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Dissecting the genetics of athletic performance and conformation traits in horses

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Dissecting the genetics of athletic performance and conformation traits in horses

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Abstract

The overall aims of this thesis were to investigate the genetics of athletic performance and the level of inbreeding in Coldblooded trotters, as well as the genetics of conformation traits in Icelandic horses. In paper I, genomic-based inbreeding coefficients in Coldblooded trotters were estimated and compared with pedigree-based coefficients. The results showed that the inbreeding level has gradually increased during the investigated years 2000-2009 and that genomic measures of inbreeding could be used to minimize further increased inbreeding. In papers II-IV, different genetic mapping methods were used to dissect the genetics of athletic performance in Coldblooded trotters. The results from papers II and III indicated that athletic performance requires not only physical characteristics but also mental qualities such as memory and learning. In paper IV, we investigated the genetics of athletic performance and exercise related blood pressure. An associated haplotype to these traits was identified. The relative quantitative proteomics analysis showed that proteins linked to inflammatory and coagulation processes were differentially expressed between the elite and sub-elite performing haplotype groups at rest and during exercise.

In paper V, we performed a genome-wide association study in Icelandic horses assessed at breeding field tests to dissect the genetics of conformation of back and croup. A quantitative trait locus for conformation of back and croup and quality of lateral gaits was identified. The genomic region harbored genes linked to anthropometric traits, scoliosis, and motor laterality in humans.

In conclusion, this thesis contributes to increased knowledge about the genetics of complex traits such as athletic performance, blood pressure regulation, inflammation, and anthropometric traits in horses. This information may also be of value for humans and other mammalian species.

Keywords: Athletic performance, Blood pressure, Conformation, Equine, Inbreeding, Inflammation, Lateral gaits

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Sammanfattning

Det övergripande målet med denna avhandling var att undersöka den genetiska bakgrunden till atletisk prestation och inavelsgraden hos kallblodstravare samt den genetiska bakgrunden till exteriöra egenskaper hos islandshästar. I den första studien undersökte vi inavelsgraden hos kallblodstravare genom att använda både information från stamtavlor och molekylär genetisk data. Resultaten visade att inavelskoefficienten gradvis har ökat under de år som studerades (2000-2009) och att genomiska mått på inavelsgrad skulle kunna användas för att förhindra fortsatt ökad inavel. I delarbetena II-IV använde vi olika genetiska kartläggningsmetoder för att undersöka den genetiska bakgrunden till atletisk prestation hos kallblodstravare. Resultaten från delarbetena II och III indikerade att inte bara fysiska egenskaper är viktiga för atletisk prestation utan även mentala egenskaper som minne och inläring. I delarbete IV undersökte vi den genetiska bakgrunden till atletisk prestation och träningsrelaterat blodtryck. En associerad haplotyp identifierades. Den relativa kvantitativa proteomik analysen visade att uttrycket av plasma proteiner kopplade till inflammation och koagulation skilde sig mellan haplotyp grupperna associerad med elit respektive subelit atletisk prestation.

I delarbete V undersöktes genomet hos islandshästar som bedömts vid avelsbedömning för att identifiera den genetiska bakgrunden till exteriör av rygg och kors. En genomisk region identifierades som inte bara påverkade exteriör av rygg och kors utan också kvaliteten av de laterala gångarterna passgång och tölt. Den genomiska regionen innehöll gener som hos människa är kända att vara kopplade till kroppslängd, skolios och motorisk lateralitet.

Sammanfattningsvis bidrar denna avhandling till ökad kunskap om den genetiska bakgrunden till komplexa egenskaper såsom atletisk prestation, blodtrycksreglering, inflammation och antropometriska egenskaper hos hästar. Denna information kan även vara värdefull för människor och andra däggdjursarter.

Nyckelord: Atletisk prestation, Blodtryck, Exteriör, Häst, Inflammation, Inavel, Laterala gångarter

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Preface

“Any living cell carries with it the experiences of a billion years of experimentation by its ancestors.” Max Delbruck, 1949.

Dedication

To my family, friends, and all wonderful horses.

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List of publications

This thesis is based on the work contained in the following papers, referred to by Roman numerals in the text:

- I. Velie BD, Solé M, Jäderkvist Fegraeus K, **Rosengren MK**, Røed KH, Ihler CF, Strand E, Lindgren G. (2019). Genomic measures of inbreeding in the Norwegian–Swedish Coldblooded Trotter and their associations with known QTL for reproduction and health traits. *Genetics Selection Evolution*. 51, 22. <https://doi.org/10.1186/s12711-019-0465-7>
- II. Velie BD, Lillie M, Jäderkvist Fegraeus K, **Rosengren MK**, Solé M, Ihler CF, Strand E, Lindgren G. (2019). Exploring the genetics of trotting racing ability in horses using a unique Nordic horse model. *BMC Genomics*. 20, 104. <https://doi.org/10.1186/s12864-019-5484-9>
- III. Velie BD, Jäderkvist Fegraeus K, Solé M, **Rosengren MK**, Røed KH, Ihler CF, Strand E, Lindgren G. (2018). A genome-wide association study for harness racing success in the Norwegian-Swedish Coldblooded Trotter reveals genes for energy metabolism and learning. *BMC Genetics*. 19, 80. <https://doi.org/10.1186/s12863-018-0670-3>
- IV. **Rosengren MK**, Naboulsi R, Åbrink M, Meadows JRS, Velie BD, Negro JJ, Jouni A, Thorsell A, Sihlbom C, Mattsson CM, Andersson G & Lindgren G. (2022). Genetic regulation of racing performance and exercise related blood pressure in Coldblooded trotters. *Manuscript*

V. **Rosengren MK**, Sigurðardóttir H, Eriksson S, Naboulsi R, Jouni A, Novoa-Bravo M, Albertsdóttir E, Kristjánsson Þ, Rhodin M, Viklund Á, Velie BD, Negro JJ, Solé M, Lindgren G. (2021). A QTL for conformation of back and croup influences lateral gait quality in Icelandic horses. *BMC Genomics*. 22, 267.
<https://doi.org/10.1186/s12864-021-07454-z>

Papers I-III and V are reproduced with the permission of the publishers.

The contribution of Maria Rosengren to the papers included in this thesis was as follows:

- I. Contributed to sample collection. Extracted DNA and prepared samples for array analysis. Read, commented, and approved the final manuscript.
- II. Contributed to sample collection. Extracted DNA and prepared samples for sequencing. Discussed and contributed to data analysis. Read, commented, and approved the final manuscript.
- III. Contributed to sample collection. Extracted DNA and prepared samples for array analysis. Read, commented, and approved the final manuscript.
- IV. Collected samples and measured blood pressure. Performed data analysis, and most of the bioinformatics and whole-genome sequence data analysis. Performed lab work. Extracted DNA, prepared samples for sequencing, performed genotyping, long-range PCR and gel electrophoresis. Interpretation of the results together with the co-authors. Drafted the manuscript.
- V. Collected samples, extracted DNA, and prepared the samples for array analysis. Performed the GWAS and haplotype association analysis. Interpreted the results together with co-authors. Drafted the manuscript together with the co-authors Heiðrún and Marina. Had the responsibility for the final version of the manuscript, submission process, and correspondence with the journal.

Related work by the author

(Not included in the thesis)

Johansson MK, Jäderkvist Fegraeus K, Ekesten B & Lindgren G. The refractive state of the eye in Icelandic horses with the Silver mutation. *BMC Veterinary Research*. 2017,13:153. doi: 10.1186/s12917-017-1059-7.

Jäderkvist Fegraeus K, Lawrence C, Petäjistö K, **Johansson MK**, Wiklund M, Olsson C, Andersson L, Andersson LS, Røed KH, Ihler C-F, Strand E, Lindgren G, Velie BD. Lack of significant associations with early career performance suggest no link between the DMRT3 "Gait Keeper" mutation and precocity in Coldblooded trotters. *PLoS One*. 2017; 12(5): e0177351. doi: 10.1371/journal.pone.0177351.

Velie BD, Smith PM, Fjordbakk CT, Solé M, Jäderkvist Fegraeus K, **Rosengren MK**, Røed KH, Ihler CF, Lindgren G, Strand E. Exploring the genetics underpinning dynamic laryngeal collapse associated with poll flexion in Norwegian-Swedish Coldblooded Trotter racehorses. 2019 Aug 28 *Equine vet J*. doi: 10.1111/evj.13171.

Ghosh S, Davis BW, **Rosengren MK**, Jevit MJ, Castaneda C, Arnold C, Jaxheimer J, Love CC, Varner DD, Lindgren G, Wade CM, Raudsepp T. Characterization of a Homozygous Deletion of Steroid Hormone Biosynthesis Genes in Horse Chromosome 29 as a Risk Factor for Disorders of Sex Development and Reproduction. *Genes* 2020, 11, 251. <https://doi.org/10.3390/genes11030251>

Lindholm J, **Rosengren MK**, Tengvall K, Hansson-Hamlin H, Hanson J. Survival analysis in 51 Standard Poodles diagnosed with naturally occurring hypoadrenocorticism (NOHA). Journal of Veterinary Internal Medicine. Submitted January 2022.

Sample collection (hair, blood, and biopsies), phenotyping, lab work, and data analysis within the ongoing hair growth project where we use the horse as a model to identify genetic factors associated with hair growth.

Abbreviations

ACE	Angiotensin converting enzyme
ChIP-Seq	Chromatin Immunoprecipitation Sequencing
DMRT3	Doublesex and mab-3 related transcription factor 3
DNA	Deoxyribonucleic acid
EBV	Estimated breeding values
ECA	<i>Equus Caballus</i>
EDNRA	Endothelin receptor type A
EDNRB	Endothelin receptor type B
ELISA	Enzyme-Linked Immunosorbent Assay
EMSA	Electrophoretic mobility shift assay
FAANG	Functional Annotation of Animal Genomes
F _{PED}	Inbreeding coefficient based on pedigree data
F _{ROH}	Inbreeding coefficient based on runs of homozygosity
F _{st}	Fixation index
GERP	Genomic Evolutionary Rate Profiling
GSDMC	Gasdermin C
GWAS	Genome-wide association study
HDO	High-Definition Oscillometry
Hi-C	Genome-wide chromosome conformation capture

HiCap	Hi-C with sequence capture
LD	Linkage disequilibrium
MAP	Mean arterial pressure
NGS	Next-generation sequencing
NSCT	Norwegian–Swedish Coldblooded trotter
NSD	North Swedish draught horse
ONT	Oxford Nanopore Technologies
PacBio	Pacific Biosciences
PCR	Polymerase chain reaction
PHYLOP	Phylogenetic P-Values
TBP	TATA-box binding protein
TSS	Transcription start sites
QTL	Quantitative trait locus
RAAS	Renin angiotensin aldosterone system
RNA	Ribonucleic acid
ROH	Runs of homozygosity
SNP	Single nucleotide polymorphism
SORCS3	Sortilin related VPS10 domain containing receptor 3
TBARS	Thiobarbituric Acid Reactive Substances
WGS	Whole-genome sequencing
ZFPM1	Zink finger protein multitype 1

1. Introduction

1.1 History of the horse and domestication

Improved whole-genome sequencing (WGS) techniques have made it possible to analyze ancient DNA from bone and other preserved remains from Equids. The earliest evidence of horse domestication is from about 5500 years ago in the Botai culture located in Kazakhstan. The Botai population was hunting wild horses, but gradually they started to utilize the wild horses for food production and transportation (Outram *et al.* 2009).

The Przewalski horse was previously considered a wild horse and the ancestor of our modern domestic horse (*Equus ferus caballus*). However, by investigating ancient DNA, it has been shown that Przewalski horses originated from the first domestic horses in Botai that returned wild, a process called feralisation. Thus, Przewalski horses are not ancestors to modern domestic horses (Gaunitz *et al.* 2018). Two separate extant horse lines have been found in Iberia (IBE) and Siberia (*E. lenensis*), which did not contribute as ancestors to the modern domestic horse (Fages *et al.* 2019). Recent information from ancient genomes reveals that all modern domestic horse breeds originate from Western Eurasia, mainly from the Volga-Don region of Russia, about 4200 years ago. These horses were strongly selected for weight-bearing and mental traits suited for riding and chariotry (Librado *et al.* 2021). They did not carry the mutation in the *Doublesex and mab-3 related transcription factor 3 (DMRT3)* gene known to play a significant role in gait ability (Andersson *et al.* 2012). However, they differed from the wild horse population at two other loci close to the *Gasdermin C (GSDMC)* gene associated with back pain and the *Zink finger protein multitype 1 (ZFPPI)* gene associated with fear memory and temperament (Aki & Miczek 2014;

Suri *et al.* 2018; Librado *et al.* 2021). These horses expanded and replaced other existing horse populations in Eurasia (Librado *et al.* 2021).

Horses have played an essential role in human history. The horse made it possible for humans to travel more efficiently, which influenced how genes, languages, and diseases were spread worldwide. Horses also affected warfare by making mounted battles possible (Hyland 2003; Drews 2004). Horses were widely used for transportations and as animal power for farming until about the mid-20th century when machinery took over. In some parts of the world, horses are still used for meat production and agriculture (FAO 2021). Today, in the western societies, horses are mainly used for riding and sports, and the horse racing industry comprises multi-billions of euros per year (IFHA 2018). The total number of horses worldwide is estimated at 59 million, and horses still play an important role for many people (FAO 2021). In addition to these domesticated horses, numerous feral horses roam free in different parts of the world, totaling about 600,000 individuals (Rodríguez-Rodríguez *et al in press*).

1.1.1 Horse breeding and selection

Darwin first defined the concept of natural selection in the 19th century. Natural selection means that adaptive traits that make an animal better suited for survival and reproduction within a population get more common over generations (Darwin 1859). However, during the domestication of the horse, people started to breed for desired traits such as endurance, speed, and strength. Different horse breeds have been created with a great variety in size and body type for different purposes of use, such as racing performance in Thoroughbreds (McGivney *et al.* 2020), and riding and gait ability in Icelandic horses (FEIF 2021). Another example is Swedish Warmbloods selected for sports performance specializing in jumping and/or dressage (Viklund *et al.* 2011; Ablondi *et al.* 2019). Today, there are more than 500 registered horse breeds worldwide (FAO 2021). Studbooks with pedigree recording were first established around the late 1700s to keep track of the horse breed populations and the relationship between horses (Weatherby 1791). However, most modern breeds were selected during the 19th century (Negro *et al.* 2021).

