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1 **Review: Review of feeding conserved forage to horses: recent advances and**  
2 **recommendations**

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21 **Short Title:** Feeding conserved forage to horses

22 **Abstract:** The horse is a non-ruminant herbivore adapted to eating plant-fibre or forage  
23 based diets. Some horses are stabled for most or the majority of the day with limited or no  
24 access to fresh pasture and are fed preserved forage typically as hay or haylage and  
25 sometimes silage. This raises questions with respect to the quality and suitability of these  
26 preserved forages (considering production, nutritional content, digestibility as well as  
27 hygiene) and required quantities. Especially for performance horses, forage is often  
28 replaced with energy dense feedstuffs which can result in a reduction in the proportion of the  
29 diet that is forage based. This may adversely affect the health, welfare, behaviour and even  
30 performance of the horse. In the past 20 years a large body of research work has  
31 contributed to a better and deeper understanding of equine forage needs and the  
32 physiological and behavioural consequences if these are not met. Recent nutrient  
33 requirement systems have incorporated some, but not all, of this new knowledge into their  
34 recommendations. This review paper amalgamates recommendations based on the latest  
35 understanding in forage feeding for horses, defining forage types and preservation methods,  
36 hygienic quality, feed intake behaviour, typical nutrient composition, digestion and  
37 digestibility as well as health and performance implications. Based on this, consensual  
38 applied recommendations for feeding preserved forages are provided.

39

40

41 **Key words:** forage, health, hygiene, behaviour, requirements

42

43 **Implications:** Improved consistency in the terminology used for preserved forages  
44 and the adoption of consensual recommended minimal forage intake levels.

## 45 **Introduction**

46 The horse is a non-ruminant grazing herbivore adapted to eating plant-fibre or forage  
47 based diets. A wide range of plant species can be hydrolyzed and/or fermented within  
48 its specialized gastro-intestinal tract (GIT), thanks to the presence of a mixed microbial  
49 symbiotic population throughout the whole GIT, but particularly in the hindgut, which  
50 facilitate fibre digestion (de Fombelle *et al.*, 2003; Dougal *et al.*, 2013). Gastrointestinal  
51 tract (GIT) digestion provides energy mainly through the production of short-chain fatty  
52 acids (SCFA), mostly acetate, propionate and butyrate, also often described as volatile  
53 fatty acids (VFA's) (Merritt and Julliand, 2013). The whole digestive system is well  
54 adapted to a trickle (almost continuous intake of small amounts) feeding intake pattern,  
55 with the horse naturally foraging for around 10 – 15 hours a day (Ellis, 2010).  
56 Historically the energy demands of working horses were too high, and time too short,  
57 to be met by forage alone, especially as forage quality was often poor, grazing limited  
58 and it was difficult to distribute and transport bulky forages. Therefore, oats, barley,  
59 beans and root vegetables were used as a major component of the diet (Stewart,  
60 1838). The inclusion of energy rich feedstuffs (particularly cereals and vegetable oils)  
61 in the diets of many horses, especially performance horses, continues today despite  
62 the availability of better quality forages (Richards *et al.*, 2006, Lindburg 2013). An  
63 increase in starch intake, often in combination with a reduction in the amount of forage  
64 provided, can have health, welfare and performance consequences.

65 Due mainly to environmental conditions and the lack of availability/undesirability of  
66 grazing, many horses are fed preserved forages (hay, haylage and sometimes silage),  
67 in particular preserved grass, either at specific times of the year or all year round and  
68 therefore preserved forages provide all or part of their forage intake. During the 2012  
69 meeting of the European Workshop on Equine Nutrition (EWEN) it was agreed that

70 there was a need for a consensus paper on the topic of preserved forage feeding to  
71 horses. The aim of this review paper is therefore to summarize recent findings and to  
72 provide consensual applied recommendations for feeding preserved forage.

73

#### 74 **Forage types and preservation methods**

75 Roughages are high fibre feeds (Morrisson, 1956) generally obtained as a crop  
76 residue or a by-product (Ellis *et al.*, 1988) e.g. straw and cereal hulls. Forages are also  
77 high fiber feeds obtained by cutting and preserving the whole plant (except roots).  
78 Although they define different products, the words forage and roughage are typically  
79 used interchangeably. From a behavioural aspect the term 'foraging' encompasses all  
80 feed intake activities of horses both on pasture and in housing situations.

81 Cut grass air dried/wilted in the field or a barn is termed *hay*, whereas *silage* is forage  
82 preserved moist and airtight, and thus fermented (McDonald *et al.*, 1991) The term  
83 haylage was originally used to describe silage with a DM content of around 50 %  
84 (Gordon *et al.*, 1961) and although haylage (and hay/silage) can be harvested at any  
85 stage of plant maturity today *haylages* are typically grown and cut at later growth  
86 stages similarly to hay, but baled before becoming dry resulting in DMs typically >50  
87 but <70% although occasionally up to 85% (see Fig. 1 and supplementary Table S1).  
88 Hay baled with too high a moisture content due to unsuitable weather conditions at  
89 cutting time and especially during wilting, insufficient turning especially if in rows  
90 (rather than being wilted widespread which helps speed up wilting, thereby helping to  
91 reduce protein degradation and loss of nutrients) etc. allows the development of fungal  
92 spores/bacteria and increases the risk of mycotoxin development. To stop undesired  
93 microbial growth, roughage/forage therefore has to be preserved under controlled  
94 conditions (Fig. 2). Hay and straw should be preserved during conditions of low water

