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1 **Potential side effects of biocontrol and plant-growth promoting *Bacillus amyloliquefaciens* bacteria**  
2 **on earthworms**

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8 **Abstract**

9 Many bacteria strains are now successfully used for plant-growth promotion (PGPR) and as  
10 biocontrol agents (BCA) against plant diseases. Mechanisms behind their action involve production of  
11 enzymes and antibiotics, which in high concentrations could also affect non-target organisms hence  
12 the biodiversity and processes in the soil. Despite these potential negative side effects, there is little  
13 research done on the subject to confirm whether they are significant. In three laboratory  
14 experiments, we tested the effect of the bacterial BCA *Bacillus amyloliquefaciens* UCMB5113 (BA) on  
15 two earthworm species, common in agricultural soils in temperate regions of the world and  
16 representing different ecological groups; one anecic (*Aporrectodea longa*) and one endogeic species  
17 (*A. caliginosa*). The earthworms were kept in replicated pots containing soil from local agricultural  
18 fields. They were fed on cow manure, and exposed to BA by 1) dipping into a BA solution (short-term  
19 external exposure in high concentration), 2) mixing BA solution into the soil (long term external and  
20 internal exposure) and 3) feeding earthworms with BA infested plant litter (internal exposure of the  
21 gut).

22 After 1-2 months, survival, growth and reproduction of the earthworms were recorded. We found no  
23 effect of the treatments as compared to control without BA amendments. We conclude that the use  
24 of high doses of BA with concentrations at the same magnitude as maximally expected when the

25 bacteria are used as PGPR and BCA, is not harmful to the soil dwelling earthworms tested in this  
26 project. Further studies of the ecological effects of PGPR and BCA bacteria on other non-target soil  
27 organisms are encouraged. The development of sustainable agricultural systems, where ecosystem  
28 services are optimized, has to be aided by a deeper knowledge of the combined effect of bacteria  
29 and earthworms on the promotion of plant health.

30

31

32 Key words

33 *Aporrectodea longa*; *Aporrectodea caliginosa*; growth; reproduction; toxicity test; biocontrol agents

34

35 Highlights

36 First study of PGPR and BCA bacteria's influences on earthworms

37 No harmful effects of the biocontrol bacteria on earthworms were found

38 BCA bacteria-earthworm interactions are interesting for development of sustainable agriculture

39

40

## 41 **1. Introduction**

42 In recent years, scientific attention has been drawn to the effects of rhizobacteria as beneficial to  
43 plants: plant-growth promoting rhizobacteria (PGPR), enhancing plant tolerance against abiotic  
44 stress, and biological control agents (BCA) against plant diseases and insect pests (Dimkpa et al.,  
45 2009; Lugtenberg and Kamilova, 2009; Pieterse et al., 2014). Several bacteria, including strains of the  
46 genera *Pseudomonas* and *Bacillus*, are now available commercially as BCAs and are successfully used

47 instead of chemical pesticides in crop production (Choudhary and Johri, 2009). PGPRs can stimulate  
48 plant growth in different ways, e.g. enhance seed germination and emergence, stimulate root  
49 development and thus mineral, nutrient and water uptake, as well as suppress diseases. The  
50 underlying mechanisms of beneficial rhizobacteria for protection of plants against parasitic root  
51 colonizing microorganisms include priming of induced systemic resistance and production of  
52 enzymes such as chitinases, peroxidases and proteases, and many types of antibiotics (Pieterse et al.,  
53 2014). This production does not only affect microorganisms and their interactions with plants but is  
54 also known to suppress nematodes and techniques for use of bacterial BCA against plant parasitic  
55 nematodes are being developed (Abally 2012; Mutua et al., 2011; Niazi et al., 2014; Wepuhkhulu et  
56 al., 2011).

57 It is suspected that the use of bacterial BCAs would also affect many other non-target soil organisms  
58 and therefore influence soil processes and biodiversity. This has so far not received much attention.  
59 For example, earthworms, like nematodes, have chitin in their cuticle, especially in their setae  
60 (Jamieson, 1992; Miller and Harley, 1999), and therefore could be negatively affected by addition of  
61 microorganisms producing chitinase. Although biocontrol bacteria occur naturally in soil, amending  
62 them in large concentrations to soils and plants could imply environmental risks. Therefore, thorough  
63 assessment of environmental impacts of BCAs needs to be carried out prior to their development and  
64 registration for use in plant production to avoid ecotoxicological effects at different trophic levels in  
65 the local ecosystem.