Due to the strong selection and limited stallion lines, the genetic diversity in domestic horse breeds has decreased during the last 2000 years (Lindgren *et al.* 2004; Wallner *et al.* 2013; Wutke *et al.* 2018; Fages *et al.* 2019). Genes

linked to behavior, athletic performance, and gait ability have been selected during domestication (Petersen *et al.* 2013a; Librado *et al.* 2016, 2021; Fages *et al.* 2019).

1.1.2 Inbreeding

Breeding with a strong selection for desired traits can lead to inbreeding, especially if the population size is small. Inbreeding means mating between closely related individuals. An increased level of inbreeding has been observed in multiple horse breeds for example Thoroughbreds and Coldblooded trotters. Both these breeds are strongly selected for racing performance, and the population is based on a few popular stallion lines (Arnason *et al.* 1989; Uhlin 2007; Binns *et al.* 2012; McGivney *et al.* 2020; Svensk Travsport 2020). An increased inbreeding level can negatively affect health and racing performance (Klemetsdal 1998; Todd *et al.* 2018; Samsonstuen *et al.* 2020). Inbreeding increases the risk of acquiring rare defects or diseases as a result of increased homozygosity for rare recessive variants. It may also cause a loss of genetic variants that are important (Charlesworth & Charlesworth 1999).

The level of inbreeding can be estimated by studying pedigree data or genomic data based on, for example runs of homozygosity (ROH) (Howard *et al.* 2017). In paper I, both pedigree and genomic data were used to estimate the level of inbreeding in the Coldblooded trotter population.

1.1.3 The horse as a model for genomic studies

Since domestication, the intense selection process in horses has resulted in the accumulation of new mutations linked to different phenotypes. The long history of selection and the population structure with different breeds make the horse and other domestic animals perfect models for studying functional genomics, i.e., the genetic background of different traits (Andersson 2016; Meadows & Lindblad-Toh 2017). Horses are genetically similar to humans, more than half of the chromosomes in horses display conserved synteny with human chromosomes (Wade *et al.* 2009). Many diseases and conditions that affect humans are also present in horses and by using the horse as a model we can get more information about biological mechanisms behind these conditions and thereby improve diagnosis and treatment. Conditions that have been studied in horses and that are also present in humans are for example allergic disorders (Shrestha *et al.* 2015, 2020; Velie *et al.* 2016),

insulin resistance (Lindåse *et al.* 2017; Nostell *et al.* 2019), melanoma (Rosengren Pielberg *et al.* 2008), and asthma (Couetil *et al.* 2020). High blood pressure is a common condition in humans (WHO 2021). In paper IV, we investigated the genetics of athletic performance and exercise related blood pressure regulation. Information from this study may also be of use in humans. The horse can serve as a model for athletic performance and biological processes activated by exercise. As well as elite human athletes, racehorses need to have the optimal genetic setup for athleticism to tolerate training and perform at an elite level to win races. This thesis focused on two horse breeds selected for sports performance; Coldblooded trotters and Icelandic horses.

1.1.4 Coldblooded trotters

Coldblooded trotters originate from the Norwegian Döle horse and North Swedish draught horses (Bjørnstad & Røed 2001; Uhlin 2007). The studbook for Coldblooded trotters was founded in 1965. Sweden and Norway have had a common breeding program since 2000. The overall breeding goal is to preserve and further develop a competitive, healthy trotting horse by improving the horses' competitive performance, durability, conformation, and temperament. The aim is also to prevent inbreeding. Stallions are judged at breeding evaluation and estimated breeding values (EBV) based on racing performance records are applied by the breeders when selecting horses for breeding (Uhlin 2007; Svensk Travsport 2021a). The population size is limited, and the Coldblooded trotter is included on the Swedish Board of Agriculture list of breeds that should be preserved (Jordbruksverket). Today, there are about 12 000 Coldblooded trotters in Sweden (Svensk Travsport 2021b). In paper I we investigated the level of inbreeding in Coldblooded trotters born in Sweden and Norway. The first harness races started in the 1500s and have since been developed with separate races for Coldblooded trotters and Standardbreds (Uhlin 2007). Today, the horse racing industry in Sweden includes multi-billions of SEK (Svensk Travsport 2019).

Since around the 1950s, Coldblooded trotters have been strongly selected for harness racing performance. Their racing speed has improved remarkably, and the horses are more lightly built than they were in the past when they were used for agriculture and forestry (Arnason *et al.* 1989; Uhlin 2007) (Figure 1). Coldblooded trotters were used as a unique model in papers II-IV to investigate genetic factors associated with athletic performance.



Figure 1. Coldblooded trotter on the harness racing track. Photo: Matilda Persson

1.1.5 Icelandic horses

The ancestors of Icelandic horses were brought to Iceland by the Vikings during the settlement of the island between the 9th and 10th centuries (Adalsteinsson 1981). The Icelandic horse breed originates mainly from Norway and the British Isles (Bjørnstad & Røed 2001). The Icelandic horse is the only horse breed allowed in Iceland and it is considered a pure breed, as it has been isolated for more than 1000 years with no genetic influence from other horse breeds (Figure 2). Originally, Icelandic horses were mainly used for transportation. Today, the Icelandic horse is a popular breed used for hobby riding and gait competitions in more than 20 countries worldwide. There are more than 280 000 Icelandic horses worldwide (WorldFengur 2021).

A selection process of the horses started already at the settlement of the island, as only the best horses were brought to Iceland. The first breeding field test was held in Iceland in 1906. However, it was not until 1950 that subjective scoring was established. Icelandic horses can be scored at breeding field tests for conformation and riding performance traits (FEIF 2021). Estimated breeding values and records from breeding field tests are applied to select horses for breeding.

The overall breeding goal is to breed healthy, reproductive, and durable horses. The aim is also to preserve coat color variation in the breed (FEIF 2021). The great phenotypic variation within this purebred breed and the standardized evaluation records from breeding field tests are valuable for genomic studies. In paper V we used phenotypic data from breeding field tests to identify genetic factors associated with conformation of back and croup in Icelandic horses, which is known to play a major role on sports performance (Albertsdóttir *et al.* 2008; Kristjánsson *et al.* 2016).

Icelandic horses can perform the alternate gaits tölt and pace. Tölt is a four beat lateral gait and the most important evaluated trait at the breeding field tests. Ideally, the movement at tölt should be fluid and balanced with a light front part and a strong back. Some Icelandic horses are five gaited and can perform pace, a two beat lateral gait. At pace, the horses should have a good speed and a long stride length (FEIF 2021). A loss of function mutation in the *DMRT3* gene has been shown to play a major role on the ability to perform lateral gaits (Andersson *et al.* 2012).



Figure 2. Icelandic horses on pasture in Iceland. Photo: Maria Rosengren

1.2 Genomics and trait mapping

The deoxyribonucleic acid (DNA) carries the genetic code for proteins, which are the molecular machinery for life and the system that controls the activity of the genes (Wold *et al.* 2014). Genomics means that the whole genome of an individual is investigated to get information about the structure and function and thereby link genetic factors to the studied traits. Within this thesis, we used different trait mapping methodologies to identify genetic variants associated with athletic performance and conformation traits in horses.

1.2.1 The horse genome

The first draft of the reference genome in horses was completed in 2007, followed by a second version (EquCab2) released in 2009. The reference genome was based on one Thoroughbred mare named Twilight (Wade *et al.* 2009). This horse was selected because it had low level of heterozygosity (i.e., low genetic variability). The whole horse genome contains 2.5-2.7 Giga bases and about 20 322 protein coding genes. More than half of the horse chromosomes show conserved blocks of genes to human chromosomes (Wade *et al.* 2009). An updated horse reference genome (EquCab3) was released in January 2018 (Kalbfleisch *et al.* 2018).

1.2.2 Genome-wide association studies

Genome-wide association study (GWAS) is a method for identifying associations between a particular trait and genetic variants in the DNA sequence. This trait mapping method was used in papers III and V. Traditionally, single nucleotide polymorphisms (SNPs) are used as genetic variants in GWA studies. A SNP is a difference at a single nucleotide position in the DNA sequence between individuals (Posthuma; Bush & Moore 2012). Shortly after the release of the first reference genome, the first equine genotyping array was developed. An improved high-density array was created in 2012 based on different breeds and related equids (McCue *et al.* 2012). In 2017, a next-generation high density genotyping 670k array became available, containing more markers evenly distributed across the genome (Schaefer *et al.* 2017).

GWAS can be performed as a quantitative trait and/or a case control study. Careful phenotyping is of importance and the individuals should be as

unrelated as possible to avoid false similarities that are not related to the studied trait. The relationship should at least be the same between the groups (Marchini *et al.* 2004). All samples are then genotyped for variants on the array, and quality control of the genotype data is performed. Statistical analyses are implemented to look for genotype differences between different phenotype groups. The candidate mutation/s can be located where there is a difference in allele frequency between the different phenotypes or in linkage disequilibrium (LD) with the detected significant variant/s (Posthuma; Bush & Moore 2012).

1.2.3 Selective sweep mapping

If there is a strong selection for a particular trait in a population, the genetic variation in that region decreases. There will be more homozygosity in the selected region compared to the original ancestral population. These footprints of selection can be used to detect genomic regions under selection by performing fixation index (F_{st}) analysis (Petersen *et al.* 2013b; Gurgul *et al.* 2019; Ardestani *et al.* 2020). Since domestication, horses have been selected for desired traits such as strength, gaits, speed, and endurance (Petersen *et al.* 2013b). Coldblooded trotters originate from the North Swedish draught horse used for agriculture and forestry but have been selected for more than 60 years for athletic performance. Before parental testing was introduced in the 1960s, Coldblooded trotters were to some extent crossbred with Standardbreds to further improve their athletic performance (Uhlin 2007). This makes the Coldblooded trotter a perfect model for studying athletic performance. Selective sweep mapping with F_{st} analysis was used in paper II by comparing pooled WGS data of Coldblooded trotters, North Swedish draught horses, and Standardbreds.

1.2.4 Whole-genome sequencing

Whole-genome sequencing (WGS) is as the name implies, a method that analyses the nucleotide sequence of an individual's whole genome. The major advance in sequencing technology came in the 1970s when Frederick Sanger and his team developed the chain-terminating inhibitor method (Sanger *et al.* 1977). Today, Sanger sequencing is still used, mainly when shorter parts of the genome are investigated. We used the Sanger sequencing technique in paper IV to verify target regions of the long-range PCR products and identify the breakpoints of a potential duplication detected from WGS.

The sequencing techniques have over the years been greatly improved. Next-generation sequencing (NGS) became available in 2005, which generates a lot of sequence data fast and cost effectively. With NGS, the DNA molecules are fragmented and sequenced in parallel with a method called sequencing by synthesis. Fluorescently tagged nucleotides are incorporated with the sequenced DNA molecule as a template. Currently, Illumina-based NGS technology is one of the most commonly used NGS methods (Heather & Chain 2016). We applied WGS using Illumina technology in papers II and IV to identify genetic variants associated with athletic performance.

Long-read sequencing is the third generation of sequencing where the whole DNA molecule can be sequenced at once without fragmentation. This increases the chance to detect larger structural variants (Mantere *et al.* 2019; Begum *et al.* 2021). In paper IV, we performed targeted long-read sequencing using MinION Oxford Nanopore Technologies (ONT) (Clarke *et al.* 2009). Another platform that provides long-read sequencing is the Pacific Biosciences (PacBio) (Eid *et al.* 2009). PacBio has a lower error rate than ONT but is more expensive, and ONT gives a higher yield (Weirather *et al.* 2017)

1.2.5 Fine-mapping and identification of candidate causal genetic variants

Trait mapping methods usually generate large genomic regions with many variants that need to be ranked and selected for functional validation to finally identify the causative variant or variants.

In general, candidate variants are prioritized based on genotyping quality, association with the studied trait, and if they are predicted to affect the encoded protein product. Different variant annotation tools can be used for predicting the effect of the variant, for example, the Ensembl Variant Effect Predictor (VEP) (McClaren *et al.* 2016) and ANNOVAR (Wang *et al.* 2010). If the identified variants are located in non-coding regions, they may regulate gene expression. Genetic variants can have a potential regulatory role, for example, if they overlap predicted regulatory elements, antisense transcripts, transcription factor binding sites, or histone marks indicating regulatory elements. Genomic conservation is an important factor when prioritizing variants. If the sequence is well conserved between species, it likely has an important function. Different tools can be used for predicting conservation, such as Phylogenetic P-Values (PHYLOP), and Genomic Evolutionary Rate

Profiling (GERP) scores (Jalali Sefid Dashti & Gamiieldien 2017). Recently, the Zoonomia project published a powerful resource containing genome assemblies of 240 different mammals (Zoonomia Consortium, 2020).

1.2.6 Functional genomics

To verify the effect of a candidate causative mutation, it needs to be validated in additional individuals and evaluated with functional tests. The equine Functional Annotation of Animal Genomes (FAANG) group has performed Ribonucleic acid (RNA) sequencing to get information about gene expression in different tissues and Chromatin Immunoprecipitation Sequencing (ChIP-Seq) to identify signatures of regulatory elements within the horse genome (Burns *et al.* 2018; Kingsley *et al.* 2020, 2021). Variants within non-coding regions can affect gene expression, for example, if they change the chromatin structure, overlap transcription factor binding sites located in transcriptional regulatory elements such as promoters, enhancers, and locus control regions (Jalali Sefid Dashti & Gamiieldien 2017). Promoters are located 5' of transcription start sites (TSS) and divided into core and regulatory promoter. The core promoter defines the TSS and binds general transcription factors such as TFIID with TATA-box binding protein (TBP) and TBP-associated proteins. The regulatory promoter binds a multitude of transcriptional activators and repressors (Nakatani *et al.* 1990; Haberle & Stark 2018). In contrast to promoters, enhancers can be located far away (> 1 Mbp), either 5' or 3' from their target gene/s as distant genomic regions can be brought together by looping (Müller *et al.* 1989; Krivega & Dean 2012; van der Valk *et al.* 2015). Chromatin interaction analysis is an informative method that can be applied to identify candidate causative mutations and their target genes (Lieberman-Aiden *et al.* 2009; Sahlén *et al.* 2015, 2021; Roytman *et al.* 2018). Genome-wide chromosome conformation capture (Hi-C) is a chromatin interaction analysis method that applies restriction enzymes and NGS to map looping structures of the genome (Lieberman-Aiden *et al.* 2009). Recently, HiCap (Hi-C with sequence capture) was developed. HiCap is an improved chromatin interactions mapping method that can map individual enhancer interactions with known promoters in the whole genome (Sahlén *et al.* 2015).

