (Gardener's Direct, Hertford, UK) contained 13% P with an aqueous suspension having a pH of 6.6. The RP was milled in a 25 mm ceramic cylinder with 20 mm diameter ceramic balls at a frequency of 30 beats/s for 1 min in an MM2000 ball mill (Glen Creston Ltd., London, UK) and sieved to obtain particles of size 151–355 µm. Individual RP particles with visible morphological differences under light microscopy were analyzed using energy dispersive X-ray analysis (EDXA) to semi-quantitatively study the variation in P content. This analysis showed that the RP contained an average of ~6% P and a maximum percentage of ~11% P in a single particle (data not shown).

Fungal interaction with RP particles

MEA was prepared according to manufacturers' instructions (Lab M, Bury, UK) and amended with RP at 0.5% (w/v), which corresponds to the dose of the P source usually adopted in microbial phosphate solubilization studies (Nautiyal 1999). The RP and MEA were sterilized separately by autoclaving at 121 °C for 15 min, before mixing once MEA had cooled to approximately 50 °C. Control plates were not inoculated while test plates were inoculated with 4 mm diameter plugs of fungal mycelium cut from the edge of a 7-day old actively growing colony using a sterile cork-borer. All plates were incubated at 25 °C in the dark. Colony diameters on control and test plates were measured by taking the means of the shortest and longest diameters of the colony at specific time periods, usually twice daily. Changes in colony diameter with time were used to calculate the average growth rate. To examine direct interactions between *A. niger* and RP particles, MEA plates were set up with RP particles (151–355 µm) placed on the agar surface. Plates were incubated for 3 days at 25 °C in the dark in the presence or absence of *A. niger*. Full plate images were taken using a Sony Cybershot DSC-H20 from a 10 cm distance.

All experiments were carried out in triplicate and data subjected to analysis of variance ($p < 0.05$) using SigmaPlot 12.5. Normality and homoscedasticity of data were checked using Shapiro–Wilk and Brown–Forsythe tests, respectively.

Abiotic phosphate solubilization

To determine the significance of organic acids in mineral transformation under solid media conditions, abiotic experiments were carried out in 9-cm diameter Petri dishes containing MEA amended with 0.5% (w/v) RP. Plates containing 0.5% (w/v) of β -tricalcium phosphate ($\text{Ca}_3(\text{PO}_4)_2$, >98%, Sigma-Aldrich, Darmstadt, Germany) were used as a positive control since $\text{Ca}_3(\text{PO}_4)_2$ is an effective test for phosphate solubilization (Nautiyal 1999). Using a sterile cork borer, 4 mm diameter cylinders of agar were cut and removed from the agar, thereby creating a well. 20 μl aliquots of Milli-Q water (control) or a 0.5 mol/l solution of either citric, oxalic, or gluconic acid (all purchased from Sigma-Aldrich, Darmstadt, Germany) were added to the wells. These organic acids are significant metabolites of *A. niger* (Gadd 1999) and have previously shown to be involved in phosphate solubilization (Mendes et al. 2014). Plates were incubated overnight at 25 °C prior to measuring the solubilization halo diameters around the wells. Each plate had 3 wells, with triplicate plates being measured. Data were subjected to analysis of variance ($p < 0.05$).

Extraction of RP particles from solid media

RP particles embedded in MEA medium were extracted after incubation by melting the agar in Milli-Q water in a crystallizing dish. Samples were gently heated to around 80 °C to melt the agar and the liquid containing dissolved medium and mycelium were then siphoned off and replaced with fresh Milli-Q water. This procedure was repeated three times for each sample. Finally, extracted minerals and secondary myogenic minerals were transferred into Eppendorf tubes, centrifuged at 1150 g for 1 min (Centrifuge 5242, Eppendorf, Hamburg, Germany), and air-dried in a desiccator at ambient temperature for ≥ 48 h before analysis.

Scanning electron microscopy (SEM)

For SEM, air-dried RP samples were mounted on adhesive carbon tape on standard aluminum electron microscopy stubs. For energy dispersive X-ray analysis (EDXA), samples remained uncoated and were viewed and analyzed in the scanning electron microscope (XL30, Philips, Eindhoven, The Netherlands) using the Phoenix microanalysis system, operating at an accelerating voltage of 20.0 kV. To obtain SEM images, samples were coated with 5 nm gold and palladium combination (Au/Pd) using a rotating sputter device (208HR, Cressington Scientific Instruments Ltd, Watford, UK). Samples were then viewed using an accelerating voltage of 15.0 kV.

Cryogenic SEM was used to examine direct interactions between *A. niger* and RP particles. MEA medium blocks

($\sim 6 \times 4$ mm) containing single RP particles (colonized by the fungus or not) were cut using a sterile scalpel and mounted on electron microscopy gold stubs. Samples were frozen in liquid nitrogen slush within the freezing chamber of a cryopreparation system (Alto 2500, Oxford instruments, Abingdon, UK). Samples were then transferred under vacuum to the cryopreparation stage attached to the electron microscope and fractured to reveal the internal structure. The vacuum chamber was warmed to -95 °C and held at this temperature for 5 min to remove surface water from the samples, and after sublimation, the chamber was re-cooled to -115 °C before coating with 30 nm Au/Pd inside the microscope. The electron beam voltage was 15 kV.