66 Many *Bacillus* species are ubiquitously present in soil and can become enriched in the rhizosphere  
67 depending on root exudates. Phenotypically high ecological diversity has been found among different  
68 *Bacillus* species with plant interaction resulting both in epiphytic and endophytic colonization (Mc  
69 Spadden Gardener, 2004). Many strains of *Bacillus subtilis*, *Bacillus cereus* and *Bacillus*  
70 *amyloliuefaciens* have been found to interact with plants and produce beneficial effects including  
71 disease suppression (Choudhary and Johri, 2009). The type strain of plant-associated *B.*

72 *amyloliquefaciens* FZB42 has been shown to produce a variety of secondary metabolites involved in  
73 microbial antagonism and thus supporting disease suppression of plants (Chen et al., 2009), and this  
74 also includes chitinase (Niazi et al., 2014).

75 In the present study we have tested the effect of the bacterial BCA *Bacillus amyloliquefaciens*  
76 UCMB5113 (Here after abbreviated as BA) on the survival, growth and reproduction of two  
77 earthworm species that are common in agricultural soils in temperate regions of the world and  
78 represent two different ecological groups (Bouché, 1977). Although the BA bacteria are not yet  
79 available as a commercial BCA, substantial research has been done on its effect on plant growth and  
80 health as well as the underlying mechanisms of action (Danielsson et al., 2007; Sarosh et al., 2009)  
81 and genomic and phenotypic analysis infer a close relationship with the type strain FZB42 (Niazi et  
82 al., 2014).

83 The aim of the study was to ascertain whether *B. amyloliquefaciens* UCMB 5113 (BA) has any effect  
84 on earthworms when exposed directly to a solution of the bacteria, or to soil or feed inoculated with  
85 the bacteria.

## 86 2. Material and Methods

### 87 2.1 Test organisms

88 The tested earthworm species were *Aporrectodea longa* (Ude) and *Aporrectodea caliginosa*  
89 (Savigny). The former belongs to the ecological category of anecic earthworms. It generally feeds on  
90 plant litter on the surface, buries litter into the soil and creates burrows from the surface down  
91 through the soil profile. The latter is an endogeic species that lives and feeds in the soil profile where  
92 it consumes large quantities of soil and organic matter but are not so selective towards fresh litter.  
93 The earthworms used were collected from agricultural and garden soils in the vicinity of Uppsala by  
94 digging and hand sorting. Prior to their use in the experiments, the earthworms were maintained in a  
95 climate chamber at 18 °C for up to two months, in 6-litre boxes with soil of the same quality as used

96 in the experiments (see description of soil below), and were fed with rehydrated dry cow dung added  
97 once a month and mixed into the superficial layer of the soil. We used new earthworms for each  
98 experiment. They were adults with fully developed clitellum or subadults with early signs of clitellum  
99 development and all chosen specimens were in full vigour.

100 *Bacillus amyloliquefaciens* subsp. *plantarum* UCMB5113 (Borriss et al. 2011) (BA) was grown in LB  
101 medium at 28 °C with agitation until stationary phase was reached. The suspension was heat  
102 shocked for 5 min at 65 °C and surviving spores collected by centrifugation. After washing the pellet  
103 in sterile MilliQ water, the density was determined using colony forming unit counts and the  
104 concentration adjusted with sterile water to 10<sup>7</sup> ml<sup>-1</sup>.

## 105 2.2 Experimental set up

106 The study was conducted in laboratories, based at the Swedish University of Agricultural Sciences  
107 (SLU), Uppsala (59°49'05''N, 17°39'28''E). In mesocosm experiments, we exposed earthworms to BA  
108 by 1) dipping into a bacteria solution (short term external exposure in high concentration), 2) mixing  
109 the bacteria into the soil where the earthworms were kept (long term external and internal  
110 exposure) and 3) feeding earthworms with bacteria infested plant litter (internal exposure of the  
111 gut).