95 activity in the crop (i.e. preserved at a DM content ideally above 85%) and given that  
96 the bales will be exposed to air, relative humidity must be lower than required for mould  
97 growth at the given temperature (Fig. 2) and therefore ideally <70%. Forages may also  
98 be preserved utilizing airtight conditions, with or without lactic acid fermentation. In  
99 silage the water soluble carbohydrates in the forage are fermented by anaerobic  
100 lactic acid bacteria (naturally occurring on the crop or added as inoculants (Fig. 1). For  
101 sufficient lactic acid production to occur, the crop DM content must ideally be around  
102 30% or less and certainly <50 % (Fig. 3). If the acids produced sufficiently increase  
103 hydrogen ion concentrations, undesirable microbial growth will be inhibited. To confirm  
104 proper ensiling of forages (without butyrate) with a DM 15-50%, pH can be used as an  
105 indicator. Weissbach, 1996 and Spörndly *et al.*, 2003 suggest that the pH should be  
106 less than:  $0.0257 \times \text{DM}\% + 3.71$  (Weissbach, 1996) but according to Field and Wilman,  
107 1996 a higher pH ( $0.0028 \times \text{DM}(\text{g}/\text{kg}) + 4.209$  (equation estimated from their figure)  
108 might be allowed if silage is preserved in bales and not in bunkers. Preservation at  
109 higher DM contents relies on maintenance of airtight packaging not a low pH (Mihin,  
110 1940). In haylage/silage bale production, at least 4 layers of film are needed to  
111 promote good preservation (Keles *et al.*, 2009; McEniry *et al.*, 2011) but adding layers  
112 (6- 10) increases the CO<sub>2</sub> content (Müller, 2005) and for bales to cope with handling,  
113 transportation, birds and long-term storage 8 layers or more are recommended  
114 (Jacobsson, 2002; Spörndly pers. com.). In haylages with very high DM contents there  
115 may be an increased risk of perforation (allowing air entrance and mould growth) by  
116 stiff and sharp stems. To facilitate safer preservation and storage such haylages  
117 require more layers of film than forages with a lower DM content. Some professional  
118 horse haylage producers may use 12-20 layers (Jansson A, pers. com.). More  
119 research is needed into the impact of the number of layers used. Mature and rough

120 crop may be difficult to preserve correctly since more air can be trapped in such bales,  
121 increasing the risk of localized mould growth. Higher density baling, late tossing, lower  
122 dry matter (75%), as well as rain occurring after cutting strongly increased mould  
123 counts in hay (Seguin *et al.*, 2010) but baling for haylage when too dry (86-88% DM)  
124 also increased mould counts (Martinson *et al.*, 2011).. Bales of forage with very low  
125 DM contents (<30 %) are also at risk of losing airtightness due to their heavy weight,  
126 plant structure collapse during fermentation, formation of effluent etc. all of which result  
127 in increased pressure on the film. In forage with DM content < 40 % *clostridial*  
128 fermentation can also occur (McDonald *et al.*, 2002) and the prevalence of *clostridial*  
129 fermentation seems to be higher in bales with unchopped vs chopped forage (Pauly,  
130 1999). In well-made haylage, whilst there will still be some microbial activity, reduction  
131 in WSC and production of by-products of fermentation this will be much more limited  
132 compared with that for silage (Müller *et al.*, 2007; Muhonen *et al.*, 2009).

133 Figures 1, 2 and 3 here

#### 134 **Nutritional composition**

135 The nutritive value of forage to the horse is determined predominantly by its nutrient  
136 content and digestibility, which in turn may be influenced by level of intake and feed  
137 interactions. Nutrient content depends on many plant related factors as well as the  
138 environment (Buxton, 1996). Herbage maturity at harvest, however, is a key influencer  
139 (Virkajärvi *et al.*, 2012). As the plant matures, the fibre fraction increases and crude  
140 protein decreases, leading to a decline in digestibility plus overall energy availability  
141 (Ragnarsson and Lindberg, 2008; 2010; Müller, 2012) (Fig. 4). Nutrient content is also  
142 closely related to plant species and morphology, with legumes typically higher in  
143 protein and calcium than grasses. The proportion of leaf to stem will also affect

144 nutritive value, even between cultivars of the same species (Van Soest, 1994;  
145 Bélanger and McQueen, 1997).

146 Figure 4 Here

147 Forage quality is also influenced by management factors especially fertilizer  
148 applications, harvest techniques and storage conditions (Van Soest, 1994; Cookson  
149 *et al.*, 2000; Rotz, 2003). Hay nutrient losses are normally higher during harvest (e.g.  
150 plant leaves left in the field due to mechanical handling of the dried crop) in contrast  
151 with silage where losses are larger during storage, mostly due to fermentation  
152 activities, air infiltration and silage effluents (McGechan, 1989, 1990; Dürr, 2004 ;  
153 Müller, 2012). Environmental factors affecting herbage development (including  
154 temperature, precipitation, solar radiation and soil nutrient availability) obviously can  
155 vary with geographical location resulting in a range of climatic and soil conditions.

156 Not surprisingly forage chemical composition and consequent nutritional value  
157 therefore shows a great variation between regions, crop years, harvest time, seasons  
158 and even within farming places (see Supplementary Table S1). The variability  
159 highlights the importance of forage analysis when designing forage based rations,  
160 especially when feeding brood mares, growing and performance horses (Jansson *et al.*  
161 *et al.*, 2012) or those with particular clinical conditions (such as laminitis, HYPP etc.).  
162 Ideally mineral content should also be analysed. Legume forages tend to have higher  
163 protein and calcium contents than grass forages and this needs to be taken into  
164 consideration when formulating the total ration (which should preferably be based on  
165 nutrient analysis).