X-ray diffraction (XRD) and X-ray powder diffraction (XRPD)

For X-ray diffraction (XRD) analysis, mineral samples were individually mounted onto a steel sample holder with an Al_2O_3 fixative, and an X-ray beam was fired at the sample using an X-ray diffractometer (Hilton-Brooks, Cheshire, UK) with a monochromatic Cu-K_α radiation source and curved graphite, single-crystal chronometer (30 mA, 40 kV). Diffraction intensities were taken over the range of 3 – 60° 2θ , at a scan rate of $1^\circ/\text{min}$ in 0.1° increments. For X-ray powder diffraction (XRPD), diffraction patterns were scanned and recorded from 3 to 60° 2θ using Ni-filtered Co-K_α radiation, scanning from 3 – 60° 2θ counting for 300 seconds per step on an X'Pert Pro Multipurpose Diffractometer (PANalytical, Cambridge, UK) using an X'Celerator detector (PANalytical, Cambridge, UK). Phases were identified with reference to patterns from the International Center for Diffraction Data (ICDD) Powder Diffraction File (PDF) database and the Inorganic Crystal Structure Database (ICSD).

Results

Interactions between fungi and RP particles

RP addition had no effect on fungal growth and average growth rates on media containing RP were 12, 11, and 17 mm/day for *A. niger*, *S. himantioides* and *T. versicolor*, respectively (Table 1). Under all treatments, RP particles incubated with test fungi became gradually surrounded by a white, cloudy precipitate (Figure 1). Most precipitation was observed closest to the site of fungal inoculation. Rock phosphate particles from within these cloudy zones of precipitation were extracted and examined using SEM (Figure 2) and XRPD (Figure 3). Scanning electron microscopy of the

Table 1. Growth rates (mm/day \pm standard error of mean) of test fungi on MEA plates with (+) or without (–) rock phosphate at 0.5% (w/v).

Fungus	Rock phosphate	
	+	–
<i>Aspergillus niger</i>	12 \pm 0.7	12 \pm 0.7
<i>Serpula himantioides</i>	11 \pm 1.3	10 \pm 0.9
<i>Trametes versicolor</i>	16 \pm 1.2	18 \pm 0.4

There were no differences in growth rate due to RP amendment ($p > 0.05$).