Protein interactions with the DNA sequence can be experimentally tested *in vitro* with electrophoretic mobility shift assay (EMSA) (Fried & Crothers 1981; Hellman & Fried 2007). For example, EMSA can investigate genetic

variants ability to bind both known DNA binding transcription factors as well as hitherto unknown transcription factors. Luciferase assays can be used to test if a genomic sequence regulates transcription of a candidate gene. A construct with the sequence of interest and regulatory region of the candidate gene is inserted in front of the reporter gene Luciferase, which emits fluorescent light when activated (Smale 2010).

Functional validation of potential causative mutations is performed using genome editing techniques, such as CRISPR-Cas9, in animal models or cell lines (Jiang & Doudna 2017).

1.2.7 Protein analysis

Proteins are the products of protein encoding genes. The proteome is the total number of proteins expressed in an organism or a specified organ, tissue, or cell type, and proteomics means that the proteome is studied. Proteomics is a valuable component of functional genomics to understand the structure, interactions, and functions of proteins. The concentration of proteins changes with different conditions, and proteins can be modified after translation which can affect their function (Tyers & Mann 2003). Quantification and differences in protein levels between biological samples or cell types can be analyzed with different techniques, such as Mass spectrometry, Enzyme-Linked Immunosorbent Assay (ELISA), and Western blot (Patterson & Aebersold 1995; Blackstock & Weir 1999; Mathesius *et al.* 2003).

In paper IV, we used Relative Quantitative Mass Spectrometry with tandem mass tags and ELISA to investigate differences in plasma protein levels between the different haplotype groups, at rest and during exercise. ELISA is a technique to quantify specific proteins by using antibodies against the protein of interest (Alhadjj & Farhana 2021). Relative Quantitative Mass Spectrometry with tandem mass tags can compare differences in the proteome between multiple samples. The proteins within the sample are digested into peptides and labeled with specific isobaric tags. The samples are then fractionated in a mass spectrometer and the peptides can be detected and quantified by a reporter ion within the isobaric tag. The fragments and peptides are then matched using databases with known peptide sequences to identify proteins (Zhang & Elias 2017).

1.3 Athletic performance

Athletic performance is a complex trait influenced by environmental, genetic, and epigenetic factors. The horses need to have the correct phenotypic characteristics to function successfully for the desired purpose on the race track as well as for leisure riding. Genes linked to muscle mass and locomotion patterns have previously been reported to have a major impact on athletic performance in horses, for example, the *Myostatin* gene, which encodes a protein belonging to the transforming growth factor β family of growth factors that regulate skeletal muscle mass (McPherron *et al.* 1997). *Myostatin* is associated with conformation, racing performance, and endurance (Hill *et al.* 2010; Tozaki *et al.* 2011; McGivney *et al.* 2012; François *et al.* 2016). The *DMRT3* gene encodes a transcription factor that affects locomotion patterns and plays a major role in lateral gait ability, riding traits, and harness racing performance (Andersson *et al.* 2012; Jäderkvist *et al.* 2014; Kristjánsson 2014; Jäderkvist Fegraeus *et al.* 2015; Jäderkvist *et al.* 2015).

In papers II and III, the horse genome was investigated to identify genetic variants associated with athletic performance. In paper IV we investigated genetic regulation of athletic performance and exercise related blood pressure. During exercise, the blood pressure needs to be highly regulated to supply the working muscles with energy and oxygen (Erickson 1993; Rowell 1993).

1.3.1 Blood pressure regulation

Horses have a heart rate of about 28-40 beats per minute at rest (Erickson & Poole 2004). Intense exercise activates the sympathetic system with the “fight or flight” response to obtain more energy and oxygen for the working muscles. β -adrenoreceptors in the cardiac muscle respond by increasing heart rate and the contractility of the heart so that the blood is circulated more effectively (Rose & Evans 1987). During exercise, the heart rate rises to about 180-240 beats per minute. Cardiac output is defined as the blood volume pumped out from the heart per minute. During exercise, the cardiac output is increased up to 400 liters per minute to supply the working muscles with oxygen and energy (Poole & Erickson 2004). Blood pressure is defined as the pressure of the blood pumped against the vascular walls, and it is dependent on the cardiac output and the total resistance in the peripheral blood vessels. The diameter of the arteries and the viscosity of the blood

affect the total peripheral resistance. The blood flows more efficiently with less turbulence if the arteries are wide and if the blood is less viscous (Poole & Erickson 2004). During exercise, more blood needs to be distributed to the working muscles. The blood flow to skeletal musculature can increase during exercise more than 60 times compared to rest. This redistribution is mainly achieved by regional vasodilation in the working muscles and vasoconstriction to the, for exercise, non-essential tissues, i.e., the kidneys and intestines. Moreover, an increased blood volume is needed during work for sweating and temperature regulation. All these factors are essential for athletic performance and are highly regulated (Erickson 1993; Rowell 1993).

The Vasoactive peptide Endothelin and nitrogen oxide

The peripheral resistance is mainly regulated by dilation or constriction of the blood vessels. Endothelins regulate the diameter of blood vessels (Yanagisawa *et al.* 1988). There are three isopeptides of Endothelin; Endothelin 1 (EDN1), Endothelin 2 (EDN2), and Endothelin 3 (EDN3). Separate genes encode each of them. Endothelin binds to specific G protein coupled receptors which are comprised of two main subtypes, Endothelin receptor type A (EDNRA) and Endothelin receptor type B (EDNRB). EDNRA has a high specificity for EDN1 and EDN2 but a low affinity for EDN3 (Arai *et al.* 1990). EDNRB has the same affinity for all three Endothelin types (Sakurai *et al.* 1990). Endothelins act regionally and the effect of the receptor subtypes varies depending on the tissue type. In general, EDNRA activation results in constriction of the blood vessels and increased blood pressure. In contrast, stimulation of EDNRB results in vasodilation by nitric oxide secretion and thus a decreased blood pressure. By regulating the diameter of the blood vessels locally, EDN1 and EDN3 contribute to the redistribution of the blood that is essential during exercise, i.e. dilatation in the working muscles and constriction to the non-essential tissues (Weitzberg *et al.* 1995; Maeda *et al.* 2002). EDN3 stimulates the release of vasopressin, which increases the blood volume by saving water and sodium in the kidneys. Vasopressin also acts as a vasoconstrictor (Rossi 1993, 1995).

Antidiuretic hormone (vasopressin)

Vasopressin is a hormone released from the pituitary gland to regulate the blood volume and salt balance in the body. Vasopressin is secreted in response to a decreased blood volume reported by the baroreceptors in the heart and lungs, decreased arterial blood pressure reported by baroreceptors

in the arteries, and by osmolality differences of the plasma sensed by the cells in the hypothalamus (Wade & Freund 1990). In addition, EDN3 can stimulate the secretion of vasopressin (Rossi 1993, 1995). Vasopressin constricts arteries, saves water in the kidneys, and increases water and sodium absorption in the colon (Wade & Freund 1990; Mckeever *et al.* 1991). Vasopressin secretion results in increased blood volume, making the blood less viscous so it can flow more easily to supply the working muscles with oxygen and nutrients. A larger blood volume is also needed for thermoregulation during exercise (McKeever & Hinchcliff 1995). Plasma concentration of vasopressin increases linearly with increasing exercise intensity in horses and humans (Wade & Freund 1990; Mckeever *et al.* 1991).

The renin angiotensin aldosterone system (RAAS)

Renin is a hormone secreted from the juxtaglomerular part of the kidneys. Renin stimulates the transformation of angiotensinogen into angiotensin I, which is then converted into angiotensin II by angiotensin converting enzyme (ACE). Angiotensin II is a potent vasoconstrictor that also triggers the secretion of vasopressin and aldosterone, which both acts on the kidneys to save sodium and water (Wade & Freund 1990). Plasma concentration of renin increases with exercise and is correlated with the intensity of the work in horses (McKeever *et al.* 1992). The renin angiotensin aldosterone system (RAAS) and Endothelin are two of the most powerful vasoregulatory systems. There are many interactions between these two systems that affect blood pressure (Rossi *et al.* 1999). Angiotensin II can activate the transcription of EDN1 (Chua *et al.* 1993; Sung *et al.* 1994). On the other hand, EDN1 can also activate the synthesis of angiotensin II probably by enhanced action of ACE (Kawaguchi *et al.* 1990, 1991). Another example of interaction between the two systems is that EDN1 can activate aldosterone secretion (Mazzocchi *et al.* 1990, 1992).

1.3.2 Blood pressure measurement

Blood pressure can be measured by using invasive or non-invasive methods. With the invasive method, a catheter is placed within an artery. An electronic pressure transducer, either located at the end of the catheter or connected via an external system, continuously measures the blood pressure. This method is more accurate but is invasive and thus inflicts pain and a risk of infection

(Erhardt *et al.* 2007). Doppler or oscillometric techniques are most commonly used as non-invasive methods. Oscillometry is a technique where the mean arterial pressure (MAP) is measured when the largest pressure fluctuations (oscillations) in the artery wall are detected. In horses, non-invasive blood pressure is usually measured by placing a cuff around the base of the tail over the middle coccygeal artery. An updated oscillometry method, High-Definition Oscillometry (HDO), was developed in 2006 and HDO has been used in many different species, including horses (Söder *et al.* 2012; Nostell *et al.* 2016, 2021). With HDO, the deflation rate can be adjusted and pulse waves analyzed in real time in the MDSWIN Analyse Software. In addition, HDO gives information on parameters such as systemic vascular resistance, stroke volume, and arrhythmia. We used the oscillometry method to measure blood pressure in paper IV.

1.4 Conformation of back and croup and gait quality

The back of the horse has a central role in the musculoskeletal system. The back plays a major role in the locomotion of the horse where the spine act as a spring in the stride cycle. The structure of the spine is complex and has two major functions; to support the body weight and to transfer the force from the hind legs in the forward movement (Slijper 1946). Back problems can cause pain and impaired performance. In general, back problems are hard to diagnose clinically in horses because the strong muscles and ligaments of the back can mask mild clinical signs (Jeffcott 1980).

It is well known that conformation traits are important for the health of the musculoskeletal system and sports performance in horses (Albertsdóttir *et al.* 2008; Rustin *et al.* 2009; Holmström & Back 2013; Solé *et al.* 2013; Jönsson *et al.* 2014a; b; Kristjánsson *et al.* 2016; Sánchez-Guerrero *et al.* 2016; Janczarek *et al.* 2017; Sánchez-Guerrero *et al.* 2017). Traditionally, conformation traits are used in multiple horse breeds when horses are selected for breeding and performance. In Coldblooded trotters, the stallions are assessed for conformation traits at breeding evaluation (Svensk Travsport 2021a). Icelandic horses are assessed at breeding field tests and the evaluation includes both conformation and riding ability traits (FEIF 2021).

An uphill conformation (higher front part of the horse compared to hind), croup dimensions, and chest width are known to be the most important qualities for riding ability in Icelandic horses (Albertsdóttir *et al.* 2008;

Kristjansson *et al.* 2016). The uphill conformation is subjectively assessed using for reference the difference between measurements of the height of the withers and the height at the croup. This should ideally be accompanied by a well-balanced backline and an even trunk with a breast that is not too deep. The backline is defined as the line from withers back to the lumbosacral joint. It is subjectively assessed based on the difference between the measurements of the lowest point of the back and the height of the croup as reference. The lowest point of the back should ideally be located in the middle of the back (FEIF 2021).

In paper V, we investigated the genetics underlying the conformation of back and croup and how this trait affects gait performance in Icelandic horses. The score given at breeding field tests for back and croup is a subjective comprised score that involves both skeletal and muscular qualities. High scores of back and croup are associated with a well-balanced backline (i.e. not forward sloping or swayback), broad and well-muscled back as well as a long, even, and well-muscled croup. On the other hand, low scores for back and croup are associated with a forward sloping or stiff backline, a narrow poor muscled back, a short or too long uneven croup lacking muscles (FEIF 2021) (Figure 3).

Gait qualities such as collection, ease, suppleness, speed, stride length, and action of the legs are highly valued when the gaits are assessed at the breeding field tests (FEIF 2021).



Figure 3. Icelandic horses that represent high and low scores of back and croup, respectively. A. Icelandic horse representing high score for back and croup. The backline is well balanced, and the croup is long and well-muscled. B. Icelandic horse representing low score for back and croup. The back is forward sloping, and the croup is short and lacks muscles. Photo: The Swedish Icelandic horse breeding organization, SIF Avel

2. Aims of the thesis

The overall aims of the thesis were to identify genetic variants associated with athletic performance and conformation traits in horses, as well as to investigate the level of inbreeding in Coldblooded trotters.

The specific aims were to:

- Estimate genomic inbreeding coefficient for Coldblooded trotters and compare with the inbreeding level based on pedigree data in the whole population.
- Identify genetic variants associated with athletic performance in Coldblooded trotters and investigate their potential effect on exercise related blood pressure regulation.
- Identify genomic regions linked to conformation of back and croup in Icelandic horses and investigate their effects on riding performance traits assessed at breeding field tests.

3. Summary of studies (I-V)

This thesis consists of five studies with the aims to investigate the inbreeding level in Coldblooded trotters and identify genetic variants associated with athletic performance and conformation traits in horses.

3.1 Study I – Genomic measures of inbreeding in the Norwegian–Swedish Coldblooded Trotter and their associations with known QTL for reproduction and health traits

The inbreeding level in Coldblooded trotters, also known as Norwegian–Swedish Coldblooded trotters (NSCT), was estimated using both pedigree data and high-density genotyping array data in 566 raced horses born between 2000 and 2009. Of the 566 horses, 265 were born in Norway and 301 in Sweden. NSCT has been strongly selected for athletic performance for more than sixty years, and their racing performance has greatly improved. However, this strong selection has led to an increased inbreeding level, which can negatively affect health and athletic performance (Klemetsdal 1998; Todd *et al.* 2018; Samsonstuen *et al.* 2020). The aims of study I were to estimate genomic inbreeding coefficient for NSCT and compare it with the inbreeding level based on pedigree data in the whole NSCT population.

Results and discussion

The inbreeding level had gradually increased in NSCT during the investigated years 2000 to 2009. The average inbreeding coefficient based on genomic data (F_{ROH}) ranged from 1.78 to 13.95%. A greater increase could be observed in horses born after 2005. F_{ROH} was higher than the inbreeding coefficient based on pedigree data (F_{PED}), however, both methods showed a progressively increased level of inbreeding. F_{ROH} increased by 3.15% and F_{PED} by 1.48% (Figure 4).