166

167 **Digestion and Digestibility**



168 Digestion can be defined as the process in the gastrointestinal tract by which forage  
169 constituents are converted by physical and chemical breakdown into substances  
170 (mainly SCFA) that can be absorbed and assimilated by the body (Argenzio *et al.*,  
171 1974). Water soluble, as well as enzymatically digestible carbohydrates (WSC, starch  
172 non-starch polysaccharides, soluble fibre) are digested from the stomach onwards  
173 along the whole GIT as long as they are not trapped within insoluble fibre. Insoluble  
174 fibre digestion depends exclusively on microorganisms that have the ability to  
175 hydrolyze the predominant  $\beta$ 1-4 linked polysaccharides of cell walls. It occurs primarily  
176 in the horse's hindgut with a limited amount of fermentation in the foregut (de Fombelle  
177 *et al.*, 2003; Jouany *et al.*, 2009). The extent of any forage digestion depends on both  
178 the fibrolytic microbial activity of the hindgut ecosystem and the total time during which  
179 parietal components are exposed to this activity (Merritt and Julliard, 2013). This  
180 duration is defined as the mean retention time (MRT) of digesta (Miyaji *et al.*, 2008b).  
181

182 Several intrinsic and extrinsic factors can affect forage digestion and digestibility  
183 (digestibility = total nutrient/energy ingested minus nutrient/energy excreted and  
184 expressed as % of total ingested). The Influence of intrinsic factors such as horse  
185 breed, individuality and age has been studied on forage digestion and digestibility. It  
186 has been anecdotally suggested, for example, that 'easy keepers' may have higher  
187 digestion efficiency than other horses. However, in a controlled study (Ragnarsson  
188 and Jansson, 2011) where two haylages (cut at early and late maturity stage) were  
189 fed to both Icelandic horses (easy keepers) and Standardbred horses no such effect  
190 could be observed (Table 1). Recent studies also showed that DM, organic matter and  
191 neutral detergent fibre fraction apparent digestibility were not different between  
192 weaned foals (6 and 12 months) and adults (14 years) (Ringler *et al.*, 2009; Earing *et*

193 *al.*, 2013), which complemented and confirmed previous data reporting no variation of  
194 total apparent digestibility of DM between weanlings (5 months), and those aged 8  
195 and 12 months old (Cymbaluk *et al.*, 1989). At two months of age, the SCFAs profile  
196 of foal's faeces remained constant and cellulolytic bacterial concentration was  
197 comparable with adult values suggesting that the fibre-degrading capacity in foals was  
198 established by two months of age (Faubladier *et al.*, 2013).

199

200 Extrinsic factors such as botanical characteristics are known to influence growth and  
201 metabolism of plants, and therefore their nutritive value as forages. In horses, the  
202 average DM digestibility decreased significantly in forages having higher NDF and  
203 lower crude protein contents (Edouard *et al.*, 2008). More specifically, alfalfa has  
204 higher DM and CP digestibility coefficients than tall fescue, caucasian bluestem  
205 (Crozier *et al.*, 1997), coastal Bermuda grass (Sturgeon *et al.*, 2000; Potts *et al.*, 2010)  
206 and Matua grass (Sturgeon *et al.*, 2000). Alfalfa also had higher OM digestibility than  
207 grass hays (LaCasha *et al.*, 1999) and more specifically coastal Bermuda grass (Potts  
208 *et al.*, 2010). Digestion and energy and crude protein content of forage is greatly  
209 affected by the stage of maturity of plants (Fig 4; Table 2).

210 The type of forage preservation, however, appears to have limited impact on apparent  
211 digestibility. When hay and haylage originating from the same crop (same  
212 harvest/batch) were compared, digestibility did not differ (Bergero and Peiretti, 2011)  
213 although when hay and silage from the same crop was compared, digestibility was  
214 slightly higher in silage compared to hay (Muhonen *et al.*, 2008b). Feed changes from  
215 hay to silage or haylage (even from the same harvest/batch) may, however, affect the  
216 microflora as well as the DM of the hindgut contents (Muhonen *et al.* 2008, Muhonen  
217 2009) Similarly changes between forage batches with different CP contents should

218 be made slowly, as colonic pH, for example, has been shown not to be stable within  
219 3 weeks after such a change (Muhonen *et al.*, 2008a) although further studies are  
220 needed to assess the importance of these alterations.. Until we do know more it is  
221 recommended that changes between forage batches should therefore be made  
222 carefully to minimise the risk of disturbances. In support of these recommendations  
223 the incidence of certain types of colic has been shown to increase especially in the  
224 first 7 days (but up to 28days) after a change in forage feeding (Hillyer et al 2002).

225

226 Insert Table 1 and 2

227

## 228 **Hygienic quality**

229 This is a key issue, as forages fed to horses have often been reported to be of poor  
230 hygienic quality (Wichert *et al.*, 2008) which can lead to significant health problems for  
231 example, mould spore exposure, especially from *Aspergillus fumigatus* has been  
232 implicated in the aetiology of recurrent airway obstruction in horses (Pirie 2014).  
233 Although the term “poor hygienic quality” is not well defined, it may include biological  
234 contaminants (e.g. pests, microorganism and their related toxins), chemical  
235 contaminants (e.g. fertilizer, heavy metals), and physical contaminants such as soil.  
236 In that context, feed hygiene, as stated by EU legislation (Regulation (EC) No  
237 1831/2003), includes all aspects that must be considered in order to produce, sell and  
238 feed a safe feedstuff that will not result in any harmful effects on the animal, and  
239 therefore applies to forage as well. EU legislation (EC No 32/2002) defines maximum  
240 acceptable levels for a variety of contaminants in feedstuffs including for heavy metals,  
241 aflatoxin B1, rye ergot and substances such as pesticides.