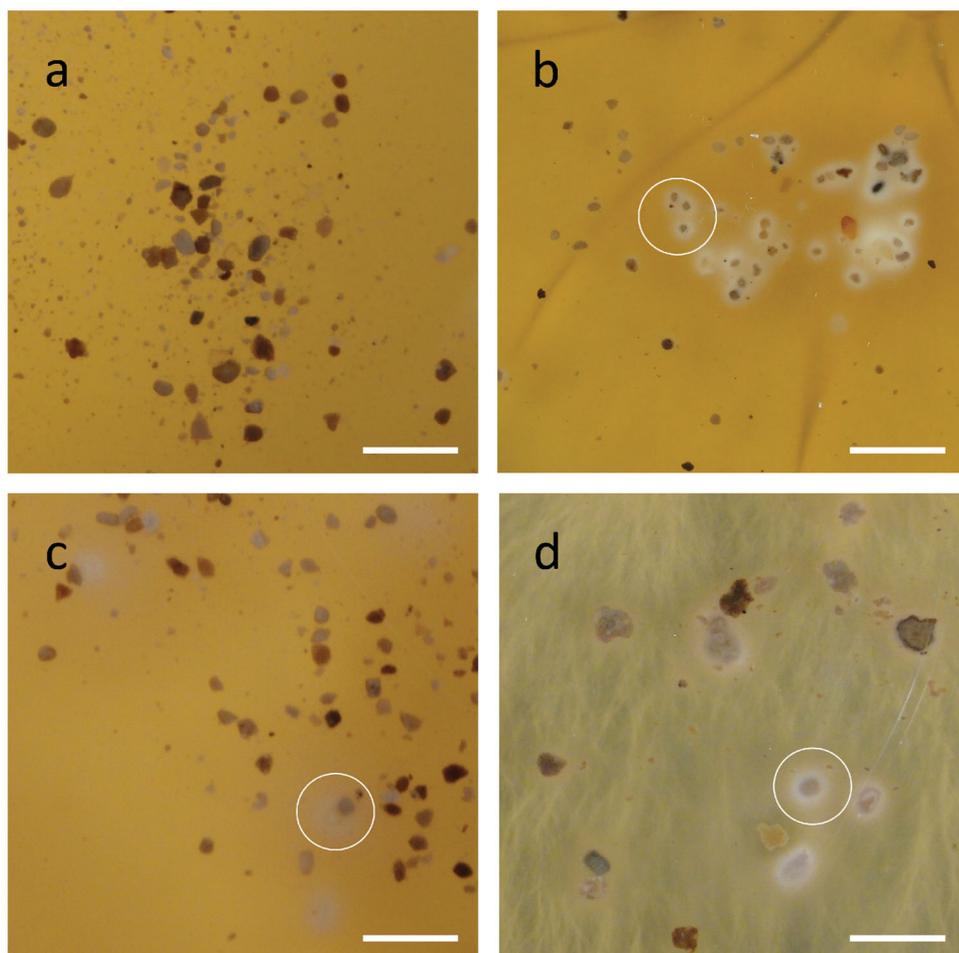


Figure 1. Fungal interaction with rock phosphate (RP) particles on MEA plates. Images show RP incubated with (a) no fungal inoculant, (b) *Aspergillus niger*, (c) *Trametes versicolor*, and (d) *Serpula himantoides* for 7 days at 25 °C in the dark, with examples of mineral precipitation or dissolution circled. Scale bars: 1 mm. Images shown are typical of several images.

samples revealed that the surface structure of the RP was altered by each of the test fungi in a distinct manner (Figure 2).

XRPD showed that control RP, not exposed to fungal metabolites, consisted mainly of apatite (Figure 3(a)) and traces of quartz (not shown). Rock phosphate incubated with test fungi revealed the additional presence of whewellite (calcium oxalate monohydrate, $\text{CaC}_2\text{O}_4 \cdot \text{H}_2\text{O}$). Overall, the intensity of peaks relating to apatite decreased with fungal incubation indicating the progressive conversion of apatite into whewellite. Since the XRPD patterns for all test fungi were similar, only data for *A. niger* are shown (Figure 3(b)). It is worth mentioning that there were no differences observed in RP particles that had or had not been subjected to the extraction method from solid media, meaning that any changes in mineral morphology were due to the test fungi and the metabolites they produce.

Role of organic acids in transformations of P-containing minerals

The formation of whewellite on the surface of RP particles provided strong evidence of the role of oxalic acid in the release of phosphate. To determine the significance of

individual organic acids on mineral solubilization, abiotic tests with organic acids commonly associated with fungal P-mineral solubilization were conducted in agar plates containing 0.5% (w/v) $\text{Ca}_3(\text{PO}_4)_2$ or RP. Figure 4(a–d) shows the effect of organic acids on $\text{Ca}_3(\text{PO}_4)_2$ solubilization, with sterile Milli-Q water serving as a control. The halo zone surrounding the wells filled with oxalic acid was the most pronounced (~9 mm diameter), followed by citric acid (~5 mm diameter), with no halo zones surrounding the gluconic acid and control wells. In the plates containing RP, oxalic acid again produced the most distinct changes in mineral/plate appearance (Figure 4(e–h)) and was the only organic acid that resulted in any noticeable changes. The light-colored precipitates observed in these experiments resembled those seen around RP particles incubated with various fungi in earlier experiments (Figure 1), suggesting that oxalic acid is mainly responsible for the reaction seen.

When RP particles were extracted from the agar surrounding the wells and examined using SEM, it was clear that the white precipitates observed when RP was exposed to oxalic acid corresponded to changes in surface morphology and structure of the RP particles (Figure 5). Citric and gluconic acid created no detectable changes in the surface appearance of RP.