Different thresholds of runs of homozygosity (ROH) were tested. The threshold that resulted in the highest correlation ($R=0.5629$) between F_{ROH} and F_{PED} was selected to detect longer homozygous regions that were represented in the population.

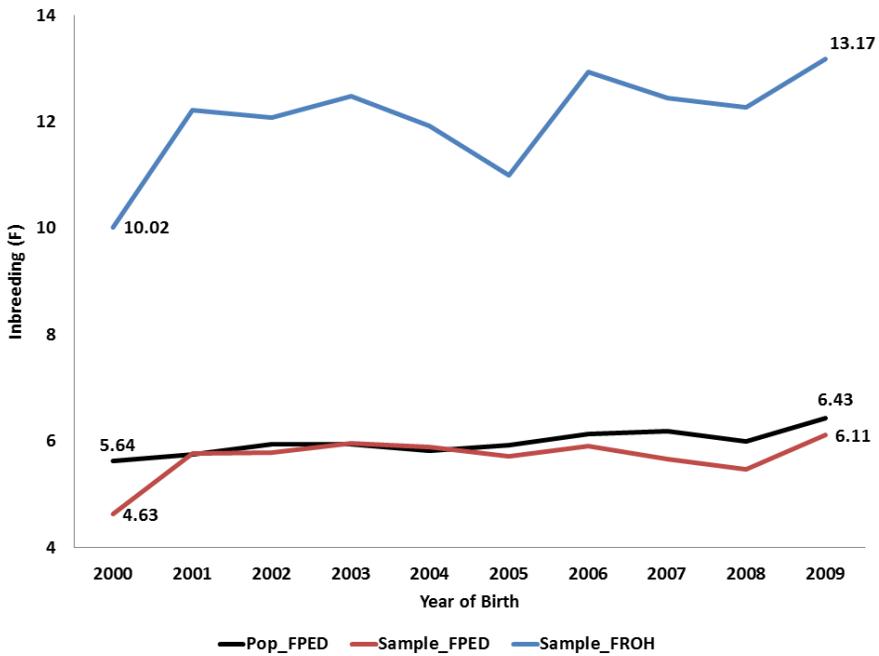


Figure 4. Median inbreeding coefficients in Norwegian–Swedish Coldblooded trotter (NSCT) by birth year, from 2000 to 2009. Sample_FROH: inbreeding based on genomic data for all the 566 studied horses. Pop_FPED: inbreeding based on pedigree data for the whole population (n=14 547). Sample_FROH: Genomic inbreeding based on genomic data in the 566 studied horses (Velie et al. 2019b).

In total, 1403 chromosomal regions of ROH were identified that were common in at least 95% of the 566 studied horses. Chromosomes 31 and 1 had the largest and smallest proportion of ROH, respectively (Figure 5).

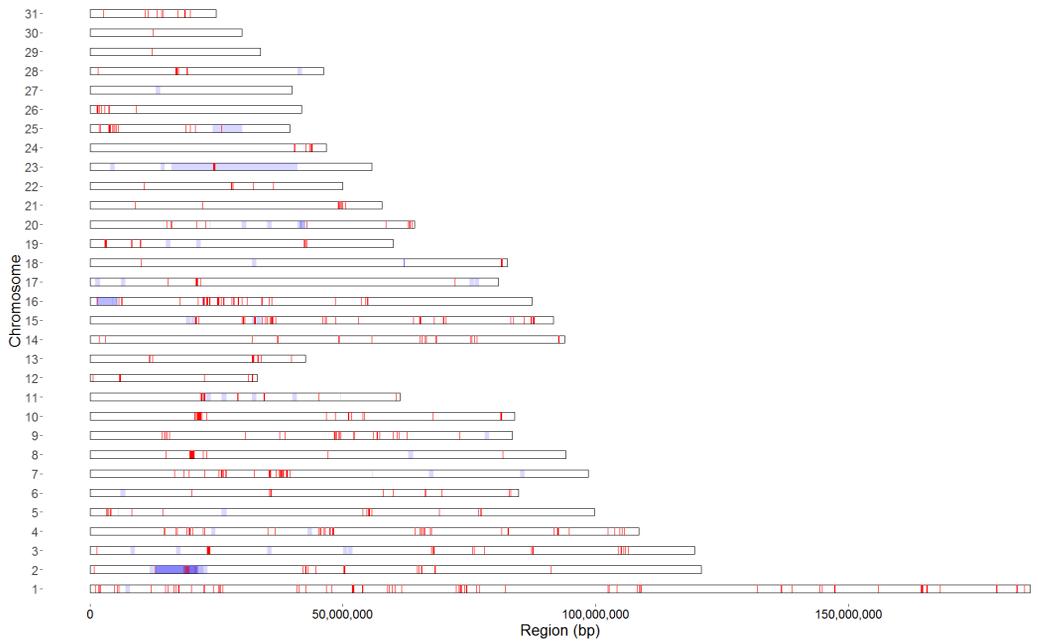


Figure 5. Chromosomal positions of identified runs of homozygosity in the studied Norwegian–Swedish Coldblooded trotter population marked in red. Regions marked in blue overlapped previously identified quantitative trait loci for reproduction and health (darker blue as the number of overlapped QTL increases). (Velie et al. 2019b).

The ROH regions that were common in more than 95% of the studied NSCT were compared to known QTL for health and reproduction. Of 1403 identified ROH regions, 38 overlapped known QTL for health traits. The majority involved QTL linked to osteochondrosis and the muscle disorder recurrent exertional rhabdomyolysis. Only one ROH overlapped a QTL associated with reproduction, which indicates that reproduction is currently not a major problem in NSCT. However, if the level of inbreeding continues to increase, health and reproduction problems are likely to occur.

F_{PED} and estimated breeding values are already available for owners and breeders when selecting horses for breeding. Genomic data on regions sensitive for inbreeding could further aid in the selection to preserve genetic diversity.

3.2 Study II – Exploring the genetics of trotting racing ability in horses using a unique Nordic horse model

In study II, Coldblooded trotters were used as a unique model to identify genomic regions associated with harness racing performance. Coldblooded trotters are derived from the North Swedish draught horse (NSD), a breed used for agriculture and forestry. To further improve the racing performance, Coldblooded trotters were to some extent crossbred with Standardbreds before 1969 when parentage testing was introduced (Uhlen 2007). This might have introduced genetic variants favorable for racing performance. To identify these genomic regions, pooled whole-genome sequence data from 25 North Swedish draught horses, 18 Coldblooded trotters, and 22 Standardbreds were compared using selective sweep mapping with fixation index (F_{st}) analysis. The horses were carefully selected based on estimated breeding values for athletic performance and relationship. Candidate regions were defined where Coldblooded trotters were similar to Standardbreds and at the same time different from North Swedish draught horses. F_{st} cut off was set at ($> 95\%$ percentile, $F_{ST} = 0.179$ NSCT vs. NSD; $< 5\%$ percentile, $F_{ST} = 0.013$ NSCT vs. SB). Regions that were smaller than 1000 bp or contained less than two SNPs were filtered out. Candidate regions were screened in the Ensembl Biomart function and the generated list of genes was analyzed in PANTHER to get information on the molecular functions and biological processes.

Results and discussion

The selective sweep mapping resulted in 580 genomic candidate regions for trotting racing performance. Chromosomes 7 and 11 had the highest number of SNPs (214 and 147 SNPs, respectively). Many of the candidate genes were linked to metabolic and cellular processes (Figure 6). Biological processes of the candidate genes included mainly catalytic and binding activity (Figure 7).

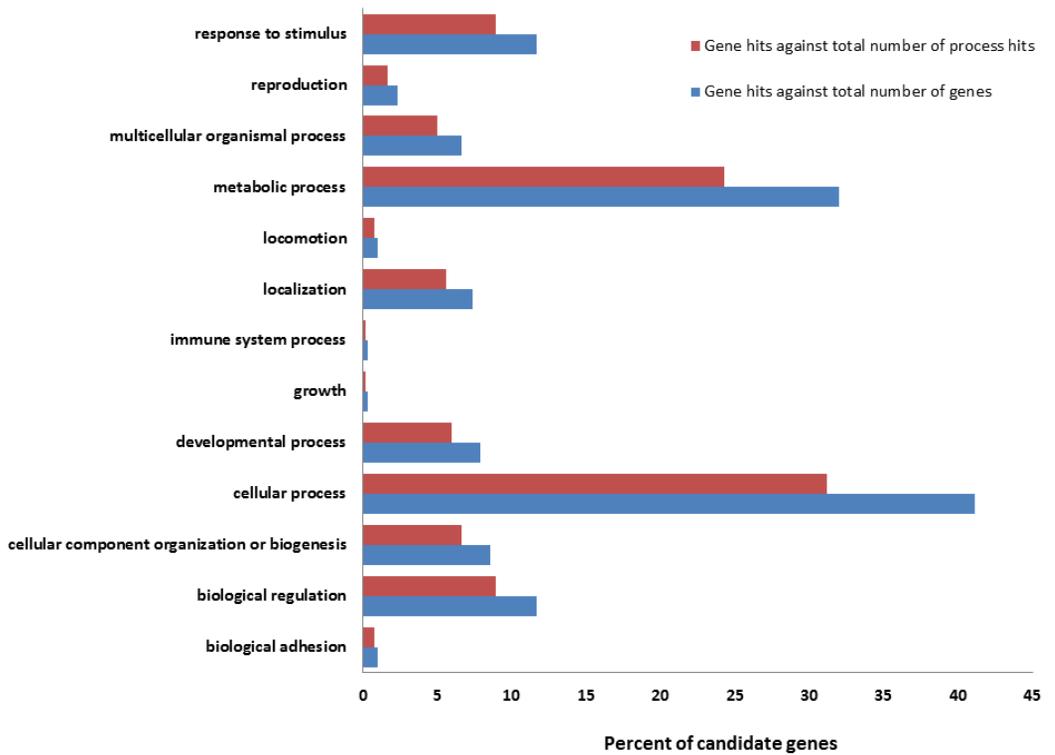


Figure 6. Molecular functions of the candidate genes analyzed in PANTHER. Definition of the classification in PANTHER: the function of the protein by itself or with directly interacting proteins at a biochemical level (Velie et al. 2019a).

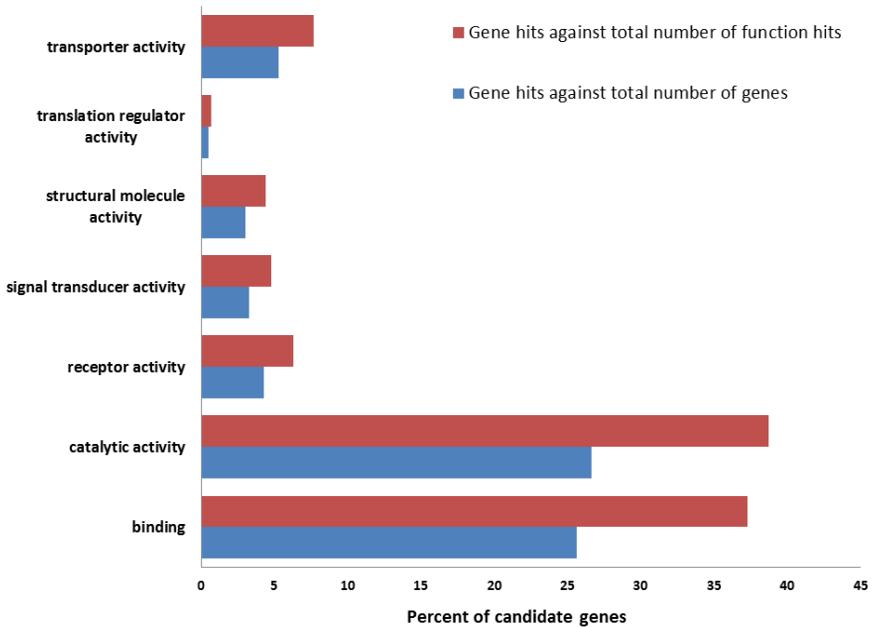


Figure 7. Biological processes of the candidate genes analyzed in PANTHER. Definition of the classification in PANTHER: the function of the protein in the context of a larger network of proteins that interact to accomplish a process at the level of the cell or organism (Velie et al. 2019a).

One of the identified regions overlapped a QTL previously known to be associated with endurance racing speed in horses (Ricard *et al.* 2017). The region was located on chromosome 1 and harbored the gene *Sortilin related VPS10 domain containing receptor 3 (SORCS3)* linked to spatial memory and learning (Breiderhoff *et al.* 2013).

Harness racing performance is a complex polygenic trait. Hundreds of candidate genes were identified in our study. The genes were linked to musculature, conformation and, also neurological processes. Indicating that not only physical qualities are important for harness racing performance, but also mental qualities such as learning and intelligence.

3.3 Study III – A genome-wide association study for harness racing success in the Norwegian-Swedish Coldblooded Trotter reveals genes for energy metabolism and learning

In study III, we performed genome-wide association analyses to detect genetic variants associated with athletic performance in 613 Coldblooded trotters. The horses were randomly selected to represent the raced population and genotyped on the High-density 670K Axiom Equine Genotyping Array. Three performance traits were analyzed; earned prize money, best km time and, number of gallops. Candidate genes linked to energy metabolism, intelligence, and the immune system were identified.

Results and discussion

Nine genome-wide significant SNPs and 23 suggestive SNPs associated with athletic performance were identified. The identified SNPs were located on *Equus caballus* chromosome (ECA) 1, 2, 6, 7, 16, 17, 23, 25, 28, 29 and 31 (Figure 8-10). In total, eight candidate genes for athletic performance were identified. One of the genes with the clearest association with performance was the *Dedicator of cytokinesis 8 (DOCK8)* gene on ECA23. Five suggestive SNPs were associated with best km time. Four of the SNPs were located within introns and the other SNP about 5.7 kb from the *DOCK8* gene. *DOCK8* can be linked to the immune system, intelligence, and motor skills (Griggs *et al.* 2008; Qin *et al.* 2015). *DOCK8* is located on the same chromosome as the *Doublesex and mab-3 related transcription factor 3 (DMRT3)* gene, which plays a major role in locomotion patterns and athletic performance (Andersson *et al.* 2012; Jäderkvist *et al.* 2014, 2015; Kristjánsson 2014; Jäderkvist Fegraeus *et al.* 2015). Interestingly, some connections or overlap have been suggested between *DOCK8* and *DMRT3* genes (Andersson *et al.* 2012; Tassano *et al.* 2016).