242

243 Several parameters can be used to evaluate hygienic quality including feel (e.g. dry,  
244 clammy), smell (e.g. typical, mouldy), colour (e.g. green, bleached), macroscopic  
245 findings (e.g. presence of sand, soil, dead animals) and/or microbial evaluation  
246 (Wichert *et al.*, 2008, Kamphues 2013, Wolf *et al.*, 2014). Knowing the DM content is  
247 key, as discussed above, as microorganisms need water for survival and multiplication  
248 (Kamphues 2013). A macroscopic evaluation should routinely be undertaken for  
249 obvious moulds, as well as the presence of sand/soil and other potential contaminants  
250 especially poisonous plants. Senecio spp., and *Taxus baccata* have been suggested  
251 to be the main poisonous plants for horses (Berny *et al.*, 2010) although regionally  
252 other poisonous plants may be important e.g. vitamin D-intoxication via Golden oat  
253 grass in parts of Germany (Bockisch *et al.*, 2015).

254 Much attention is given to the microbiological analysis of feedstuffs but there are  
255 significant effects of the analytical methods used. Pre-treatment, incubation  
256 temperature, type of incubation substrate and incubation duration can all significantly  
257 influence what is cultivated (Raymond *et al.*, 2000; Müller *et al.*, 2011). For example,  
258 the growth of thermophilic bacteria such as actinomyces is supported by an incubation  
259 temperature of 55°C for three days, whereas the growth of mesophilic aerobic bacteria  
260 is supported by a lower incubation temperature (25°C), but a longer incubation time (7  
261 days, Raymond *et al.*, 2000). In some European countries, the characterization and  
262 benchmarking for bacteria, moulds and yeasts in feedstuffs has been standardized  
263 (Tables 3 and 4) to provide upper acceptable levels of contamination which are  
264 thought not to have any adverse effects on horse health. For this purpose  
265 microorganisms are classified as being either epiphytic (i.e. normal contaminants that  
266 are present even under optimal conditions) or spoilage inducing. Importantly, both  
267 types may impact health (e.g. respiratory problems by moulds). However, it should be

268 emphasised that whilst these benchmark levels can be used to describe spoilage, the  
269 consequences on equine health through exceeding the benchmark levels requires  
270 more in-depth evaluation.

271

272 Undesirable bacteria, moulds and yeasts multiply under adverse conditions such as  
273 rainfall, high humidity, and high ambient temperatures or due to vectors like mites. As  
274 outlined above a DM content of >85% reduces microbial activity

275

276 Mycotoxins such as ergot alkaloids, , zearalenone, deoxynivalenol, fumonisin B1 and  
277 B2, ochratoxin A or aflatoxin B1 are secondary metabolites produced by fungi that are  
278 capable of causing toxicity (see review Riet-Correa *et al.*, 2013). However, little is  
279 known about the impact of mycotoxins on equine health status. Several mycotoxins  
280 are designated as undesirable substances with maximum levels in human food,  
281 whereas only aflatoxin B1 and rye-ergot are currently designated as undesirable  
282 substances with maximum levels for animal feedstuffs. Further analyses, therefore,  
283 may be required including detailed mycotoxin determinations e.g. clinical signs of  
284 leukoencephalomalacia requires fumonisin analysis especially when corn silage with  
285 corn cobs is being fed to the affected horses. Whilst it is impossible to fully eliminate  
286 mycotoxins, it is important to reduce contamination by optimizing harvesting and  
287 storage conditions.

288 Botulism occurs following exposure to toxins (8 different serotypes) produced by  
289 anaerobic spore-forming bacterium *Clostridium botulinum* and other botulinum toxin-  
290 producing clostridia (Galey 2001). In adult horses, food-borne botulism is acquired by  
291 the ingestion of preformed toxins (mainly type B, C or A). Feeding big bale silage or  
292 haylage have been often associated with botulism outbreaks in horses, mules or

293 cattle (e.g. Ricketts *et al.*, 1984; Divers *et al.*, 1986; Wollanke 2004; Myllykoski *et al.*,  
294 2009), although there have been outbreaks linked with hay feeding (Wichtel and  
295 Whitlock 1991; Johnson *et al.*, 2010) . Botulinum toxin production typically occurs due  
296 to contamination with animals, soil or poultry slurry. Equivocal results have been  
297 reported regarding the potential risk of silage or haylage contamination with  
298 *Clostridium botulinum* spores through using the wastage from biogas anaerobic  
299 digesters for fertilization (Müller *et al.*, 2013; Neuhaus *et al.*, 2015).

300

### 301 *Methods of sampling*

302 A good sampling procedure is crucial so that any analysis (e.g. nutrients, microbial  
303 counts) represents the mean value for the whole batch. For the official feed control EU  
304 legislation (EC No 691/2013) suggests for example that 5 up to 40 individual samples  
305 should be aggregated to form an initial overall sample of between 1 (low specific  
306 gravity, e.g. hay or straw) and 4 kg as fed (e.g. silage). .