112 Three different experiments were done with various combinations of exposition methods and  
113 earthworm species, summarized in table 1. The experiments were preceded by preliminary studies  
114 where soil mixture, moisture level and feeding were tested. Water content appeared to be the most  
115 critical since the soil became hard and impenetrable for the earthworms if allowed to dry out. The  
116 vessels used in experiment 1 and 2, were cylinders made from PVC plastic sewage pipes with 14.5 cm  
117 inner diameter and 30 cm height. At the bottom of the cylinders, nylon mesh (mesh size 1 mm) was  
118 attached with a rubber band to allow good drainage of the soil and prevent earthworms from  
119 escaping. The walls of the cylinders extended ca 15 cm above the level of the soil surface, to prevent  
120 earthworms from escaping. The top of the cylinders were loosely covered with transparent

121 polyethylene plastic bags in order to minimize evaporation. For experiment 3, opaque plastic boxes (27  
122 cm x 17 cm wide x 13 cm deep) were used. They were perforated in the bottom to allow drainage  
123 and the internal base of the vessels was covered with nylon net to prevent escape of earthworms.  
124 The boxes had no lid and were covered with a net and a nylon sheet in order to avoid excessive  
125 evaporation and infection of *Sciaridae* flies (See table 1). The boxes provided a greater soil volume  
126 than the cylinders allowing for a higher number of earthworms and less laborious handling.

127 The vessels were filled with a moist soil mixture (15% water) consisting of 60 % clay-loam soil and 30  
128 % sandy soil and 10 % cow manure. The clay-loam soil contained 36.5 % clay, total carbon content  
129 was 1.5 %, pH 6.6, and was classified as Eutric cambisol (Kirchmann et al. 1994). The sandy soil  
130 contained 2.7 % carbon and pH was 6.3. Both soils were collected from the experimental farm area  
131 of the SLU University in the vicinity of Uppsala. The soils were hand sorted to remove roots, debris,  
132 stones and macrofauna (e.g. earthworms and beetles) and thereafter frozen (48 h, -20 °C) and  
133 thawed (48 h, +20 °C) twice to reduce the remaining indigenous fauna. This would be efficient for  
134 reduction of macro- and mesofauna but not for nematodes and other microfauna (Sulkava and  
135 Huhta, 2003). Dried cow manure (Weibulls® concentrated, dried organic cow manure) was wetted to  
136 50 % moisture content before being mixed into the experimental soil as feed for the earthworms.  
137 The particle size of the manure was on average less than 1 mm with no particles larger than 3 mm. In  
138 experiment 1, an additional amount of 100 g of wetted cow manure was added per cylinder at day 29  
139 of the experiment as feed for the worms. The manure was evenly mixed into the soil in all  
140 experiments and also when additional manure was added in experiment 1. The water content of the  
141 mineral soil was ca. 15 % by wet weight at the start of the experiment and the soil mixture was  
142 wetted to field capacity before introducing the earthworms.

143 The procedure for the three exposure methods was as follows: In the dipping method (treatments DS  
144 and D in experiments 1 and 3) earthworm specimens were dipped for 15 seconds into a BA spore  
145 solution in sterile water with  $1 \times 10^7$  cells ml<sup>-1</sup>. In the Control (C), worms were dipped into deionised

146 water for 15 sec before being added to soil-filled cylinders. In treatments with BA mixed into the soil  
147 (Experiments 1 and 2; treatments S, DS, SL, and SL+) 150 ml of BA spore solution in sterile water ( $1 \times$   
148  $10^7$  cells  $\text{ml}^{-1}$ ) was poured over the soil. To distribute it more evenly, we did not pour the whole  
149 solution on top of the soil. Instead, it was added in three portions; after filling 1/3, 2/3 and 3/3 of the  
150 whole soil volume. Amendment to leaves (treatment L+ in experiment 2) was done by keeping leaves  
151 in the BA solution for 1 min and then the excess liquid was shaken off gently to mimic spray  
152 administration of *Bacillus* with subsequent runoff. Leaves treated with water only, served as a  
153 control (treatment L). In a similar way to what we did with the BA solution to distribute it more  
154 evenly, we added 4 g of amended or control leaves on top of the first third of the total amount of soil  
155 mixture, then another third of soil was added to the cylinder and 4 g more of amended leaves, and so  
156 with the third portions of soil and leaves. An additional 4 g of leaves was added on the surface after  
157 one and two weeks in L+, L and SL+ treatments (see table 2).