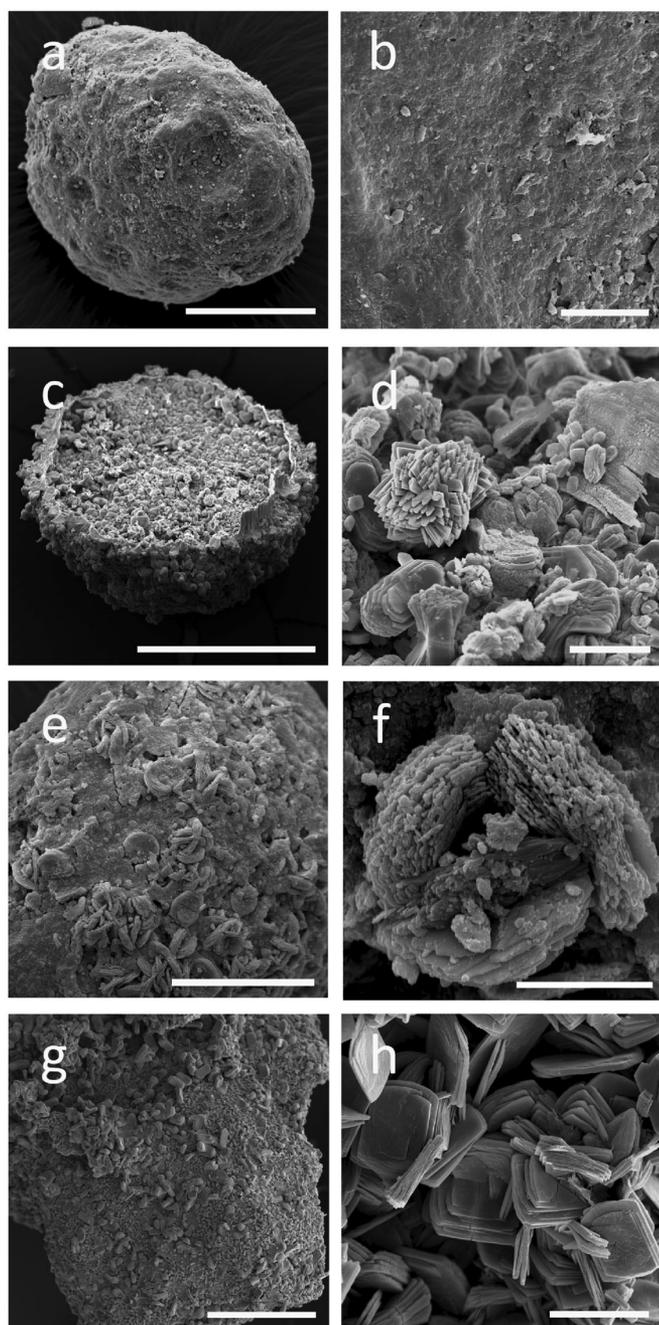


Figure 2. Alterations of rock phosphate (RP) surface structure caused by fungi. SEM images of RP incubated with (a, b) no fungi, (c, d) *Aspergillus niger*, (e, f) *Trametes versicolor*, (g, h) *Serpula himantioides* on MEA until fungal colonies had reached the edge of the Petri dishes. Higher magnification images in the right panel all relate to the respective panel on the left. Scale bars: (a) 100 μm , (b, d, h) 20 μm , (c, g) 200 μm , (e) 50 μm , (f) 5 μm . Images shown are typical of several images.

Rock phosphate particles extracted from these plates were analyzed for mineral composition using XRPD (Figure 3(c–e)). Significant mineralogical changes were only detected with oxalic acid, which again showed the presence of whewellite, also found when RP was incubated with test fungi (Figure 3(b)).

Physical mechanisms of RP bioweathering by *A. niger*

The effect of fungal metabolites, particularly oxalic acid, has therefore been demonstrated to play a key role in

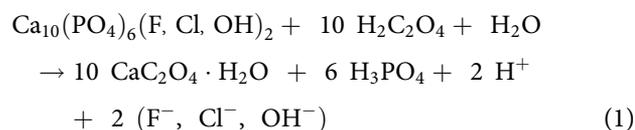
biochemical alteration of RP. However, the physical effects of fungal colonization cannot be overlooked. Therefore, RP particles were incubated in the presence or absence of *A. niger*, with no barrier between the particles and the fungus, thereby allowing direct colonization of the RP. *Aspergillus niger* was chosen for this aspect because it produced pronounced chemical changes in the RP particles and has a well-known ability to solubilize mineral phosphates (Mendes et al. 2014; Whitelaw 1999).

Extensive RP colonization by *A. niger* was clearly visible on the outer and internal surfaces of the particles (Figure 6(b,d), respectively). Furthermore, hyphae were observed to show physical penetration and burrowing through the particles (Figure 6(d,f,h)), and possibly thigmotropism, that is, responding to cracks and fissures within the RP particle structure (Figure 6(d,f)).

Discussion

This research has demonstrated direct fungal interactions with RP particles, highlighting the chemical and physical transformations mediated by the fungus during RP bioweathering. Our data have demonstrated that oxalic acid had a key role in the chemical transformation of apatite in RP, resulting in the formation of whewellite as a secondary biomineral and, hence, releasing soluble phosphate. Moreover, we present evidence of physical mechanisms of fungal bioweathering, including penetration, burrowing, and tunneling through the RP particles.

Whewellite was present in a variety of crystalline forms after incubation of RP with *A. niger*, *S. himantioides*, and *T. versicolor*. It has been found that RP solubilization using oxalic acid in an aqueous medium produces calcium oxalate minerals, with a predominance of whewellite over weddellite (Mendes et al., unpublished data). Whewellite ($\text{CaC}_2\text{O}_4 \cdot \text{H}_2\text{O}$) is hydrated calcium oxalate, and as a monohydrate, it is more stable than its dihydrate form (weddellite) and one of the most common forms of oxalate encountered in the environment (Gadd 1999; Gadd et al. 2014). In terrestrial environments, calcium oxalate crystals are commonly found associated with fungi, and many species are capable of their formation due to secretion of oxalic acid: both weddellite and whewellite have been recorded (Fomina et al. 2010; Gadd 1999, 2017b; Gadd et al. 2014; Pylro et al. 2013). In our experiments, the source of the calcium was primarily from the apatite component of the RP, confirming that the whewellite product results from the interaction between the fungus and the apatite, following the equation:



The phosphate from the apatite is released into the external medium and can either remain free in the medium, be taken up by the fungi or other organisms (bacteria, plants) in the vicinity, or reprecipitate as a secondary phosphate