Another candidate gene *Glutamate ionotropic receptor NMDA type subunit 2B (GRIN2B)*, located on ECA6, is associated with memory and learning (Kim *et al.* 2018). Two significant SNPs for earned prize money were located within the introns of the *GRIN2B* gene.

The genome-wide association analyses of earned prize money revealed many significant SNPs ($p_{raw} < 1.39 \times 10^{-7}$), the majority of these SNPs were located on ECA6 (Figure 8). The most significant SNP ($p_{raw} = 9.01E-10$) was

detected on ECA 28 in the vicinity of the *NADH: ubiquinone oxidoreductase subunit A12 (NDUFA12)*, which is known to be involved in energy metabolism, aging and, neurodegeneration (Gowthami *et al.* 2019).

No significant SNPs were identified for best km time and number of gallops (Figure 9 and 10). However, two suggestive regions were visible on ECA17 and ECA23 for best km time (Figure 9).

In conclusion, as in study II, our results showed that not only genes associated with physical capacity are of importance for athletic performance, but also mental skills such as memory and learning. Likely, horses that have easy to learn routines for training and competition can adapt more easily and focus on winning the races. According to our results, further studies on mental skills and racing performance are warranted.

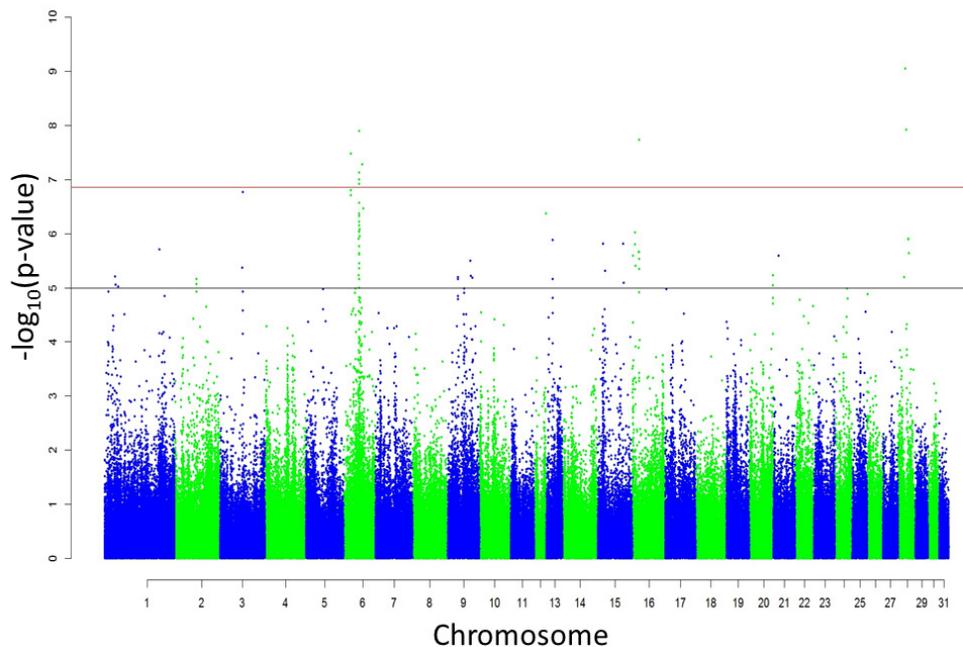


Figure 8. Manhattan plot from genome-wide association analysis of earned prize money. The red line represents Bonferroni genome-wide significance threshold and the black horizontal line indicates the suggestive genome-wide significance threshold. Uncorrected $\lambda = 1.0532$. Significant signal ECA 6, 16, and 28. Multiple suggestive signals on various chromosomes (Velie *et al.* 2018).

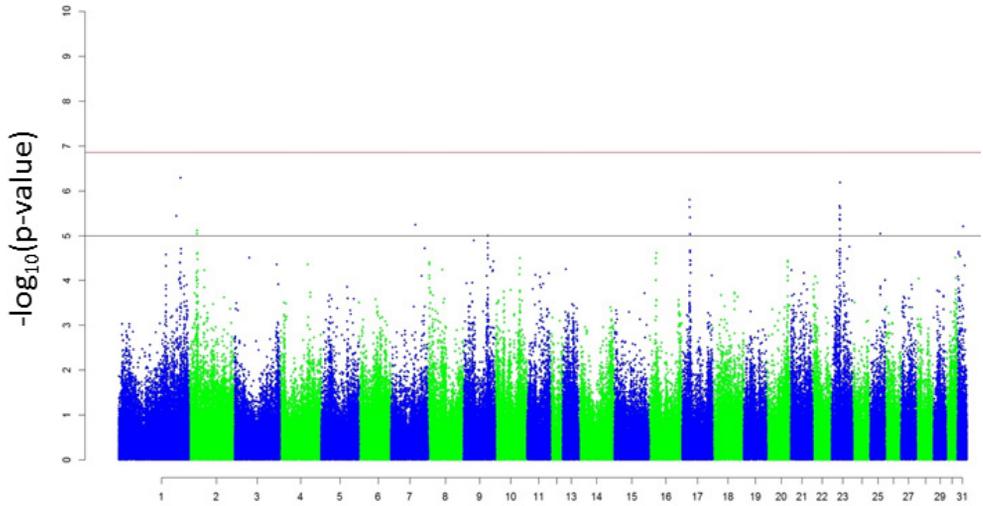


Figure 9. Manhattan plot from genome-wide association analysis of best km time. The red line represents Bonferroni genome-wide significance threshold and the black horizontal line indicates the suggestive genome-wide significance threshold. Uncorrected $\lambda = 1.0902$. Suggestive signals were detected on ECA 17 and 23. A few single suggestive SNPs were also visible on ECA 1, 2, 7, and 31 (Velie *et al.* 2018).

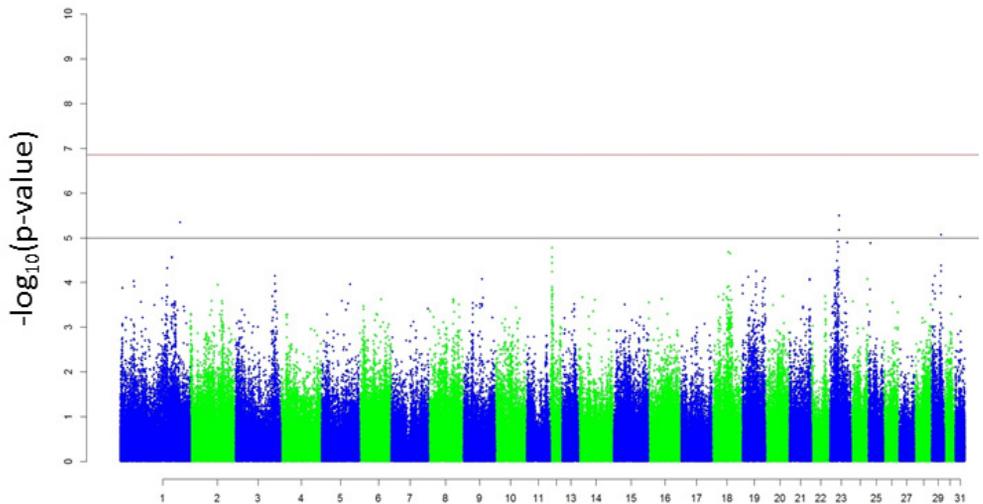


Figure 10. Manhattan plot from genome-wide association analysis of number of gallops. The red line represents Bonferroni genome-wide significance threshold and the black horizontal line indicates the suggestive genome-wide significance threshold. Uncorrected $\lambda = 1.0256$. Suggestive signals with a few single SNPs were detected on ECA 1, 23, and 29 (Velie *et al.* 2018).

3.4 Study IV – Genetic regulation of racing performance and exercise related blood pressure in Coldblooded trotters

In study IV, we dissected a genomic region from previous selective sweep mapping for athletic performance located about 50 kb from the *Endothelin 3* (*EDN3*) gene (Jäderkvist Fegraeus *et al.* 2018). The overall aim was to study regulatory genetics of athletic performance and exercise related blood pressure. We also investigated differences in plasma proteins levels between the haplotype groups (sub-elite and elite performing) before and during exercise.

Results and discussion

The associated genomic region did not harbor any protein coding genes and was syntenic to a region on human chromosome 20 between *EDN3* and *PHACTR3*. Therefore, we hypothesized that a regulatory element was located within the associated region regulating candidate gene or genes outside. To identify candidate causal genetic variants, we applied WGS and targeted long-read sequencing in Coldblooded trotters selected based on associated haplotype and harness racing performance records (i.e. earnings per start). WGS revealed genetic variants with potential regulatory functions. One SNP of which the mutant overlapped a potential transcription factor binding site. The SNP was associated with multiple performance traits. In addition, a tandem duplication of 37.6 kb that involved the *EDN3* promoter flank region and the two first exons of the *EDN3* gene was detected in horses carrying the elite performing haplotype.

Horses carrying the sub-elite performing haplotype had higher plasma concentration of EDN1 and lower plasma concentration of EDN3 than the elite performing haplotype group. They also had higher blood pressure during exercise and 20 minutes after the complete exercise. There were no differences in blood pressure between the haplotype groups at rest.

The relative quantitative proteomics analysis showed that proteins linked to inflammatory and coagulation processes were differentially expressed between the haplotype groups. During exercise compared to rest, horses carrying the sub-elite performing haplotype had higher plasma levels of proteins involved in complement and coagulation processes. The complement system is a part of the innate immune system (Rus *et al.* 2005).

At rest, the sub-elite haplotype group had higher levels of proteins linked to immune response, metabolism, and complement and coagulation systems. The horses in the present study performed an intense exercise. In humans as well as horses, intense exercise is known to induce physical stress response to obtain optimal energy and oxygen to the working muscles. This physical and oxidative stress also affects the inflammatory and coagulation systems (Barton *et al.* 2003; Donovan *et al.* 2007a; Holbrook *et al.* 2010; Scoppetta *et al.* 2012; Kostrzewa-Nowak *et al.* 2020). Vascular endothelial cells play a significant role in regulating blood pressure and hemostasis. Inflammatory cytokines and turbulent blood flow generated from vasoconstriction negatively affect the function of the endothelial cells (Poher & Sessa 2007; Dinh *et al.* 2014).

Horses carrying the elite performing haplotype likely perform better because they have better oxygenation of the working muscles as the blood vessels are dilated and the blood pressure is lower. On the other hand, horses carrying the sub-elite performing haplotype had higher plasma concentration of EDN1, lower plasma concentration of EDN3, and higher blood pressure during exercise, resulting in impaired blood flow to the working muscles due to vascular constriction. This further leads to anaerobic work, increased oxidative stress, and inflammation. Horses carrying the sub-elite performing haplotype needed a longer time to recover and get back to resting blood pressure values after the exercise. This information can be useful when the horses are trained.

In conclusion, further fine-mapping and functional genomic studies are needed to identify causative genetic variants. Inflammatory factors could potentially be used as biomarkers to identify horses that are more likely to tolerate intense exercise and to monitor subclinical exercise induced disorders. Our results contribute to increased knowledge about the molecular genetics of athletic performance and biological processes activated by exercise in horses. In addition, the horse can serve as a model for studying the molecular genetics behind complex biological processes activated by intense exercise.

3.5 Study V – A QTL for conformation of back and croup influences lateral gait quality in Icelandic horses

In study V a genome-wide association analysis (GWA) was performed in 177 Icelandic horses to detect genomic regions linked to conformation of back and croup. We also investigated how these regions affected riding performance traits scored at breeding field tests.

Results and discussion

A QTL for the conformation of back and croup was identified on *Equus caballus* (ECA) 22 (Figure 11). Thirteen SNPs reached the suggestive level of which ten were in LD ($r_2 \geq 0.8$) (Figure 12).

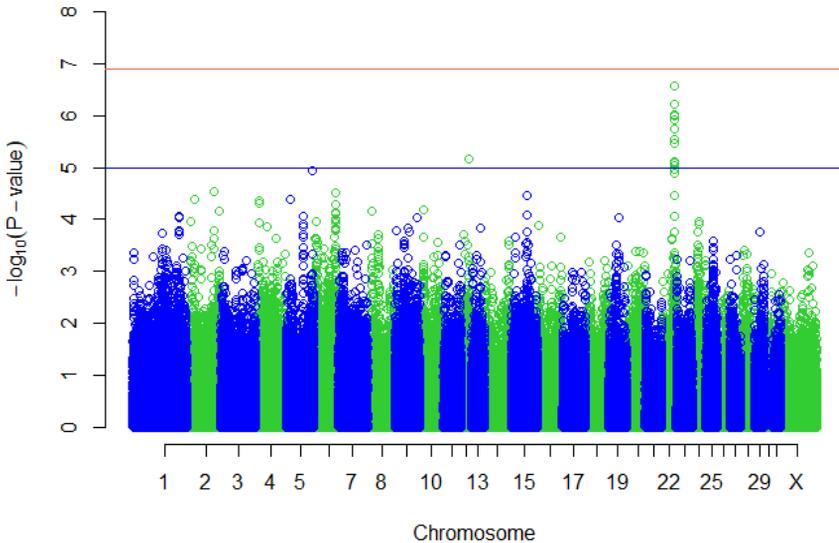


Figure 11. Manhattan plot from genome-wide association analysis of the conformation of back and croup. The red line represents Bonferroni genome-wide significance threshold ($p < 6.9 \times 10^{-8}$) and the blue horizontal line represents the suggestive genome-wide significance threshold ($p < 1.0 \times 10^{-5}$). The estimated λ was 0.98 (se 2.55×10^{-5}). A QTL was detected on ECA 22 (Rosengren *et al.* 2021).