158 Two earthworm specimens were added to each experimental cylinder and four *A. longa* or six *A.*  
159 *caliginosa* to each box. The experimental units were arranged in a randomized design and kept in  
160 darkness at 17-19 °C in a climate chamber for the duration of the experiments (see table 1). They  
161 were covered with transparent plastic bags in order to prevent excessive evaporation, and watered  
162 regularly. They were moved around every second week in order to minimize effects due to any local  
163 differences in temperature and evaporation rates. Each individual earthworm was weighed at the  
164 start and end of the experiments after being washed in cold tap water and dried on paper tissue. The  
165 individual fresh mass was also recorded at day 29 in experiment 1. At the end of the experiments, all  
166 cocoons produced were sorted out by wet sieving of the soil over a mesh (mesh size 2 mm) and  
167 counted.

168 The three experiments were arranged as indicated in tables 1 and 2. Experiment 1 included four  
169 treatments with *A longa* as follows: (1) DS: Dipping earthworms into BA solution+ mixing BA into the  
170 soil; (2) D: Dipping earthworms in BA solution+ no mixing; (3) S: No dipping + mixing BA into the soil;



171 (4) C: Control, no dipping + no mixing (table 2). To repeat and extend experiment 1, we added  
172 treatments with another earthworm species (*A. caliginosa*) and an alternative exposure method,  
173 where the earthworms were exposed to BA amended plant material (*Brassica napus* leaves) as food.  
174 In this case, both the external and internal tissues of the earthworms were exposed to the BA  
175 bacteria. Since results from experiment 1 had shown considerable earthworm weight increase and  
176 cocoon production during the first month, we judged that a shorter period would give reliable  
177 results. Hence, the experimental duration was shortened to 28 days for experiments 2 and 3. The  
178 treatments for experiment 2 were: control (C) without addition of BA; addition of BA by pouring 150  
179 ml of bacteria solution into the soil (S), like in the earlier experiments; addition of BA-amended  
180 *Brassica napus* leaves (L+); addition of leaves treated with water only (L); Addition of BA to the soil  
181 and addition of BA amended leaves (SL+). These five treatments were set up with *A. longa*  
182 (treatment 1-5) and with *A. caliginosa* (treatment 6-10). Experiment 3 included 4 treatments, dipping  
183 *A. longa* and *A. caliginosa* in BA solution and their respective controls (tables 1 and 2).

184

## 185 2.3 Statistical analysis

186 Data for earthworm mass, relative mass increase and cocoon production were analysed using a  
187 general linear model (GLM) with treatments as model components. When significant effects were  
188 found ( $P < 0.05$ ), Tukey's pairwise comparisons was used to compare treatment means. Minitab 16  
189 Software was used for all analyses.

## 190 3. Results

### 191 3.1 Experiment 1

192 The mortality of earthworms was rather high in this experiment. However, it did not differ  
193 significantly between treatments ( $P = 0.903$ ). In table 3, column n shows the number of populated  
194 mesocosms (with one or two live worms per mesocosm). The surviving earthworms grew well and

195 had on average increased from 2.2 g fresh mass at the start to 3.9 g at the end of the experiment  
196 (Table 3). There were no significant differences in earthworm individual growth between treatments  
197 after 29 days or 57 days from the start ( $P=0.25$  and  $P=0.69$ , respectively). Relative increment of  
198 earthworm biomass did not differ between treatments, either after 29 days ( $P=0.16$ ) or after 57 days  
199 ( $P=0.70$ ). Cocoon production amounted to a maximum of 0.25 cocoons per earthworm.