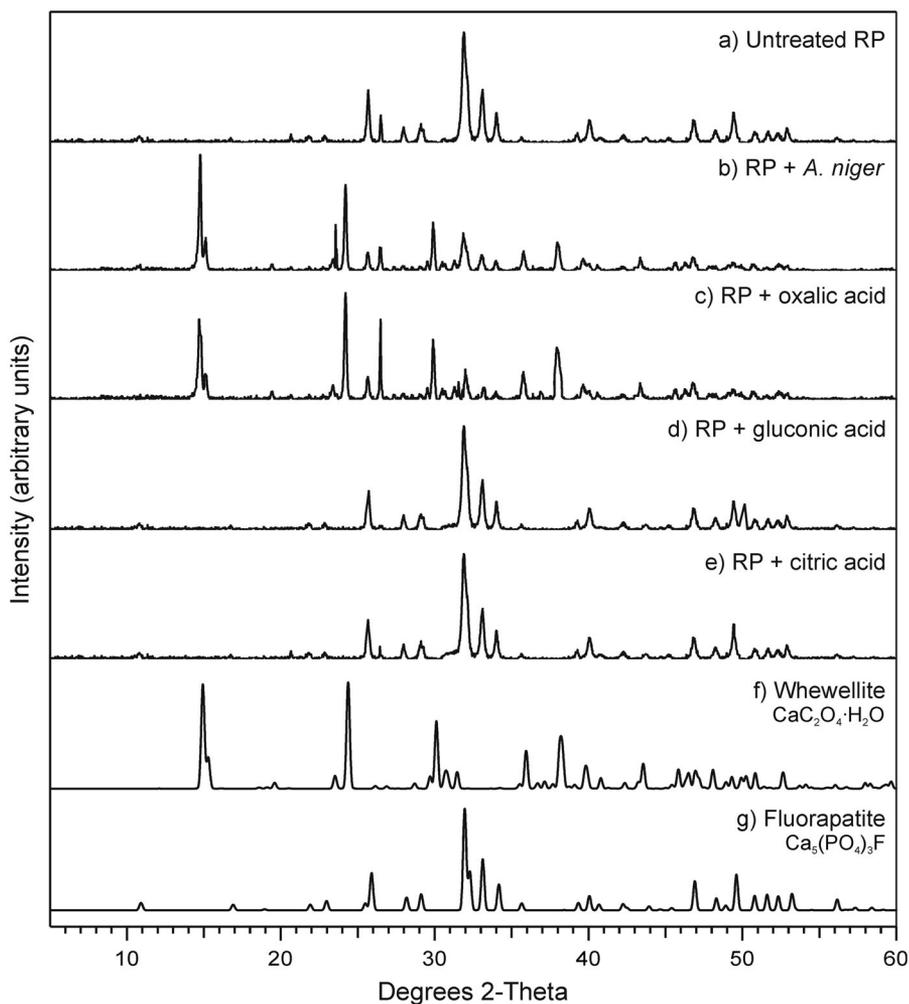


Figure 3. X-ray diffraction patterns of (a) untreated rock phosphate (RP) and RP incubated with (b) *Aspergillus niger* or with (c) oxalic, (d) gluconic or (e) citric acid at 0.5 mol/l. Standard patterns of (f) whewellite (ICSD 434201) and (g) fluorapatite (ICSD 9444) are also shown. All replicate samples from within treatments were pooled, and a subsample analyzed. Note the formation of whewellite when RP was incubated with *A. niger* and oxalic acid. Typical patterns are shown from one of several analyses.

mineral. Thus, it is clear that the excretion of oxalic acid was significant in affecting the solubilization of RP mineral components. This is also supported by previous research where oxalic acid was found to be the most effective organic acid for solubilizing RP in aqueous media (Kpombekou and Tabatabai 1994, 2003; Mendes et al. 2014, 2020).

The role of oxalic acid in RP transformations after fungal inoculation was also confirmed in abiotic tests with the most common organic acids associated with fungal phosphate solubilization, namely oxalic, citric, and gluconic acid (Arcand and Schneider 2006; Mendes et al. 2014; Richardson et al. 2011; Schneider et al. 2010; Silva et al. 2014). Oxalic acid was the most effective organic acid at solubilizing Ca₃(PO₄)₂ and RP, producing the widest halo zone on Ca₃(PO₄)₂-containing media and the greatest extent of noticeable precipitation on the RP which was similar to that observed on RP when directly exposed to fungal exudates. Images of the RP particles taken from the area surrounding the wells revealed similar morphologies between RP particles treated abiotically with oxalic acid or with test fungi. In fact, oxalic acid was found to be the only organic acid that caused a change in the surface morphology and

mineral composition of RP, resulting in whewellite production. Citric acid showed a much lower potential than oxalic acid for solubilizing the P sources. This is corroborated by other published works that report a lower RP solubilization efficiency of citric acid because oxalate can form a more stable complex with calcium (Kpombekou and Tabatabai 1994, 2003; Mendes et al., 2020). There were no obvious effects of gluconic acid, which implied that gluconate was not involved in direct solubilization of RP under these conditions. However, gluconic acid is commonly detected in assays of bacterial and fungal solubilization of phosphates (Illmer and Schinner 1995; Mendes et al. 2013, 2014, 2014; Schneider et al. 2010; Whitelaw et al. 1999), and it is possible that gluconic acid plays a minor role in the solubilization of phosphate-containing minerals, for example, by reducing the pH. Furthermore, in the natural environment, each organic acid may not be produced exclusively, and they may work in synergy or compete with each other (Carlile et al. 2001).