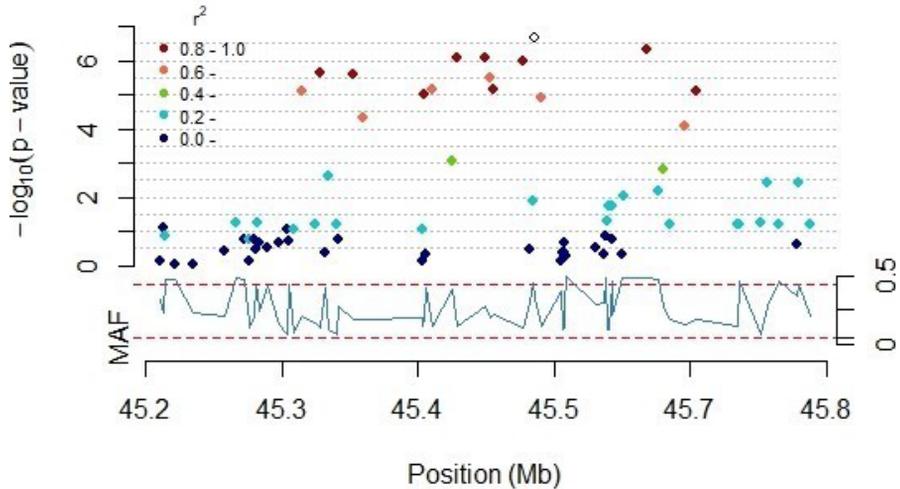


Figure 12. LD Manhattan plot from genome-wide association analysis of the conformation of back and croup. With a zoomed view of the signal on ECA 22. The top SNP, i.e. the most significant SNP is indicated as an open circle, Thirteen SNPs reached the suggestive genome-wide significance threshold ($p < 1.0 \times 10^{-5}$). Of these SNPs were ten in LD ($r^2 \geq 0.8$) (Rosengren et al. 2021).

Haplotype association analysis resulted in two opposite haplotypes associated with higher and lower scores for back and croup (p -value < 0.001). To investigate the effect of the QTL, we compared the two haplotypes. Horses with the favorable haplotype (associated with higher scores for back and croup) had on average higher scores for the lateral gaits tölt and pace, an uphill conformation with longer front legs, less deep breast, and positive remarks on backline and croup type (Table 1). An uphill conformation (higher front part of the horse compared to hind) and croup dimensions are known to be favorable for performance and gait ability and could aid collection and lightness of the front (Albertsdóttir et al, 2008; Kristjánsson et al, 2016). Further, the length of legs can influence stride length (Hoyt *et al.* 2000; Pontzer 2007; Sparrow *et al.* 2017). All these traits affect gait quality and are of great importance when the Icelandic horses are evaluated at the breeding field test (FEIF 2021).

Table 1. Significant differences in traits assessed at breeding field tests between Icelandic horses carrying the favorable and unfavorable haplotype for the score of back and croup (Rosengren *et al.* 2021).

Trait	Favorable haplotype		Unfavorable haplotype		<i>t</i> -value	df	<i>p</i> -value
	N	Mean	N	Mean			
Back and croup	34	8.29	28	7.71	4.05	58.08	<0.001
Tölt ¹	33	8.41	27	7.96	2.52	45.79	0.015
Pace ¹	33	7.18	27	6.09	2.99	50.24	0.004
Slow tölt ¹	33	8.14	26	7.73	2.14	45.19	0.038
Depth at breast ²	33	63.2	28	64.6	-3.52	56.22	0.001
Width of the hips ²	23	47.0	20	48.1	-2.21	37.54	0.033
Width between thigh bones ²	23	43.0	20	44.2	-2.23	38.86	0.031
Length of forelimbs ²	33	15.2	28	12.1	3.22	40.81	0.003
Backline ³	34	1.79	28	2.25	-2.69	58.91	0.009
Croup type ³	34	1.85	28	2.18	-2.31	53.23	0.025

¹Subjectively assessed traits (scale 5-10)

²Zoometric measurements (cm)

³Subjectively assessed sub-traits (scale 1-3)

N = Number of horses

p-values generated from Student's *t*-test

Interestingly, the QTL harbored genes potentially linked to scoliosis and anthropometric traits in humans (Kim *et al.* 2010; Liu *et al.* 2018). There are as far as we know no studies on the prevalence of scoliosis and other back problems in Icelandic horses. In general, mild back problems can be hard to detect clinically in horses because of the strong musculature and ligaments of the back (Jeffcott 1980). A few cases of scoliosis have been described in horses and the reported clinical signs were a lateral position of the head, difficulties to walk in a straight line, impaired action of the hind limbs, and inflexibility of the back (Boyd 1976; Jeffcott 1980; Wong *et al.* 2006). In humans, scoliosis is known to result in general muscle weakness and impaired exercise performance (Martínez-Llorens *et al.* 2010). In addition, scoliosis is suggested to be linked to motor laterality (handedness) in humans (Grivas *et al.* 2004, 2006; Milenkovic *et al.* 2004; Yang & Li 2011;

Catanzariti *et al.* 2014). As well as humans, horses commonly show motor laterality (McGreevy & Thomson 2006; Lucidi *et al.* 2013; Cully *et al.* 2018). Some horses have harder finding balance and straightness during training, which is of great importance for lateral gaits.

In conclusion, our results showed that the detected QTL affected not only conformation of back and croup, but also quality of the lateral gaits tölt and pace. Based on the results from this study, a genetic test has been developed that may be valuable for breeders and trainers to help in their selection of breeding horses. In addition, horses can potentially serve as a model to increase the knowledge about scoliosis and motor laterality in humans. Further studies are warranted to fully understand the biological background of the QTL and how it affects back and croup conformation, as well as the quality of the lateral gaits in Icelandic horses and other breeds.

4. General discussion and future perspectives

In this thesis, the genetics of athletic performance and conformation in horses, as well as the level of inbreeding in Coldblooded trotters, were investigated.

In paper I, the genomic inbreeding coefficient in Coldblooded trotters was estimated. The coldblooded trotters have been strongly selected for racing performance since the 1950s, and their athletic capacity has remarkably improved (Arnason *et al.* 1989; Uhlin 2007). However, the selection has also resulted in a gradual increase in the inbreeding level, which can negatively affect health and racing performance (Klemetsdal 1998; Todd *et al.* 2018; Samsonstuen *et al.* 2020). Coldblooded trotters consist of a relatively small population, and a few popular stallions dominate the breeding (Svensk Travsport 2020). Effective methods are needed to monitor and limit inbreeding to avoid future problems with hereditary diseases. The change in the level of inbreeding was investigated between the years 2000 to 2009 using both pedigree and high-density genotyping array data. Both methods showed similar results with a gradually increased inbreeding level. However, the inbreeding coefficient based on pedigree data was lower, most likely because old pedigree data may contain incorrect information. Different definitions of ROH, such as minimum length, the minimum number of SNPs, missing genotypes, and heterozygous SNPs, affect the results. We tested different thresholds and selected the one that resulted in the highest correlation with the pedigree-based level of inbreeding to capture longer ROH common within the population. Genomic data gives more exact information and detects genomic regions sensitive to inbreeding. Genomic measures of inbreeding could be introduced into the EBV as a tool when selecting horses for breeding to preserve genetic diversity.

In this thesis, different genetic mapping methods were used to investigate the genetics of athletic performance and conformation traits in horses. Both performance and conformation are complex traits influenced by multiple factors. The results from papers II and III indicated that athletic performance requires not only physical characteristics but also mental qualities. One of the candidate genomic regions detected in paper II harbored the gene *SORCS3* associated with spatial memory and learning (Breiderhoff *et al.* 2013). The genome scan in Coldblooded trotters in paper III revealed the

candidate genes *DOCK8* associated with the immune system, intelligence, and motor skills, as well as *GRIN2B* linked to memory and learning (Griggs *et al.* 2008; Kim *et al.* 2018). Likely, horses that have easy to understand routines for training and competition can better adapt and focus on winning the races. During domestication, horses were selected for mental skills suited for horseback riding and chariotry. Horses that were docile and less sensitive to stress were easier to handle. The *ZFPMI* gene associated with fear memory and temperament was selected during the domestication process (Aki & Miczek 2014; Suri *et al.* 2018; Librado *et al.* 2021). The identification of these candidate genes motivates future studies aimed to disentangle the molecular mechanisms behind mental skills and racing performance.

In paper IV we investigated regulatory genetics of athletic performance and exercise related blood pressure in Coldblooded trotters. Horses with the elite performing haplotype had higher plasma concentration of EDN3 and lower exercise induced blood pressure. In addition, these horses had lower plasma concentration of EDN1. EDN3 dilates arteries and increases the blood volume, whereas EDN1 overall has opposite effects (Yanagisawa *et al.* 1988; Rossi 1993; Rossi *et al.* 1999). The blood vessels need to be dilated during exercise to support the working muscles with energy and oxygen. Moreover, a larger blood volume is required to obtain sufficient blood flow to the active muscles and for sweating to regulate the temperature (McKeever & Hinchcliff 1995).

The relative quantitative proteomics analysis in paper IV revealed that proteins linked to inflammatory and coagulation processes were differentially expressed between the haplotype groups at rest and during exercise. During exercise compared to rest, horses carrying the sub-elite performing haplotype had higher levels of proteins related to inflammatory and coagulation processes. Intense exercise activates a physical stress response to obtain more energy for the working muscles. The released stress hormones also affect inflammatory and coagulation processes. (Donovan *et al.* 2007a; Holbrook *et al.* 2010; Scoppetta *et al.* 2012; Kim *et al.* 2021). Endothelial cells in the vascular wall have a significant role in maintaining blood flow by regulating the diameter of the blood vessels and by preventing the blood from clotting (Poher & Sessa 2007; Dinh *et al.* 2014). The blood flows more efficiently with less turbulence if the blood vessels are wide and if the blood is less viscous (Poole & Erickson 2004). The blood vessels need to be dilated during intense exercise to supply the working muscles with

energy and oxygen. However, inflammatory factors and turbulent flow disturb the function of the endothelial cells (Poher & Sessa 2007; Dinh *et al.* 2014). The oxidative stress marker Thiobarbituric Acid Reactive Substances (TBARS) is associated with decreased endurance in horses (Holbrook *et al.* 2010). Pro-inflammatory cytokines such as IL-6 and TNF- α are known to be activated during intense exercise in both horses and humans (Donovan *et al.* 2007b; Gondim *et al.* 2009; Holbrook *et al.* 2010; Scoppetta *et al.* 2012; Kim *et al.* 2021). The generated cytokines and oxidative stress markers could be used as biomarkers to monitor exercise induced disorders and identify horses that are more likely to tolerate intense training and maximum exercise. In future studies, we would like to follow up the horses included in paper IV, measure blood pressure while the horses are running, and take blood samples before, during, and as well as post exercise to validate potential biomarkers. In addition, the detected proteins need to be further validated with other protein detection methods such as ELISA and Western blot.

Conformation is essential for health, durability, and performance. It is well known that conformation of back and croup affects riding ability and performance in Icelandic horses (Albertsdóttir *et al.* 2008; Kristjánsson *et al.* 2016). In paper V, we investigated the genome of 177 Icelandic horses assessed at breeding field tests to dissect the genetics of conformation of back and croup. A genomic region was detected containing genes linked to anthropometric traits, scoliosis, and motor laterality in humans. The genomic region affected conformation of back and croup and the quality of the lateral gaits tölt and pace. Horses with the genetic variants associated with a higher score for back and croup had higher scores for tölt and pace and an uphill conformation (higher front part of the horse compared to hind). They had a less deep breast, longer front legs, and a well-balanced backline. The backline is the line from the withers and back to the lumbosacral joint. The backline should not be forward sloping or swayback. An uphill conformation and croup dimensions play a significant role in sports performance and riding ability in Icelandic horses. The detected QTL harbored genes associated with scoliosis in humans (Liu *et al.* 2018). Interestingly, there is a potential link between scoliosis and motor laterality in humans (Grivas *et al.* 2004, 2006; Milenkovic *et al.* 2004; Yang & Li 2011; Catanzariti *et al.* 2014). In addition, scoliosis is known to result in general muscle weakness and impaired exercise performance in humans (Martínez-Llorens *et al.* 2010). Just like

humans, horses display handedness, and some horses even have problems walking in a straight line when they start to be trained. Usually, these horses need more training to strengthen their balance. In future research, we would like to use biomechanical monitoring tools to perform a detailed investigation of how the detected QTL affects the gait quality in Icelandic horses as well as in other gaited breeds. In addition, as conformation and locomotion patterns are associated with athletic performance, we would like to investigate whether the detected QTL for back and croup affects harness racing performance success.

The detected significant association signals from papers IV and V were located within non-coding regions and could therefore have a transcriptional regulatory function on genes encoding proteins of importance for the studied traits. High-resolution chromosome conformation assays such as 4C and HiCap would help us identify causative regulatory genetic variants and their target genes. With HiCap, it is possible to map individual enhancer interactions with known promoters in the whole genome (Sahlén *et al.* 2015). In future research, we would like to perform chromosome conformation experiments in cell types relevant for the studied traits in wild type and mutant horses followed by functional validation of detected potential causative genetic variants using CRISPR-Cas-based editing.

In conclusion, this thesis contributes to increased knowledge about the molecular genetics of athletic performance and biological processes activated by intense exercise as well as conformation traits in horses. The obtained results will be valuable for breeders and trainers in their selection of breeding horses and how to optimize the training of their horses. The horse can also serve as an excellent model to understand the genetics and biological mechanisms behind complex traits such as athletic performance, blood pressure regulation, inflammation, locomotion pattern, and anthropometric traits of importance for humans other mammalian species.

5. Concluding remarks

This thesis contributes to the understanding of the molecular genetics of athletic performance, conformation, and quality of lateral gaits in horses. The results can be valuable for trainers to adjust the training of the horses and for breeders when selecting horses for breeding. In addition, the horse can be used as a model for studying the molecular genetics of complex traits and biological processes activated by intense exercise.

- The level of inbreeding had gradually increased in Coldblooded trotters during the investigated years 2000 to 2009. Genomic measures of relationship are more exact and can identify genomic regions sensitive for inbreeding. Thus, genomic measures of relationship between potential breeding horses could be introduced when selecting horses for breeding to minimize further increased inbreeding.
- The results from papers II and III indicated that athletic performance requires not only physical characteristics but also mental qualities such as memory and learning.
- In paper IV, candidate genetic variants influencing athletic performance and exercise related blood pressure regulation were detected. Coldblooded trotters carrying the elite performing haplotype likely perform better because they can achieve improved oxygenation of the working muscles as the blood vessels are dilated, and the blood pressure is lower. On the other hand, Coldblooded trotters carrying the sub-elite performing haplotype had higher blood pressure during exercise and needed a longer time to recover and return to resting blood pressure levels post-exercise. This information can be valuable when the horses are trained. In addition, horses harboring the sub-elite performing haplotype had higher plasma levels of proteins linked to inflammatory and coagulation processes. Inflammatory factors and oxidative stress markers could be used as biomarkers to monitor exercise induced disorders such as overtraining and identify horses that are more likely to tolerate intense training and maximum exercise.