200

### 201 3.2 Experiment 2

202 In this experiment all earthworms survived and gained mass during the four week experimental  
203 period (Table 4). The results confirmed earlier observations in experiment 1 where there was no  
204 significant effect of adding a solution of BA to the soil ( $P>0.05$ ). In addition, offering leaves amended  
205 with the BA solution as food did not affect either growth or cocoon production of any of the two  
206 species ( $P>0.05$ ). However, relative increment in mass of *A. caliginosa* was larger in treatment SL+.cal  
207 ( $P=0.029$ ), with the combined addition of BA amended leaves and BA to the soil, as compared to the  
208 control. Cocoon production was considerably higher than in experiment 1. For *A. longa*, mean for the  
209 different treatments was between 2.92 and 4.17 cocoons per individual but did not differ  
210 significantly among treatments ( $P=0.921$ ). The corresponding value for *A. caliginosa* was between  
211 6.50 and 9.58 and it did not differ significantly among treatments either ( $P=0.421$ ).

### 212 3.3 Experiment 3

213 Effects of dipping earthworms into the BA solution are shown in Table 5. Growth of earthworms in  
214 absolute or relative terms did not differ between treatments ( $P=0.778$  and  $P=0.768$  for *A. longa* and  
215  $P=0.880$  and  $P=0.976$  for *A. caliginosa*) and mean values were larger than in experiment 2. Cocoon  
216 production did not differ between treatments either ( $P=0.417$  for *A. longa* and  $P=0.613$  for *A.*  
217 *caliginosa*), but mean values were considerably lower than in experiment 2 (table 5).

218

#### 219 4. Discussion

220 We aimed to conduct the experiments in soil conditions similar to the soil where the earthworms  
221 were collected, which was the agricultural soil of the Uppsala area. In a preliminary study, we had  
222 some initial problems with the experimental conditions and found that the clay dominated soil got  
223 very hard and impenetrable when drying out, which affected earthworm survival. Therefore keeping  
224 moisture within favourable limits is a must for successful lab experiments with earthworms. Lowe  
225 and Butt (2005) suggest a moisture content of 25% wet weight for cultures of *A. longa* and three  
226 other earthworms of the same family (Lumbricidae).

227 The conditions and viability of the worms is also a delicate issue. In experiment 1, the *A. longa*  
228 specimens used were collected from the field in October-November and had been kept in cultivation  
229 boxes for two months before the start of the experiment in February. High mortality and low cocoon  
230 production could be due to less favourable conditions of the worms during storage and perhaps also,  
231 because they were at the end of their life cycle. In experiments 2 and 3, which were done during the  
232 summer, the worms were in good conditions and moisture was regularly controlled. This ensured a  
233 100 % survival and high reproduction with little variation among replicates.

234 In all experiments, the earthworms were provided with sufficient amounts of feed. This is necessary  
235 when studying the interaction of earthworms with their environment since they would otherwise go  
236 into diapause or try to escape from the experimental soil units. Boström (1988) and Boström and  
237 Lofs-Holmin (1986) found that *A. caliginosa* went into estivation in an earthworm growth  
238 experiment, as soon as the added food resource was depleted. The feed was mixed into the soil of  
239 the mesocosms of all treatments, although *A. longa* is an anecic species that feeds mainly on the soil  
240 surface. According to some authors (e.g. Boyle, 1990, Lowe and Butt, 2002), earthworms, especially  
241 anecic and epigeic species, but also endogeics, grow better if the feed is placed on the soil surface.  
242 Lofs-Holmin (1983) however, found that mixing of feed into the soil gives just as good growth and  
243 reproduction, and it is practical since the risk of drying out of fodder is minimized and infection of the