Chemically-induced mineral phosphate solubilization has been widely studied and is considered to be the main mechanism of microbial phosphate release from sparingly soluble

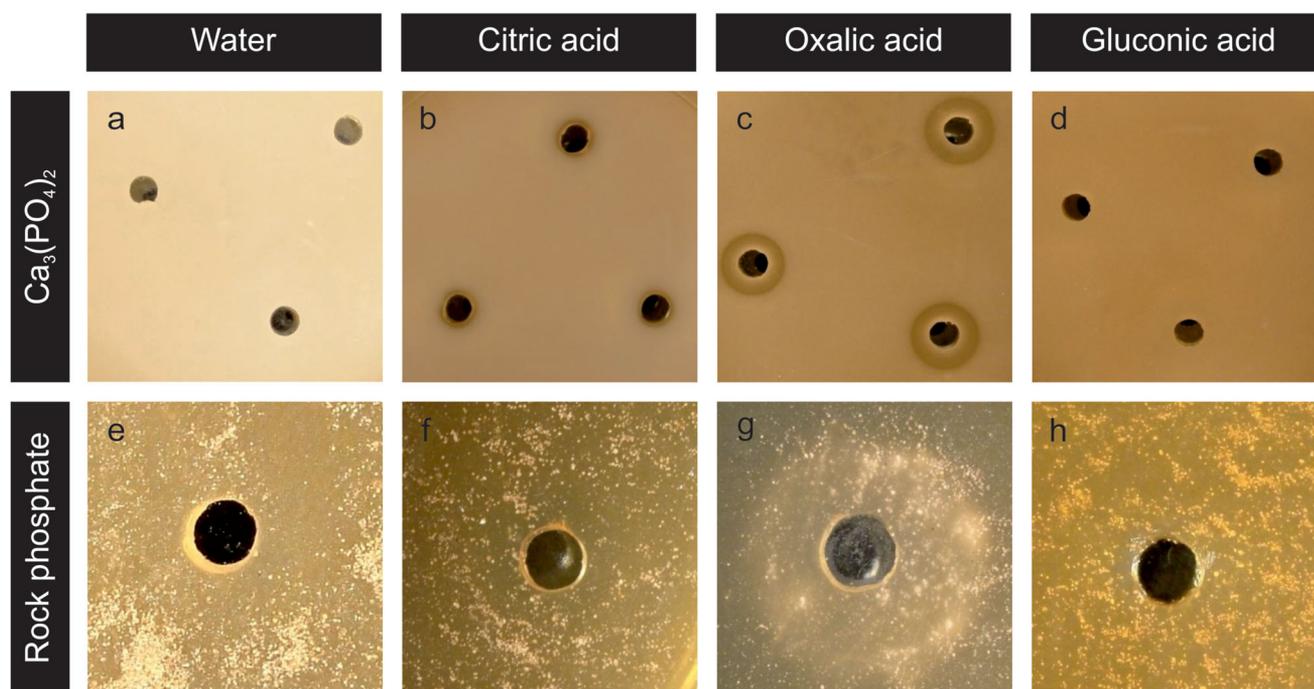


Figure 4. Solubilization halos from (a–d) $\text{Ca}_3(\text{PO}_4)_2$ or (e–h) rock phosphate by addition of individual organic acids into wells bored in MEA plates amended with 0.5% P mineral. Agar wells were filled with 20 μl (a, e) ultrapure water, (b, f) citric, (c, g) oxalic or (d, h) gluconic acid. All organic acid concentrations were 0.5 mol/l. Images shown are typical examples from several determinations.

sources. This can be employed in bioprocesses based on microbially-produced organic acids as means to obtain soluble phosphate from RP (Goldstein et al. 1993; Mendes et al. 2013, 2015, 2017; Vassilev and Vassileva 2003), wastes, such as chars and ashes (Atienza-Martínez et al. 2014; Vassilev et al. 2009, 2013, 2013; Vassileva et al. 2012) and other insoluble phosphate minerals such as struvite (Suyamud et al. 2020). On the other hand, physical mechanisms of fungal phosphate bioweathering have been largely neglected, despite their potential importance in natural environments and in agroecosystems that rely on RP. In this regard, our results demonstrate active mineral colonization by fungi, including burrowing, tunneling and associated fragmentation and transformation, suggesting that physical interactions have an important role in RP bioweathering.