307 *Table 3 and 4 here*

### 308 ***Forage Intake Behaviour and Welfare considerations***

309 Free-ranging horses perform 10-15 individual feed-bouts/day and forage for 12.5±2.5  
310 hours per day with recordings as high as 18 hours (Ellis, 2010, Supplementary Table  
311 S2). 'Non-foraging' bouts are rarely longer than 3 hours duration (2±1.3) (Souris *et al.*,  
312 2005; Van Dierendonck *et al.*, 1996; Hallam *et al.*, 2012, Ellis *et al.*, 2015;). Even when  
313 stabled, horses tend to spend a minimum time (8.5-12 hrs/day) on foraging related  
314 behaviours, at times 'topping up' their food intake behaviour by ingesting wood-  
315 shavings and performing coprophagy for up to 3hrs per day (Ellis *et al.*, 2006, Ellis

316 2010). Curtis *et al.* (2011) suggested the potential for wood-shaving intakes of up to  
317 3.5kg in a few individuals fed DM restricted diets as part of a weight loss programme  
318 and this is in line with considerable amounts of woodshavings reported in the stomach  
319 of horses on a low forage diet (Boswinkel *et al* 2007) . Intake times for chopped  
320 lucerne/alfalfa are about twice as fast as those for long hay (Ellis, 2010). Chopping  
321 very late cut hay to 3.5 cm particle length did not shorten intake times in 8 horses but  
322 adding chopped forage (~2.5cm or 4cm) to pellets at 30% inclusion rate doubled  
323 concentrate feed intake times (Ellis and Hill, 2002; Ellis *et al.*, 2005). Replacing hay  
324 with ground-hay pellets reduces chewing time/kg by 75% (Ellis *et al.*, 2010) which may  
325 partially explain the increased intakes (to 4-5% of BW) seen by Argo *et al.* (2002) and  
326 Henneke and Callaham (2009) when they fed only pellets compared to a chaff-feed or  
327 hay. Elia *et al.* (2010) found that foraging through wood-shaving bedding increased by  
328 3.5 hrs/day, when feeding just hay pellets compared to feeding hay. This strong  
329 motivation to spend a minimum of 8 (stabled) to 12 (grazing) hours on intake  
330 behaviours, irrelevant of the energy density or composition of feedstuffs is supported  
331 by many studies on voluntary intake behaviour (Supplementary Table S2) and night  
332 time observations of stabled horses (Ellis *et al.*, 2015). In addition stabled horses rest  
333 for 3-4 hours maximum before re-commencing foraging-related behavior (Ellis *et al.*,  
334 2015).

335 Furthermore, in a race horse population of 2900 animals, those fed <6.8kg/day of  
336 forage showed a significant increase in abnormal behaviours including oral  
337 stereotypies and weaving (McGreevy *et al.*, 1995). Lack of foraging opportunity has  
338 been directly linked to the onset of oral stereotypies in foals (Nicol *et al.*, 2002) and to  
339 possible stereotypic pre-cursor behaviour (increased water play and drinking,  
340 locomotion) in 3 out of 5 feed-restricted ponies (Dugdale *et al.*, 2010). Appetitive

341 behaviours have a positive feedback on motivation through the brain's pleasure  
342 centres which become active as the horse pursues its goal (McBride and Hemmings,  
343 2005). If the motivation to forage/chew is not fulfilled, other behaviours may replace  
344 the original goal achievement behaviour as highlighted in several mammals (Hughes  
345 and Duncan, 1988).

### 346 **Health considerations**

347 Horses as previously stated are adapted to a slow and almost continual intake of a  
348 diet rich in structural fibre and low in rapidly hydrolysable carbohydrates. For many  
349 commonly encountered equine health issues, forage feeding management is an  
350 important factor in helping to reduce risk and maintain health.

351 Particle length influences motility and transit time within the GI tract, as larger particles  
352 move more slowly (Drogoul *et al.*, 2000) and forage also influences gastric emptying  
353 rate and/or the passage rate through the small intestine (Jensen *et al.*, 2012). A more  
354 stable (Willing *et al.*, 2009) and diverse microbial population, with a larger core, is  
355 found when a forage only diet is fed especially compared to a sugar and starch rich  
356 ration (Dougal *et al.*, 2014). Stabling and feeding preserved forage results in a change  
357 in gut motility and higher DM faeces despite an increased water intake (Williams *et al.*,  
358 2011, 2015). Horses have a lower pH in the proximal stomach during early morning  
359 (1:00–9:00 AM), when stomach-fill tends to be lower especially in stabled animals  
360 (Husted *et al.*, 2008). It is therefore not surprising that several nutritional risk factors  
361 for equine gastric ulcer syndrome (EGUS), colic and diarrhea have been identified and  
362 many of them are related to limited, reduced or a changed intake of forage (see table  
363 5).



364 Recurrent airway obstruction (RAO) is the most common cause of chronic coughing  
365 in horses in temperate countries, with up to 14% prevalence in the UK (Hotchkiss *et*  
366 *al.*, 2007). This lower airway inflammatory disease results in a range of clinical signs  
367 from exercise intolerance/poor performance, to severe expiratory dyspnea (Pirie *et al.*,  
368 2002; Pirie *et al.*, 2003; McGorum and Pirie 2008). Exposure to airborne organic dust  
369 (mostly endotoxins) via stabling and feeding of hay/straw with a high mould count  
370 plays a primary role (Couetil and Ward 2003) as discussed above.