244 substrate with, e.g. *Sciaridae* fly larvae is less likely to occur. Increase in mass for both species gives  
245 an indication that experimental conditions were favourable for their activity. In the case of *A.*  
246 *caliginosa* this increase ranged between 41 – 112 %, which is lower than the average 196% increase  
247 reported by Eriksen-Hamel and Whalen (2006), and by Vercesi et al (2006). If relative mass increase  
248 of the earthworms is a response factor, it is important to have specimens within the same mass  
249 range since relative growth rate decreases as the animals become larger. Based on this, it should be  
250 noted that, whereas juveniles were used in these experiments, in our experiment only adults and  
251 sub-adults were used, hence lower growth rates are expected. In the case of *A. longa*, their relative  
252 increase in body mass, ranging 50-138%, more than doubled the 25.81% obtained by Butt (1993) in a  
253 3-month long study. The higher relative increment in treatment 5 of experiment 2 (Table 4) could  
254 also be a result of somewhat smaller worms used in that treatment as compared to the other  
255 treatments. Cocoon production, which ranged 0.027-0.287 and 0.004-0.104 cocoon worm<sup>-1</sup> day<sup>-1</sup>, for  
256 *A. caliginosa* and *A. longa*, respectively, showed a higher production for the former than for the  
257 latter. Reported values for cocoon production in similar temperatures as in our study for *A.*  
258 *caliginosa* include averages of 0.09 and 0.221 cocoon worm<sup>-1</sup> day<sup>-1</sup> (Boström, 1988; Garvín et al.,  
259 2002; Vercesi et al., 2006); the lowest value may also be due to the inclusion of juveniles in the study,  
260 while the highest value is within our range. Butt (1993) and Holmstrup (1999) report that *A. longa*  
261 produced an average of 0.052 and 0.090 cocoon worm<sup>-1</sup> day<sup>-1</sup> in their experiments, respectively. The  
262 former being included in our range, while the latter is slightly higher. The low cocoon production in  
263 experiment 3 (Table 5) could also be a result of smaller specimens used as compared to those used in  
264 experiment 2– the earthworms may not yet have reached their full maturity and was still allocating  
265 most resources to body mass increase. Growth of individuals decline asymptotically with increasing  
266 body mass (Lowe and Butt, 2005) and there is a trade-off between body-mass increase and  
267 reproduction.

268 If laboratory reared earthworms had been used instead of specimens collected from the field,  
269 differences in fecundity, growth and survival between experiments due to seasonal changes caused

270 by the phenology of the earthworms could have been avoided. Under constant environmental  
271 conditions, earthworms have been shown to maintain both activity and reproductive conditions  
272 throughout the year. However, reproductive fatigue and high death rate can occur compared to  
273 those kept under fluctuating temperatures (Lowe and Butt, 2005). Although use of laboratory reared  
274 earthworms of the same age would have given more reliable and replicable data we chose to use  
275 field-collected ones since resources and time were not available to produce the amounts of  
276 specimens needed for our experiments.

277 This is the first study focussing on the impact of BCA bacteria on earthworms and from the results we  
278 can conclude that no harmful effects of *B. amyloliquefaciens* UCMB5113 on the tested earthworm  
279 species were recorded. Previous studies on the interaction between BCA bacteria and earthworms,  
280 focused on the opposite direction of the interaction: earthworm effect on bacteria, rather than  
281 bacteria effect on earthworms. These were conducted with the genus *Pseudomonas*, and the only  
282 reference made to the effect of these on earthworms was the lack of earthworm mortality during the  
283 experiments. No records of weight change or cocoon production have been reported (Stephens et al.  
284 1993; Doube, et al. 1994; and Schimdt, et al, 1997). Further studies of interactions of BCA bacteria  
285 and earthworms could concern other species of bacteria and earthworms. Earthworms by  
286 themselves also have positive effects on plant production. The underlying mechanisms for these  
287 positive effects include (i) biocontrol of pests and diseases, (ii) stimulation of microbial plant  
288 symbionts, (iii) production of plant growth-stimulating substances, (iv) soil structure improvements,  
289 and (v) increase of soil nutrient availability (Brown et al., 1999). Though recent studies focused on  
290 the first three mechanisms, van Groenigen et al. (2014) suggest that the last one is the most  
291 important. Earthworm activity influences the microbial community of soils directly by consumption,  
292 digestion and distribution of microorganisms and indirectly by modification of the soil environment  
293 (Byzov et al., 2007; Postma-Blaauw et al. 2006; Scheu et al., 2002; Schrader et al. 2013). This could  
294 either enhance or hamper the effects of bacterial BCAs. Their potential synergy becomes a relevant  
295 line for future research since the combined effects of earthworms and BCA bacteria on plant health

296 and productivity are of great interest for development of sustainable agricultural methods with  
297 minimum use of chemical pesticides and optimal use of ecosystem services.