- A QTL for back and croup was detected in Icelandic horses that affected the conformation of back and croup and the quality of lateral gaits. Based on the results from this study a genetic test was developed that can be useful when horses are selected for breeding. The results also highlighted potential links between scoliosis and motor laterality.

References

- Ablondi, M., Eriksson, S., Tetu, S., Sabbioni, A., Viklund, Å. & Mikko, S. (2019). Genomic Divergence in Swedish Warmblood Horses Selected for Equestrian Disciplines. *Genes*, vol. 10 (12), 976
- Adalsteinsson, S. (1981). Origin and conservation of farm animal populations in Iceland. *Journal of Animal Breeding and Genetics*, vol. 98 (1–4), pp. 258–264
- Aki, T. & Miczek, K.A. (2014). Neurogenetics of Aggressive Behavior – Studies in Rodents. *Curr Top Behav Neurosci.*, pp. 1–41
- Albertsdóttir, E., Eriksson, S., Näsholm, A., Strandberg, E. & Árnason, T. (2008). Genetic correlations between competition traits and traits scored at breeding field-tests in Icelandic horses. *Livestock Science*, vol. 114 (2–3), pp. 181–187
- Alhaji, M. & Farhana, A. (2021). Enzyme Linked Immunosorbent Assay. *StatPearls*, StatPearls Publishing.
- Andersson, L. (2016). Domestic animals as models for biomedical research. *Uppsala Journal of Medical Sciences*, vol. 121 (1), pp. 1–11
- Andersson, L.S., Larhammar, M., Memic, F., Wootz, H., Schwochow, D., Rubin, C.J., Patra, K., Arnason, T., Wellbring, L., Hjälml, G., Imstrand, F., Petersen, J.L., McCue, M.E., Mickelson, J.R., Cothran, G., Ahituv, N., Roepstorff, L., Mikko, S., Vallstedt, A., Lindgren, G., Andersson, L. & Kullander, K. (2012). Mutations in DMRT3 affect locomotion in horses and spinal circuit function in mice. *Nature*, vol. 488 (7413), pp. 642–646 Nature Publishing Group.
- Arai, H., Hori, S., Aramori, Ichiro, Ohkubo, H. & Nakanishi, S. (1990). Cloning and expression of a cDNA encoding an endothelin receptor. *Nature*, vol. 348 (December), pp. 730–732
- Ardestani, S.S., Aminafshar, M., Maryam, M.B.Z.B., Banabazi, M.H., Sargolzaei, M. & Miar, Y. (2020). Whole-genome signatures of selection in sport horses revealed selection footprints related to musculoskeletal system development processes. *Animals*, vol. 10 (1)
- Arnason, T., Bendroth, M., Philipsson, J., Henriksson, K. & Darenius, A. (1989). Genetic evaluation of Swedish trotters. In State of breeding evaluation in trotters., 1989. pp. 106–130. EAAP
- Barton, M.H., Williamson, L., Jacks, S. & Norton, N. (2003). Effects on plasma endotoxin and eicosanoid concentrations and serum cytokine activities in horses competing in a 48-, 83-, or 159-km endurance ride under similar

- terrain and weather conditions. *American Journal of Veterinary Research*, vol. 64 (6), pp. 754–761
- Begum, G., Albanna, A., Bankapur, A., Nassir, N., Tambi, R., Berdiev, B.K., Akter, H., Karuvantevida, N., Kellam, B., Alhashmi, D., Sung, W.W.L., Thiruvahindrapuram, B., Alsheikh-ali, A., Scherer, S.W. & Uddin, M. (2021). Long-Read Sequencing Improves the Detection of Structural Variations Impacting Complex Non-Coding Elements of the Genome. *International Journal of Molecular Sciences*, pp. 1–14
- Binns, M.M., Boehler, D.A., Bailey, E., Lear, T.L., Cardwell, J.M. & Lambert, D.H. (2012). Inbreeding in the Thoroughbred horse. *Animal Genetics*, vol. 43 (3), pp. 340–342
- Bjørnstad, G. & Røed, K.H. (2001). Breed demarcation and potential for breed allocation of horses assessed by microsatellite markers. *Animal Genetics*, vol. 32, pp. 59–65
- Blackstock, W.P. & Weir, M.P. (1999). Proteomics: quantitative and physical mapping of cellular proteins. *Trends in Biotechnology*, vol. 17 (3), pp. 121–127 Elsevier Current Trends.
- Boyd, J.S. (1976). Congenital Deformities in Two Clydesdale Foals. *Equine Veterinary Journal*, vol. 8 (4), pp. 161–164
- Breiderhoff, T., Christiansen, G.B., Pallesen, L.T., Vaegter, C., Nykjaer, A., Holm, M.M., Glerup, S. & Willnow, T.E. (2013). Sortilin-Related Receptor SORCS3 Is a Postsynaptic Modulator of Synaptic Depression and Fear Extinction. *PLoS ONE*, vol. 8 (9), pp. 1–15
- Burns, E.N., Bordbari, M.H., Mienaltowski, M.J., Affolter, V.K., Barro, M. V., Gianino, F., Gianino, G., Giulotto, E., Kalbfleisch, T.S., Katzman, S.A., Lassaline, M., Leeb, T., Mack, M., Müller, E.J., MacLeod, J.N., Ming-Whitfield, B., Alanis, C.R., Raudsepp, T., Scott, E., Vig, S., Zhou, H., Petersen, J.L., Bellone, R.R. & Finno, C.J. (2018). Generation of an equine biobank to be used for Functional Annotation of Animal Genomes project. *Animal Genetics*, vol. 49 (6), pp. 564–570
- Bush, W.S. & Moore, J.H. (2012). Chapter 11 : Genome-Wide Association Studies. *PLoS Comput Biol.*, vol. 8 (12): e1002822
- Catanzariti, J.F., Guyot, M.A., Agnani, O., Demaille, S., Kolanowski, E. & Donze, C. (2014). Eye-hand laterality and right thoracic idiopathic scoliosis. *European Spine Journal*, vol. 23 (6), pp. 1232–1236 Springer Verlag.
- Charlesworth, B. & Charlesworth, D. (1999). The genetic basis of inbreeding depression. *Genet Res*, pp. 329–340 SLU Library.
- Chua, B.H., Chua, C.C., Diglio, C.A. & Siu, B.B. (1993). Regulation of endothelin-1 mRNA by angiotensin II in rat heart endothelial cells. *Biochimica et Biophysica Acta*, vol. 1178, pp. 201–206
- Clarke, J., Wu, H.C., Jayasinghe, L., Patel, A., Reid, S. & Bayley, H. (2009). Continuous base identification for single-molecule nanopore DNA

- sequencing. *Nature Nanotechnology*, vol. 4 (4), pp. 265–270 Nature Publishing Group.
- Couetil, L., Cardwell, J.M., Leguillette, R., Mazan, M., Richard, E., Bienzle, D., Bullone, M., Gerber, V., Ivester, K., Lavoie, J.P., Martin, J., Moran, G., Niedźwiedz, A., Pusterla, N. & Swiderski, C. (2020). Equine Asthma: Current Understanding and Future Directions. *Frontiers in Veterinary Science*, vol. 7 (July), pp. 1–21
- Cully, P., Nielsen, B., Lancaster, B., Martin, J. & McGreevy, P. (2018). The laterality of the gallop gait in Thoroughbred racehorses. (Bartos, L., ed.) *PLOS ONE*, vol. 13 (6), p. e0198545 Public Library of Science.
- Darwin, C. (1859). *On the origins of species by means of natural selection or the preservation of favoured races in the struggle for life*. London: John Murray.
- Dinh, Q.N., Drummond, G.R., Sobey, C.G. & Chrissobolis, S. (2014). Roles of inflammation, oxidative stress, and vascular dysfunction in hypertension. *BioMed Research International*, vol. 2014
- Donovan, D.C., Jackson, C.A., Colahan, P.T., Norton, N. & Hurley, D.J. (2007a). Exercise-induced alterations in pro-inflammatory cytokines and prostaglandin F2 a in horses. vol. 118, pp. 263–269
- Donovan, D.C., Jackson, C.A., Colahan, P.T., Norton, N. & Hurley, D.J. (2007b). Exercise-induced alterations in pro-inflammatory cytokines and prostaglandin F2 α in horses. *Veterinary Immunology and Immunopathology*, vol. 118 (3–4), pp. 263–269 Elsevier.
- Drews, R. (2004). *Early riders: The beginnings of mounted warfare in Asia and Europe*. New York and London: Routledge.
- Erickson, H. (1993). Exercise physiology. In: Swenson, M. & Reece, W. (eds.) *Dukes physiology of domestic animals*. Ithaca: Cornell University Press, 11th. ed, p. 305.
- Erickson, H. & Poole, D. (2004). The Cardiovascular System. In: Reece, W.O. (ed.) *Dukes' Physiology of Domestic Animals*. Ithaca: Cornell University Press, 12. ed.
- Fages, A., Hanghøj, K., Khan, N., Gaunitz, C., Seguin-Orlando, A., Leonardi, M., McCrory Constantz, C., Gamba, C., Al-Rasheid, K.A.S, Albizuri, S., Alfarhan, A.H., Allentoft, M., Alquraishi, S., Anthony, D., Baimukhanov, N., Barrett, J.H., Bayarsaikhan, J., Benecke, N., Bernáldez-Sánchez, E., Berrocal-Rangel, L., Biglari, F., Boessenkool, S., Boldgiv, B., Brem, G., Brown, D., Burger, J., Crubézy, E., Dagnora, L., Davoudi, H., de Barros Damgaard, P., de Los Ángeles de Chorro Y de Villa-Ceballos, M., Deschler-Erb, S., Detry, C., Dill, N., do Mar Oom, M., Dohr, A., Ellingvåg, S., Erdenebaatar, D., Fathi, H., Felkel, S., Fernández-Rodríguez, C., García-Viñas, E., Germonpré, M., Granada, J.D., Hallsson, J.H., Hemmer, H., Hofreiter, M., Kasparov, A., Khasanov, M., Khazaeli, R., Kosintsev, P.,

- Kristiansen, K., Kubatbek, T., Kuderna, L., Kuznetsov, P., Laleh, H., Leonard, J.A., Lhuillier, J., Liesau von Lettow-Vorbeck, C., Logvin, A., Lõugas, L., Ludwig, A., Luis, C., Arruda, A.M., Marques-Bonet, T., Matoso Silva, R., Merz, V., Mijiddorj, E., Miller, B.K., Monchalov, O., Mohaseb, F.A., Morales, A., Nieto-Espinet, A., Nistelberger, H., Onar, V., Pálsdóttir, A.H., Pitulko, V., Pitskhelauri, K., Pruvost, M., Rajic Sikanjic, P., Rapan Papeša, A., Roslyakova, N., Sardari, A., Sauer, E., Schafberg, R., Scheu, A., Schibler, J., Schlumbaum, A., Serrand, N., Serres-Armero, A., Shapiro, B., Sheikhi Seno, S., Shevnina, I., Shidrang, S., Southon, J., Star, B., Sykes, N., Taheri, K., Taylor, W., Teegen, W.R., Trbojević Vukičević, T., Trixl, S., Tumen, D., Undrakhbold, S., Usmanova, E., Vahdati, A., Valenzuela-Lamas, S., Viegas, C., Wallner, B., Weinstock, J., Zaibert, V., Clavel, B., Lepetz, S., Mashkour, M., Helgason, A., Stefánsson, K., Barrey, E., Willerslev, E., Outram, A.K., Librado, P. & Orlando, L. (2019). Tracking Five Millennia of Horse Management with Extensive Ancient Genome Time Series. *Cell*, vol. 177(6), pp. 1419-1435
- FAO (2021). *Food and Agriculture Organization of the United Nations FAOSTAT Statistics Database*. Available at: <http://www.fao.org/faostat/en> [2021-05-07]
- FEIF (2021). *General Rules and Regulations Breeding Rules and Regulations. FEIF International Federation of Icelandic Horse Associations*. Available at: https://www.feiffengur.com/documents/FEIF_Breeding_Rules_Regulation_s_2021.pdf [2021-05-07]
- François, L., Fegraeus, K.J., Eriksson, S., Andersson, L.S., Tesfayonas, Y.G., Viluma, A., Imsland, F., Buys, N., Mikko, S., Lindgren, G. & Velie, B.D. (2016). Conformation Traits and Gaits in the Icelandic Horse are Associated with Genetic Variants in Myostatin (MSTN). *Journal of Heredity*, vol. 107 (5), pp. 431–437
- Fried, M. & Crothers, D.. (1981). Equilibria and kinetics of lac repressor-operator interactions by polyacrylamide gel electrophoresis. *Nucleic acids research*, vol. 9 (23), pp. 6505–6525
- Gaunitz, C., Fages, A., Hanghøj, K., Albrechtsen, A., Khan, N., Schubert, M., Seguin-Orlando, A., Owens, I.J., Felkel, S., Bignon-Lau, O., Damgaard, P. de B., Mittnik, A., Mohaseb, A.F., Davoudi, H., Alquraishi, S., Alfarhan, A.H., Al-Rasheid, K.A.S., Crubézy, E., Benecke, N., Olsen, S., Brown, D., Anthony, D., Massy, K., Pitulko, V., Kasparov, A., Brem, G., Hofreiter, M., Mukhtarova, G., Baimukhanov, N., Lõugas, L., Onar, V., Stockhammer, P.W., Krause, J., Boldgiv, B., Undrakhbold, S., Erdenebaatar, D., Lepetz, S., Mashkour, M., Ludwig, A., Wallner, B., Merz, V., Merz, I., Zaibert, V., Willerslev, E., Librado, P., Outram, A.K. & Orlando, L. (2018). Ancient genomes revisit the ancestry of domestic and Przewalski's horses. *Science*, vol. 360 (6384), pp. 111–114