371 The management strategies for horses at increased risk of several conditions, such  
372 as certain forms of laminitis (including those with the Equine Metabolic Syndrome and  
373 Pituitary Pars Intermedia Dysfunction) as well as various muscular disorders including  
374 the Equine Rhabdomyolysis Syndrome, includes reducing the intake of non-structural  
375 carbohydrates (NSC) (MacLeay *et al.*, 2000; McKenzie *et al.*, 2003; Valentine *et al.*,  
376 2001; Hunt *et al.*, 2008). Such horses are often put on restricted or no pasture access  
377 (especially when the levels of fructans, starch and sugar are high or there is a high  
378 herbage yield) and are fed a diet based on preserved forage or forage replacers (e.g.  
379 commercial fibre based products) with a known and low content of NSC (<10-12%  
380 DM) to minimize post-feeding glycemic and insulinaemic responses (Borgia *et al.*,  
381 2011; Geor and Harris 2013; Harris *et al.*, 2013). Recent observations also indicate  
382 that forage CP content might be of importance for the insulin response (Ringmark and  
383 Jansson 2013).

384 Table 5 here

### 385 **Performance considerations**

386 Historically, the use of forage in diets of performance horses has been limited  
387 (Jansson and Harris 2013). Recent studies (Connysson *et al.*, 2006; Muhonen *et al.*,

388 2008b; Connysson *et al.*, 2010; Essén-Gustavsson *et al.*, 2010; Jansson and Lindberg  
389 2012; Ringmark *et al.*, 2012; Ringmark *et al.*, 2015) show that forage-only diets can  
390 meet the high energy requirements of horses in very heavy training (equal to or more  
391 than twice maintenance requirements) if the energy density is high enough. The  
392 energy density of such forage must correspond to at least 10.5 MJ ME/kg DM  
393 (Jansson and Lindberg 2012; Ringmark 2014). These studies indicate no adverse  
394 effects on performance with exception for slightly lower (ca -10 %) muscle glycogen  
395 contents within 3 days after high intensity work compared to a high starch diet  
396 (Jansson and Lindberg 2012). However, very high muscle glycogen contents and  
397 indications of rapid glycogen recovery have been documented on forage-only diets  
398 providing crude protein intakes above current feeding recommendations and forage  
399 WSC contents of 8.5-13.5 % of DM (Essén-Gustavsson *et al.*, 2010; Ringmark, 2014).  
400 In the study by Jansson and Lindberg (2012) plasma lactate response was decreased,  
401 venous pH and blood glucose was increased during exercise on the forage-only diet  
402 compared to the traditional high starch diet, indicating a metabolic pattern that might  
403 improve performance. A recent study (Ringmark 2014) has also shown that it is  
404 possible to get Standardbred yearlings into racing condition at the age of 3, and also  
405 to win races, on a high-energy forage diet supplemented only with minerals and  
406 vitamins. Horses also maintained plasma volume longer during 12 h of feed  
407 deprivation on a forage only-diet compared to a high starch diet (Connysson *et al.*,  
408 2010). Altogether these studies suggest that high energy preserved forage can be an  
409 alternative to high starch feeds in exercising horses and that such diets might promote  
410 both health and performance.

411 ***Author's Recommendations for best practice***

412 Based on the above and other published work (as well as personal views where  
413 stated).

414

415 A. The general use of the following terms is recommended

416

417 I. *haylage* for forages stored airtight and with DM content  $\geq 50\%$ .

418 II. *silage* for forages stored airtight with DM contents below 50%

419 III. *hay* for forage preserved at a DM content ideally above 85%

420

421 B. To fully understand the value of a forage, and its impact as part of the horse's  
422 diet, nutrient analysis is recommended and an estimation of the energy content  
423 should be made. This becomes particularly important when feeding horses with health  
424 disorders (e.g. low BCS, obesity, laminitis, PSSM).

425

426 C. Hygienic quality of forages must be a key consideration as poor hygienic  
427 quality can lead to significant health problems. As a minimum, regular visual and  
428 olfactory inspection should be undertaken and poor quality forage appropriately  
429 disposed of. Benchmarks for microorganism contamination should be considered.

430

431 D. Changes in forage should be done gradually especially if the nutrient  
432 composition (energy, protein, WSC) is unknown or known to differ considerably due  
433 to changes in plant-species, growth stage and preservation technique. In such  
434 instances at least 2 and possibly more than 3 weeks adaptation period may be  
435 required.

436

437 In terms of forage requirements for horses it is recommended that:

438 1. The basis of any horse's diet should be fresh or preserved forage  
439 (recommended particle length >2.5 cm) offered ideally ad libitum or spread  
440 throughout the day to avoid prolonged periods (i.e. >4-5hrs) in a stable without  
441 foraging opportunity. However, in older horses with chewing difficulties, using ground  
442 high fibre products in a soaked mash form may be beneficial.

443 2. A horse requires sufficient forage fed in a form that supports species specific  
444 feed intake behaviour ideally for a minimum of 8 hrs and preferably 10 hours/24hrs.

445 3. The amount of preserved forage in the ration should be calculated on a g DM  
446 /kg BW rather than % of ration or an as-fed basis due to the great variation in DM  
447 content of different forages.

448 4. The lower limit of daily forage intake should be 15g DM/kg BW in addition to  
449 complementary (concentrate/compound) feeding, with an absolute minimum  
450 recommendation of 12.5g DM/kg BW. Previously recommended minimums of forage  
451 at 8-10g DM per kg BW/day are not acceptable according to the latest understanding  
452 of equine ethological needs and health considerations.

453 5. Any lower amounts should only be fed solus (i.e. no other fibre provision), under  
454 exceptional clinical circumstances (e.g. as low energy providing hays for animals  
455 undergoing severe restriction for weight loss purposes, post-surgery etc.), under  
456 veterinary supervision and with an appropriate forage vitamin/mineral/protein  
457 balancer.