## 298 5. Conclusions

299 From the experiments described above, we can conclude that the use of high doses of BA with  
300 concentrations of the same magnitude as maximally could be expected when the bacteria are used  
301 as BCA, is not harmful to the soil dwelling earthworms tested in this project. BA does not have  
302 negative impact on survival, growth or reproduction of two of the most common earthworm species  
303 in Swedish agricultural soils when these earthworms are exposed to BA by short-term external  
304 contact with high concentration (dipping), long-term external contact with lower dose (mixing into  
305 soil) and internal contact with the gut (feeding with BA-amended leaves). The combined effects of  
306 earthworms and BCA bacteria for promotion of plant health are of interest for the development of  
307 biological control and sustainable agriculture with reduced use of chemical pesticides.

308

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314

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428

429

430 TABLES

431 Table 1: Characteristics of three laboratory experiments testing effects of the biocontrol and plant-  
432 growth promoting *Bacillus amyloliquefaciens* UCMB5113 bacteria to the earthworms *Aporrectodea*  
433 *longa* and *Aporrectodea caliginosa*.

Experiment	1	2	3
Species	<i>A. longa</i>	<i>A. longa</i> <i>A. caliginosa</i>	<i>A. longa</i> <i>A. caliginosa</i>
Exposition methods	short term external; long term external and internal exposure.	long term external and internal exposure; internal exposure of the gut.	short term external
Vessels	3 L cylinders	3 L cylinders	6 L boxes
Moist Soil* (kg)	1.5	1.5	4.0
Treatments	4	10	4
Replicates	6	6	3
Starting date	February 2, 2014	July 28, 2014	August 20, 2014
Duration (days)	57	28	28

434

435 Notes: \*15% water content

436

437

438 Table 2. Treatments in the three lab experiment testing effects of *Bacillus amyloliquefaciens*

439 UCMB5113 (BA) on earthworms.

Experiment:	1	2	3
Treatment			
1	DS.long	C.long	D.long
2	D.long	S.long	C.long
3	S.long	L+.long	D.cal
4	C.long	L.long	C.cal
5		SL+.long	
6		C.cal	
7		S.cal	
8		L+.cal	
9		L.cal	
10		SL+.cal	

440

441 Notes: D= dipping earthworms into BA solution; S= addition of BA by pouring 150 ml of bacteria

442 solution into the soil; C= control; L+= addition of BA-amended *Brassica napus* leaves; L= addition of

443 leaves treated with water only; long= *Aporrectodea longa*; cal= *Aporrectodea caliginosa*

444

445

446 Table 3. Experiment 1: survival, biomass evolution and cocoon production of the earthworm  
 447 *Aporrectodea longa* in a mesocosm experiment testing influence of *Bacillus amyloliquefaciens*  
 448 UCMB5113 (BA).

Treatment	Start		29 days			57 days			Cocoons
	n	Fresh mass (g ind <sup>-1</sup> )	n	Fresh mass (g ind <sup>-1</sup> )	Relative increment %	n	Fresh mass (g ind <sup>-1</sup> )	Relative increment %	
1. DS	6	2.20 (0.38)	4	3.37 (0.17)	53.2 (23.9)	4	4.51 (0.65)	105 (38.8)	0.12
2. D	6	2.10 (0.39)	5	2.73 (0.38)	30.0 (6.6)	5	3.62 (0.40)	72.4 (25.9)	0.20
3. S	6	1.99 (0.34)	4	3.78 (0.41)	89.9 (16.7)	4	3.82 (0.86)	92.0 (37.1)	0.25
4. C	6	1.82 (0.35)	4	3.33 (0.40)	83.0 (10.7)	4	4.33 (0.44)	137.9 (10.6)	0.25
P value		0.91		0.25	0.16		0.69	0.70	

449 Note: Mean individual fresh mass (g per individual), relative increment from the start (Relative  
 450 increment %), and SE (within brackets) of the number of mesocosms per treatment with live  
 451 earthworms (n), which decreased during the course of the experiment; at the start, and at 29 days  
 452 and 57 days after start. Treatments: 1. DS=dipping into BA solution, mixing BA into the soil; 2.  
 453 D=dipping into BA solution; 3. S=mixing BA into the soil; 4. C= Control, no dipping or mixing into the  
 454 soil. P value=Testing differences between treatments with Anova.