The physical force exerted by growing fungal hyphae has been well explored, but mainly focused on topics where fungi are pathogenic (Brand 2012; Money 2004; Money and Howard 1996), or where they cause damage to human-made structures (Gadd 2017c; Gadd et al. 2014; Sterflinger 2010). However, physical mechanisms of mineral biodeterioration have been demonstrated to be important in black slate bioweathering, along with biochemical mechanisms, by the fungus *Schizophyllum commune* (Kirtzel et al. 2020). Direct mineral interactions were also found to be necessary for more efficient bioweathering of cobalt-containing laterites and pyritic ores (Yang et al. 2019) and manganese oxides (Ferrier et al. 2019). Examples of fungal tunneling (Fomina et al. 2010; Hoffland et al. 2003; Jongmans et al. 1997; Pinzari et al. 2013) and ‘footprints’ (the marks left by fungal interactions) (Jongmans et al. 1997; Kirtzel et al. 2020; Pinzari et al. 2013; Wei et al. 2012) have been widely

demonstrated as components of bioweathering, though importantly, not necessarily isolated from organic acids and their impact. We have evidence for similar mechanisms of fungal penetration of RP particles. This confirms that *A. niger* has a turgor pressure suitable for probing RP or other minerals, and that physical interactions are also important in the biodeterioration of RP. In the silicate mineral feldspar, fungal tunneling contributed to approximately 2% of the total weathering (Smits et al. 2005). Interestingly, the frequency of tunneling in mineral samples was found to increase with nutrient limitation (Hoffland et al. 2004), so it can be hypothesized that fungal tunneling of RP would also be more evident at limiting P, allowing for a more significant contribution of physical interactions to bioweathering. Indeed, it was demonstrated that mesh bags filled with apatite stimulated mycelial growth of mycorrhizal fungi around them, with a higher stimulus in P-deficient soil than in soil with adequate P levels (Hagerberg et al. 2003). Such mechanisms of phosphate solubilization in a free-living saprotrophic fungus such as *A. niger* supports the possibility of applying phosphate-solubilizing fungi as a soil inoculant in agriculture (Vassilev et al. 2019; Vassilev and Mendes 2018), especially in tropical areas where P-deficient soils predominate (Roy et al. 2016) and the use of RP as fertilizer is widespread. In the soil, where the production and diffusion of organic acids may be limited, direct hyphal interactions with RP particles are very likely to be an important contributory mechanism for phosphate solubilization.

In summary, this research reveals chemically- and physically-induced fungal transformation of RP particles, highlighting the important role of fungi in P cycling, as well as other associated elements (Gadd et al. 2014). Oxalic acid was

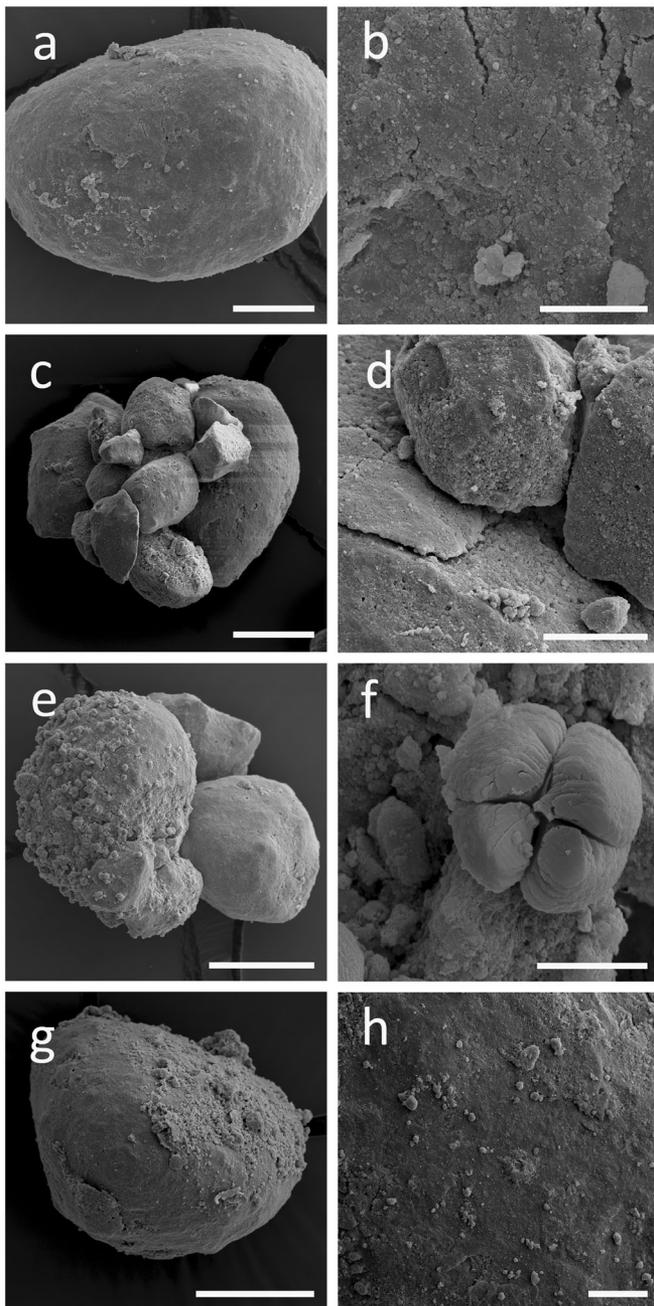


Figure 5. Alterations of RP surface structure when treated with organic acids in abiotic agar plates. SEM images showing RP exposed to (a, b) ultrapure water, (c, d) citric, (e, f) oxalic, or (g, h) gluconic acid. Scale bars: (a) 50 μm , (c, e, g) 100 μm , (b) 10 μm , (d, h) 25 μm , (f) 5 μm . Images shown are typical of several images.

shown to be a key factor in this process. Beyond participating in P mineral bioweathering in natural environments, these fungi present a potential to enhance phosphate uptake by plants in agronomic systems. In contrast to other studies using liquid media, our data demonstrate that *A. niger* can interact and transform RP in a solid matrix, which supports the idea of using fungal inoculants combined with RP to promote phosphate release in agricultural soil. The mechanisms exhibited by free-living saprophytic fungi may be critical in releasing phosphate to other organisms in colonizing ecosystems influenced by the geological, geomorphic, and agricultural change.