458 6. Straw, if required, should be introduced into the diet very slowly, which helps  
459 the horse to adapt its chewing behavior and reduce the risk of impaction, although this  
460 remains a significant risk with certain individuals. The risk of gastric ulceration also

461 may increase when straw is the main roughage. Some of the authors (PH, AE, VJ, NL)  
462 *personally recommend* that not more than 30% of the forage DM ration should be  
463 straw (other than for donkeys). Others (AJ, IV) may recommend higher proportions  
464 providing the straw is of a good hygienic standard and the overall ration is balanced  
465 for protein vitamins, minerals and trace elements.

466 7. For horses requiring increased energy intake due to reproduction status or  
467 work level, less mature forages that have a higher energy content, should be  
468 considered. Forages cut at an earlier plant maturity stage are less likely to require  
469 protein supplementation.

470 8. Intakes above those recommended here may be fed or consumed by horses,  
471 as long as a healthy energy balance (body condition) is maintained. If body weight  
472 increases even when feeding the minimum recommended intake, a forage with a  
473 lower energy content should be fed rather than reducing the dry matter forage intake.

474

475

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482 and Andreas Olt.

483

484

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633

634

635 Table 1. *Coefficients of total tract apparent digestibility in Icelandic and Standardbred*  
636 *horses fed the same early and late cut forages (after Ragnarsson and Jansson, 2011)*

	Early cut Icelandic	Early cut Standardbred	Late cut Icelandic	Late cut Standardbred	SE
OM	0.536 <sup>a</sup>	0.565 <sup>b</sup>	0.431	0.427	0.006
CP	0.636	0.660	0.478	0.479	0.012
NDF	0.517	0.536	0.322	0.320	0.008
ADF	-	-	-	-	-
Energy	0.517 <sup>a</sup>	0.540 <sup>b</sup>	0.407	0.400	0.007

637 <sup>a, b</sup>. Values in the same row without common superscripts differ ( $p < 0.05$ ) and indicate breed  
638 differences.

639 OM : Organic matter, CP : Crude Protein; NDF : Neutral detergent fibre; ADF : Acid Detergent  
640 fibre

641

642

643

644

645 Table 2. *Coefficients of total tract apparent digestibility of haylages cut (first cut) at different*  
646 *stages of maturity (after Ragnarsson and Lindberg 2008; Ragnarsson and Lindberg, 2009)*

647

	Timothy haylage					Mixed grass haylage				
	Cut 1	Cut 2	Cut 3	Cut 4	SE	Cut 1	Cut 2	Cut 3	Cut 4	SE
DM	0.716 <sup>a</sup>	0.626 <sup>b</sup>	0.513 <sup>c</sup>	0.457 <sup>d</sup>	0.011	0.691 <sup>a</sup>	0.616 <sup>b</sup>	0.619 <sup>b</sup>	0.556 <sup>c</sup>	0.010
OM	0.747 <sup>a</sup>	0.647 <sup>b</sup>	0.527 <sup>c</sup>	0.485 <sup>c</sup>	0.010	0.710 <sup>a</sup>	0.630 <sup>b</sup>	0.640 <sup>b</sup>	0.578 <sup>c</sup>	0.010
CP	0.809 <sup>a</sup>	0.735 <sup>b</sup>	0.642 <sup>c</sup>	0.639 <sup>c</sup>	0.014	0.765 <sup>a</sup>	0.740 <sup>ab</sup>	0.708 <sup>ab</sup>	0.688 <sup>b</sup>	0.015
NDF	0.770 <sup>a</sup>	0.646 <sup>b</sup>	0.516 <sup>c</sup>	0.440 <sup>d</sup>	0.009	0.717 <sup>a</sup>	0.584 <sup>bc</sup>	0.594 <sup>b</sup>	0.520 <sup>c</sup>	0.013
ADF	0.746 <sup>a</sup>	0.629 <sup>b</sup>	0.480 <sup>c</sup>	0.400 <sup>d</sup>	0.008	0.685 <sup>a</sup>	0.545 <sup>b</sup>	0.555 <sup>b</sup>	0.485 <sup>b</sup>	0.017
Energy	0.733 <sup>a</sup>	0.633 <sup>b</sup>	0.515 <sup>c</sup>	0.468 <sup>c</sup>	0.006	0.670 <sup>a</sup>	0.600 <sup>b</sup>	0.595 <sup>b</sup>	0.560 <sup>b</sup>	0.011

648 <sup>a, b, c</sup>, Values in the same row without common superscript differ (p<0.05).

649 DM : Dry matter; OM : Organic matter, CP : Crude Protein; NDF : Neutral detergent fibre; ADF  
650 : Acid Detergent fibre

651

652

653

654 Table 3: *Classification of microorganisms in feedstuffs according to Kamphues (2013)*

Microorganism	Classification	Category	Species examples
Aerobic bacteria	Epiphytic	1	<i>Flavobacterium</i>
			<i>Pseudomonas</i>
			<i>Xanthomonas</i>
	Spoilage	2	<i>Erwinia</i>
			<i>Bacillus spp</i>
			<i>Staphylococcus</i>
			<i>Micrococcus</i>
Aerobic moulds	Epiphytic	3	<i>Streptomyces spp.</i>
			<i>Verticillium</i>
			<i>Acremonium</i>
	Spoilage	4	<i>Fusarium</i>
			<i>Aurebasidium</i>
Yeasts	Spoilage	5	<i>Aspergillus</i>
			<i>Penicillium</i>
			<i>Scopulariopsis</i>
			<i>Wallemia</i>
		6	<i>Mucor spp.</i>
		7	All species

655

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660 Table 4: *Benchmark for microorganisms in feedstuffs (cfu per g feed) according to Kamphues*  
661 *(2013)*