455

456 Table 4. Experiment 2: individual fresh mass (g per individual) and individual cocoon production of  
 457 the earthworm *Aporrectodea longa* or *Aporrectodea caliginosa* in a mesocosm experiment testing  
 458 influence of *Bacillus amyloliquefaciens* UCMB5113 (BA).

459

*Aporrectodea longa*

Treatment	Initial fresh mass (g ind <sup>-1</sup> )	Final fresh mass (g ind <sup>-1</sup> )	Relative increment %	Cocoons per worm
1. Control	2.71 (0.20)	4.44 (0.23)	63.8 (8.7)	2.92 (0.93)
2. S	2.75 (0.20)	4.10 (0.22)	49.1 (5.4)	3.42 (0.80)
3. L+	2.53 (0.20)	4.15 (0.29)	64.0 (10.6)	3.67 (0.99)
4. L	2.72 (0.23)	4.71 (0.26)	73.2 (9.8)	3.33 (0.79)
5. S L+	2.86 (0.31)	4.81 (0.29)	68.2 (10.8)	4.17 (1.25)
Anova P	0.894	0.223	0.358	0.921

values

460

*Aporrectodea caliginosa*

Treatment	Initial fresh mass	Final fresh mass	Relative increment %	Cocoons per worm
6. Control	1.71 (0.13)	2.40 (0.17)	40.4 (3.9) B	9.58 (2.22)
7. S	1.61 (0.08)	2.62 (0.14)	62.7 (7.8) AB	9.75 (1.45)
8. L+	1.56 (0.10)	2.45 (0.13)	57.1 (5.3) AB	8.92 (1.08)
9. L	1.58 (0.06)	2.42 (0.10)	53.2 (3.7) AB	6.83 (1.48)
10. S L+	1.40 (0.07)	2.32 (0.13)	65.7 (13.7) A	6.50 (1.85)
Anova P	0.245	0.627	0.029 *	0.421

461

462 Note: Mean and SE (within brackets), n=6. 28 days experimental time (28/7-25/8 2014). Treatments:  
463 1. Control=no application of BA or *Brassica napus* leaves; 2. S=mixing BA into the soil, no leaves  
464 added; 3. L+=leaves with BA added, no BA into the soil; 4. L-=Leaves without BA added, no BA into  
465 the soil; 5. SL+= mixing BA into the soil, leaves with BA added. Testing differences between  
466 treatments = Anova P value (\* = significant difference). All earthworms in all treatments survived the  
467 experimental time. Values with different letters in a column indicate significant differences (P<0.05).  
468



469 Table 5. Experiment 3: dipping earthworms (*Aporrectodea longa*, *Aporrectodea caliginosa*) into a  
 470 bacteria solution of *Bacillus amyloliquefaciens* ( $10^7$  cells ml<sup>-1</sup>) and into water (control).

Treatment/species	Initial fresh mass (g ind <sup>-1</sup> )	Final fresh mass (g ind <sup>-1</sup> )	Relative increment %	Cocoons per worm
<i>A. longa</i>				
- Water dipping	2.18 (0.18)	4.47 (0.35)	105.0 (9.44)	0.68 (0.55)
- Bacteria dipping	2.23 (0.17)	4.35 (0.23)	95.1 (13.9)	0.17 (0.08)
Anova P values	0.826	0.778	0.768	0.417
<i>A. caliginosa</i>				
- Water dipping	0.98 (0.046)	2.03 (0.079)	107.1 (9.8)	0.67 (0.17)
- Bacteria dipping	0.99 (0.051)	2.01 (0.075)	103.0 (13.0)	0.83 (0.26)
Anova P values	0.906	0.880	0.976	0.613

471 Note: Mean and SE (within brackets) of fresh mass of earthworms at the start and after 29 days,  
 472 relative increment and cocoon production per individual. Mean of 4 worms per container for *A. longa*  
 473 and 6 worms per container for *A. caliginosa*, replicated in 3 containers with 4 l of soil. Testing  
 474 differences between treatments = Anova P value.

475