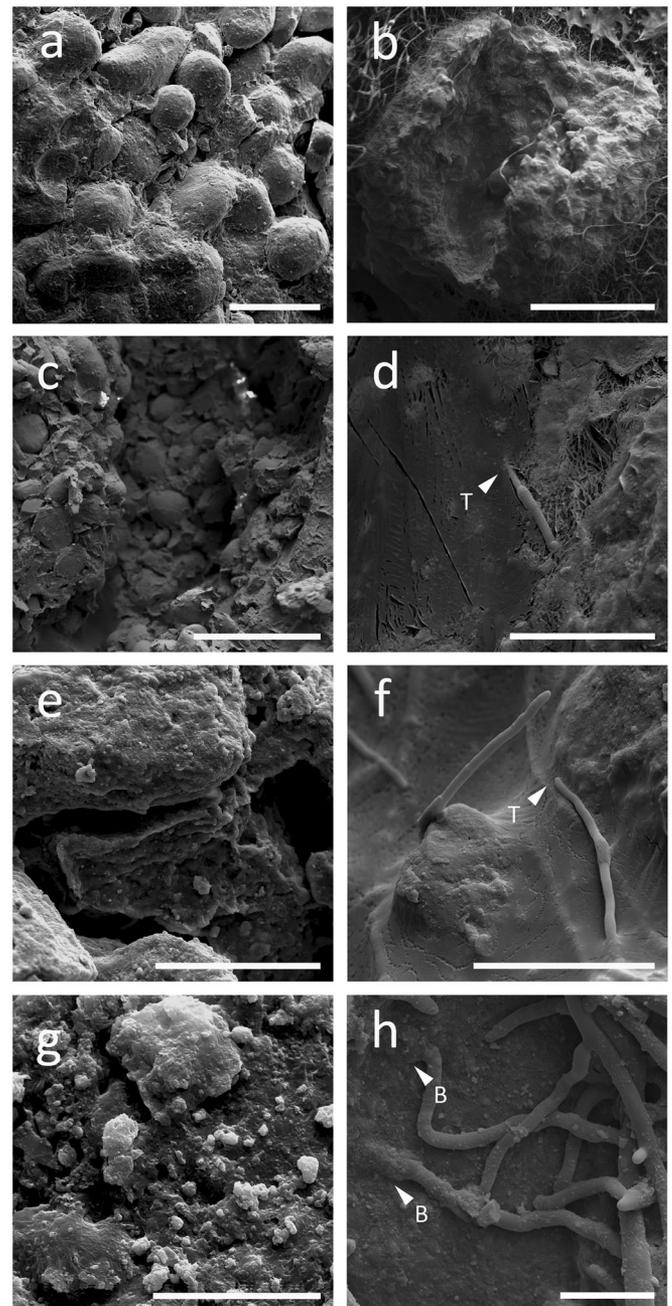


Figure 6. Colonization of RP particles by *Aspergillus niger*. Left (a, c, e, g) and right (b, d, f, h) panels show cryogenic scanning electron micrographs of non-inoculated and inoculated RP particles, respectively. Note the complex aggregate structure (a, c) and the porosity (e, g) of RP particles. The images on the right show a RP particle with the middle section removed, allowing the interior of the particle to be studied. Note the ability of *A. niger* to effectively colonize RP particles (b), as well as thigmotropically (T) responding to cracks and fissures within the structure (d, f), physically burrowing (B) through RP particles (h) and depositing minerals or forming mineral nucleation sites on their hyphal surfaces (h). Scale bars: (a) 200 μm , (b) 1 mm, (c) 500 μm , (d–f) 50 μm , (g) 10 μm , and (h) 20 μm .

Conclusions

Fungi can cause chemical and physical alterations to rock phosphate particles, using mechanisms that actively participate in environmental P cycling. Oxalic acid has a key role in such transformation of RP and related phosphate minerals. Furthermore, the results demonstrate that the

production of oxalic acid is responsible for the formation of the secondary biomineral whewellite as a result of apatite solubilization. Moreover, our data provide clear evidence of fungal penetration and tunneling through RP particles, showing that physical interactions are also important for RP solubilization and bioweathering.

Disclosure statement

No potential conflict of interest was reported by the author(s).

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ORCID

Gilberto de Oliveira Mendes  <http://orcid.org/0000-0003-3900-719X>
 Stephen Hillier  <http://orcid.org/0000-0001-9652-9950>
 Timothy S. George  <http://orcid.org/0000-0003-3231-2159>
 Geoffrey Michael Gadd  <http://orcid.org/0000-0001-6874-870X>

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