Category <sup>1</sup>	Aerobic bacteria x 10 <sup>6</sup> cfu, g			Moulds x 10 <sup>3</sup> cfu, g			Yeasts x 10 <sup>3</sup> , g
	1	2	3	4	5	6	7
Hay	30	2	0.15	200	100	3	150
Straw	100	2	0.15	200	100	5	400
Grass silage	0.2	0.2	0.01	5	5	5	200
Corn silage	0.4	0.2	0.03	5	5	5	1000

662 <sup>1</sup>*Categories; 1: Epiphytic bacteria; 2 and 3: Spoilage indicating bacteria; 4: Epiphytic moulds; 5*  
663 *and 6: Spoilage indicating moulds; 7: yeasts all species*

664

665

666

667 Table 5 Some suggested forage related risk factors for gastric ulcers and colic (see also Andrews  
 668 *et al.*, 2015, Durham, 2013)

Disease	Risk factor	Possible (non-exclusive) explanation	Ref
Gastric ulcers squamous	Frequency of forage feeding. Intervals > 6h may increase risk of ulcers	Decreased production of saliva, slower passage rate and reduced buffering capacity in the stomach	Luthersson <i>et al.</i> , 2009
Gastric ulcers squamous	Straw as the main forage source in horses/ponies may increase risk of ulcers	Straw may provide low levels of additional buffering support (low in protein, low in calcium). Straw may create mucosal irritation, and may affect the nature of the fibrous mat within the stomach. In addition, the potential for poor hygienic quality (including increased risk of mycotoxins) may play a role.	Luthersson <i>et al.</i> , 2009 Wichert <i>et al.</i> , 2008
Gastric ulcer - glandular	Feeding alfalfa chaff increased the number of glandular ulcers compared to	The glandular lesions could be a result of mechanical injury caused by the very small particles/physical	Fedtke <i>et al.</i> , 2015



hay or alfalfa pellets properties of the alfalfa  
 in weanlings or adult chaff  
 horses

Colic – in Reduced intake of Any change in forage Cohen *et al.*, 2000  
 general grass\*, limited intake intake will cause changes Hassel *et al.*, 2004  
 of forage. Importantly in the microflora in the Hillyer *et al.*, 2002  
 a change in the type hindgut. This may cause Hudson *et al.*, 2001  
 of forage fed. \* NB increased risk of Eg. Change of hay within  
 Horses with dysfermentation, change in two weeks without  
 duodenitis-proximal motility, and several adaptation (Tinker *et al.*,  
 jejunitis were were physiological changes in 1997; Cohen *et al.*,  
 significantly more likely the GI tract. A change in 1999). Change between  
 to have grazed pasture hay within previous 2 two silages with different  
 than the control weeks may increase risk of crude protein content  
 population (Cohen et colic between 4.9 to 10 (Muhonen *et al.*, 2008).  
 al., 2006) times. Change from hay to  
 haylage or silage from the  
 same botanical origin  
 (Muhonen *et al.*, 2009).

Colic - Feeding costal Indigestible fibers have Little and Blikslager 2002  
 impaction Bermuda hay reduced water holding and  
 releasing capacity, can  
 cause changes in the  
 microflora and can cause  
 altered motility

670 **Figures**

671 Figure 1. A very schematic overview giving some definitions and characteristics of hay, haylage  
672 and silage (WSC=water soluble carbohydrates) made without additives. It is important to note  
673 that DM content (and water activity) will determine the potential for fermentation so are linked with  
674 lactate production. .In general as lactate starts to be produced the WSC and pH will start to  
675 reduce, although other factors can influence pH (including buffer capacity). The WSC of any  
676 preserved crop will be lower than the fresh forage it originated from due to respiration (plant and  
677 microbial) post harvesting but the WSC content of haylage compared with the hay from the same  
678 fresh pasture will depend on whether there have been any lactic acid (or VFA production) If there  
679 is virtually no lactic acid (or VFA) production, then there will be no decrease in total WSC in  
680 haylage compared to hay (Muhonen et al 2009).

681  
682 Figure 2. A schematic figure on how minimum required air humidity (relative humidity, RH) for  
683 mould growth varies with ambient temperature (adapted from Lehmann, 1971).

684 Figure 3. The relationship between airtight stored grass forage dry matter (DM) and lactate  
685 content. At DM contents higher than 50 % (dotted line) the lactate content is very low. Data from  
686 Wilkinson *et al.*, 1976 (3 observations, 16- 45%DM); Müller, 2007 (10 observations 29-68%DM);  
687 Muhonen, 2008 (8 observations, 36-81%DM); Ragnarsson, 2009 (8 observations, 36-81%) and  
688 Sarkijärvi *et al.*, 2012 (6 observations, 36-55%DM). All observations are from grass forage  
689 preserved in wrapped bales except for Wilkinson *et al.*, 1976 which were in silos. Forages were  
690 preserved without additives except 6 observations, with lactic acid inoculant, from Muhonen, 2008  
691 and formic acid inoculant in the Sarkijärvi *et al.*, 2012 observations. Data fits ( $R^2=0.84$ ) the  
692 equation:  $y = 0,0406x^2 - 5,4306x + 179.3$  and the equation is relevant in the interval 16-70 % DM.

693 Figure 4. A schematic illustration (not to scale)of the maturation of plants and the effect on dry  
694 matter content (NB in Southern European countries the increase in DM is more dramatic than for  
695 Northern Europe as illustrated by the two lines), digestible plus metabolizable (ME) and Net (NE)  
696 energy plus digestible protein content on a wet matter (WM) basis. The extent of such changes  
697 may be affected by environmental conditions as well as management practices.

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