- Genereux, D.P., Serres, A., Armstrong, J., Johnson, J., Marinescu, V.D., Murén, E., Juan, D., Bejerano, G., Casewell, N.R., Chemnick, L.G., Damas, J., Di Palma, F., Diekhans, M., Fiddes, I.T., Garber, M., Gladyshev, V.N., Goodman, L., Haerty, W., Houck, M.L., Hubley, R., Kivioja, T., Koepfli, K.P., Kuderna, L.F.K., Lander, E.S., Meadows, J.R.S., Murphy, W.J., Nash, W., Noh, H.J., Nweeia, M., Pfennig, A.R., Pollard, K.S., Ray, D.A., Shapiro, B., Smit, A.F.A., Springer, M.S., Steiner, C.C., Swofford, R., Taipale, J., Teeling, E.C., Turner-Maier, J., Alfoldi, J., Birren, B., Ryder, O.A., Lewin, H.A., Paten, B., Marques-Bonet, T., Lindblad-Toh, K. & Karlsson, E.K. (2020). A comparative genomics multitool for scientific discovery and conservation. *Nature*, vol. 587 (7833), pp. 240–245
- Gondim, F.J., Zoppi, C.C., dos Reis Silveira, L., Pereira-da-Silva, L. & Vaz de Macedo, D. (2009). Possible Relationship Between Performance and Oxidative Stress in Endurance Horses. *Journal of Equine Veterinary Science*, vol. 29 (4), pp. 206–212
- Gowthami, N., Sunitha, B., Kumar, M., Keshava Prasad, T.S., Gayathri, N., Padmanabhan, B. & Srinivas Bharath, M.M. (2019). Mapping the protein phosphorylation sites in human mitochondrial complex I (NADH: Ubiquinone oxidoreductase): A bioinformatics study with implications for brain aging and neurodegeneration. *Journal of Chemical Neuroanatomy*, vol. 95, pp. 13–28 Elsevier.
- Griggs, B.L., Ladd, S., Saul, R.A., DuPont, B.R. & Srivastava, A.K. (2008). Deducator of cytokinesis 8 is disrupted in two patients with mental retardation and developmental disabilities. *Genomics*, vol. 91 (2), pp. 195–202 Academic Press.
- Grivas, T., Mihas, K., Vasiliadis, E., MAZIOTOU, C., KARATHANOU, S. & POLYZOIS, V. (2004). Handedness and Laterality of Trunk Rotation in Children Screened for Scoliosis., 2004. pp. 64–67
- Grivas, T.B., Vasiliadis, E.S., Polyzois, V.D. & Mouzakis, V. (2006). Trunk asymmetry and handedness in 8245 school children. *Pediatric Rehabilitation*, vol. 9 (3), pp. 259–266 Taylor & Francis.
- Gurgul, A., Jasielczuk, I., Semik-Gurgul, E., Pawlina-Tyszko, K., Stefaniuk-Szmukier, M., Szmatoła, T., Polak, G., Tomczyk-Wrona, I. & Bugno-Poniewierska, M. (2019). A genome-wide scan for diversifying selection signatures in selected horse breeds. *PLoS ONE*, vol. 14 (1), pp. 1–24
- Haberle, V. & Stark, A. (2018). Eukaryotic core promoters and the functional basis of transcription initiation. *Nature Reviews Molecular Cell Biology*, vol. 19 (10), pp. 621–637
- Heather, J.M. & Chain, B. (2016). The sequence of sequencers: The history of sequencing DNA. *Genomics*, vol. 107 (1), pp. 1–8 The Authors.

- Hellman, L.M. & Fried, M.G. (2007). Electrophoretic mobility shift assay (EMSA) for detecting protein-nucleic acid interactions. *Nature Protocols*, vol. 2 (8), pp. 1849–1861 Nature Publishing Group.
- Hill, E.W., Gu, J., Eivers, S.S., Fonseca, R.G., McGivney, B.A., Govindarajan, P., Orr, N., Katz, L.M. & MacHugh, D. (2010). A sequence polymorphism in MSTN predicts sprinting ability and racing stamina in thoroughbred horses. *PLoS ONE*, vol. 5 (1), pp. 2–7
- Holbrook, T.C., McFarlane, D. & Schott II, H.C. (2010). Neuroendocrine and non-neuroendocrine markers of inflammation associated with performance in endurance horses. *Equine Veterinary Journal*, vol. 42, pp. 123–128
- Holmström, M. & Back, W. (2013). The effects of conformation. *Equine Locomotion*. London: WB Saunders, 2nd. ed, pp. 229–243.
- Howard, J.T., Pryce, J.E., Baes, C. & Maltecca, C. (2017). Invited review: Inbreeding in the genomics era: Inbreeding, inbreeding depression, and management of genomic variability. *Journal of Dairy Science*, vol. 100 (8), pp. 6009–6024 American Dairy Science Association.
- Hoyt, D.F., Wickler, S.J. & Cogger, E.A. (2000). Time of contact and step length: the effect of limb length, running speed, load carrying and incline. *Journal of Experimental Biology*, vol. 203 (2), pp. 221–227
- Hyland, A. (2003). *The Horse in the Ancient World*. Gloucestershire: Sutton Publishing.
- IFHA (2018). *International Federation of Horseracing Authorities*. Available at: https://www.ifhaonline.org/resources/Annual_Report_2018.pdf [2021-05-07]
- Jäderkvist Fegraeus, K., Johansson, L., Maenpaa, M., Mykkanen, A., Andersson, L.S., Velie, B.D., Andersson, L., Arnason, T. & Lindgren, G. (2015). Different DMRT3 genotypes are best adapted for harness racing and riding in finnhorses. *Journal of Heredity*, vol. 106 (6), pp. 734–740
- Jäderkvist Fegraeus, K., Velie, B.D., Axelsson, J., Ang, R., Hamilton, N.A., Andersson, L., Meadows, J.R.S. & Lindgren, G. (2018). A potential regulatory region near the EDN3 gene may control both harness racing performance and coat color variation in horses. *Physiological Reports*, vol. 6 (10)
- Jäderkvist, K., Andersson, L.S., Johansson, A.M., Arnason, T., Mikko, S., Eriksson, S., Andersson, L. & Lindgren, G. (2014). The DMRT3 ‘ Gait keeper ’ mutation affects performance of Nordic and Standardbred trotters. *Journal of Animal Science*, vol. 92 (10), pp. 4279–4286
- Jäderkvist, K., Holm, N., Imsland, F., Arnason, T., Andersson, L., Andersson, L.S. & Lindgren, G. (2015). The importance of the DMRT3 “Gait keeper” mutation on riding traits and gaits in Standardbred and Icelandic horses. *Livestock Science*, vol. 176, pp. 33–39 Elsevier.

- Jalali Sefid Dashti, M. & Gamieldien, J. (2017). A practical guide to filtering and prioritizing genetic variants. *BioTechniques*, vol. 62 (January), pp. 18–30
- Janczarek, I., Wilk, I. & Strzelec, K. (2017). Correlations between body dimensions of young trotters and motion parameters and racing performance. *Pferdeheilkunde*, vol. 33 (2), pp. 139–145
- Jeffcott, L.B. (1980). Disorders of the thoracolumbar spine of the horse — a survey of 443 cases. *Equine Veterinary Journal*, vol. 12 (4), pp. 197–210 American Medical Association (AMA).
- Jiang, F. & Doudna, J.A. (2017). CRISPR – Cas9 Structures and Mechanisms. *Annual review of biophysics*, vol. 46, pp. 505–531
- Jönsson, L., Egenvall, A., Roepstorff, L., Näsholm, A., Dalin, G. & Philipsson, J. (2014a). Associations of health status and conformation with longevity and lifetime competition performance in young Swedish Warmblood riding horses: 8,238 cases (1983-2005). *Journal of the American Veterinary Medical Association*, vol. 244 (12), pp. 1449–1461 American Veterinary Medical Association.
- Jönsson, L., Näsholm, A., Roepstorff, L., Egenvall, A., Dalin, G. & Philipsson, J. (2014b). Conformation traits and their genetic and phenotypic associations with health status in young Swedish warmblood riding horses. *Livestock Science*, vol. 163 (1), pp. 12–25 Elsevier.
- Jordbruksverket *Bevara, nyttja och utveckla – handlingsplan för uthållig förvaltning av svenska husdjursraser 2010–2020*. Available at: http://www2.jordbruksverket.se/webdav/files/SJV/trycksaker/Pdf_rapporte/r/ra10_14.pdf
- Kalbfleisch, T.S., Rice, E.S., DePriest, M.S., Walenz, B.P., Hestand, M.S., Vermeesch, J.R., O'Connell, B.L., Fiddes, I.T., Vershinina, A.O., Saremi, N.F., Petersen, J.L., Finno, C.J., Bellone, R.R., McCue, M.E., Brooks, S.A., Bailey, E., Orlando, L., Green, R.E., Miller, D.C., Antczak, D.F. & MacLeod, J.N. (2018). Improved reference genome for the domestic horse increases assembly contiguity and composition. *Communications Biology*, vol. 1 (197), pp. 1–8
- Kawaguchi, H., Sawa, H. & Yasuda, H. (1990). Endothelin stimulates angiotensin I to angiotensin II conversion in cultured pulmonary artery endothelial cells. *J Mol Cell Cardiol*, vol. 22, pp. 839–842
- Kawaguchi, H., Sawa, H. & Yasuda, H. (1991). Effect of endothelin on angiotensin converting enzyme activity in cultured pulmonary artery endothelial cells. *J Hypertens*, vol. 9 (2), pp. 171–174
- Kim, D.S., Wheeler, M.T. & Ashley, E.A. (2021). The genetics of human performance. *Nature Reviews Genetics*, vol. 23 (January) Springer US.
- Kim, J.I., Yoo, J.H., Kim, D., Jeong, B. & Kim, B.N. (2018). The effects of GRIN2B and DRD4 gene variants on local functional connectivity in attention-

- deficit/hyperactivity disorder. *Brain Imaging and Behavior*, vol. 12 (1), pp. 247–257 *Brain Imaging and Behavior*.
- Kingsley, N.B., Hamilton, N.A., Lindgren, G., Orlando, L., Bailey, E., Brooks, S., McCue, M., Kalbfleisch, T.S., MacLeod, J.N., Petersen, J.L., Finno, C.J. & Bellone, R.R. (2021). “Adopt-a-Tissue” Initiative Advances Efforts to Identify Tissue-Specific Histone Marks in the Mare. *Frontiers in Genetics*, vol. 12 (March), pp. 1–9
- Kingsley, N.B., Kern, C., Creppe, C., Hales, E.N., Zhou, H., Kalbfleisch, T.S., Macleod, J.N., Petersen, J.L., Finno, C.J. & Bellone, R.R. (2020). Functionally annotating regulatory elements in the equine genome using histone mark chip-seq. *Genes*, vol. 11 (1)
- Klemetsdal, G. (1998). Inbreeding on Racing Performance. *Genetics Selection Evolution*, vol. 30, pp. 351–366
- Kostrzewa-Nowak, D., Kubaszewska, J., Nowakowska, A. & Nowak, R. (2020). Effect of Aerobic and Anaerobic Exercise on the Complement System of Proteins in Healthy Young Males. *Journal of clinical medicine*, vol. 9 (2357)
- Kristjánsson, T., Bjornsdóttir, S., Albertsdóttir, E., Sigurdsson, A., Pourcelot, P., Crevier-Denoix, N. & Arnason, T. (2016). Association of conformation and riding ability in Icelandic horses. *Livestock Science*, vol. 189 (July 2016), pp. 91–101
- Kristjánsson, Þ. (2014). *Riding Ability in Icelandic Horses – Effect of Conformation and the ‘Gait Keeper’ – Mutation in the DMRT3 Gene*.
- Krivega, I. & Dean, A. (2012). Enhancer and promoter interactions — long distance calls. *Curr Opin Genet Dev*, vol. 22 (2), pp. 79–85
- Librado, P., Fages, A., Gaunitz, C., Leonardi, M., Wagner, S., Khan, N., Hanghøj, K., Alquraishi, S.A., Alfarhan, A.H., Al-Rasheid, K.A., Der Sarkissian, C., Schubert, M. & Orlando, L. (2016). The evolutionary origin and genetic makeup of domestic horses. *Genetics*. Genetics Society of America. DOI: <https://doi.org/10.1534/genetics.116.194860>
- Librado, P., Khan, N., Fages, A., Kusliy, M.A., Suchan, T., Tonasso-Calvière, L., Schiavinato, S., Alioglu, D., Fromentier, A., Perdereau, A., Aury, J.M., Gaunitz, C., Chauvey, L., Seguin-Orlando, A., Der Sarkissian, C., Southon, J., Shapiro, B., Tishkin, A.A., Kovalev, A.A., Alquraishi, S., Alfarhan, A.H., Al-Rasheid, K.A.S., Seregély, T., Klassen, L., Iversen, R., Bignon-Lau, O., Bodu, P., Olive, M., Castel, J.C., Boudadi-Maligne, M., Alvarez, N., Germonpré, M., Moskal-Del Hoyo, M., Wilczyński, J., Pospuła, S., Lasota-Kuś, A., Tunia, K., Nowak, M., Rannamäe, E., Saarma, U., Boeskorov, G., Lõugas, L., Kyselý, R., Peške, L., Bălăşescu, A., Dumitraşcu, V., Dobrescu, R., Gerber, D., Kiss, V., Szécsényi-Nagy, A., Mende, B.G., Gallina, Z., Somogyi, K., Kulcsár, G., Gál, E., Bendrey, R., Allentoft, M.E., Sirbu, G., Dergachev, V., Shephard, H., Tomadini, N.,

- Grouard, S., Kasparov, A., Basilyan, A.E., Anisimov, M.A., Nikolskiy, P.A., Pavlova, E.Y., Pitulko, V., Brem, G., Wallner, B., Schwall, C., Keller, M., Kitagawa, K., Bessudnov, A.N., Bessudnov, A., Taylor, W., Magail, J., Gantulga, J.O., Bayarsaikhan, J., Erdenebaatar, D., Tabaldiev, K., Mijiddorj, E., Boldgiv, B., Tsagaan, T., Pruvost, M., Olsen, S., Makarewicz, C.A., Lamas, S., Albizuri Canadell, S., Nieto Espinet, A., Iborra, M.P., Lira Garrido, J., Rodríguez González, E., Celestino, S., Olària, C., Arsuaga, J.L., Kotova, N., Pryor, A., Crabtree, P., Zhumatayev, R., Toleubaev, A., Morgunova, N.L., Kuznetsova, T., Lordkipanize, D., Marzullo, M., Prato, O., Bagnasco Gianni, G., Tecchiati, U., Clavel, B., Lepetz, S., Davoudi, H., Mashkour, M., Berezina, N.Y., Stockhammer, P.W., Krause, J., Haak, W., Morales-Muñiz, A., Benecke, N., Hofreiter, M., Ludwig, A., Graphodatsky, A.S., Peters, J., Kiryushin, K.Y., Iderkhangai, T.O., Bokovenko, N.A., Vasiliev, S.K., Seregin, N.N., Chugunov, K.V., Plasteeva, N.A., Baryshnikov, G.F., Petrova, E., Sablin, M., Ananyevskaya, E., Logvin, A., Shevnina, I., Logvin, V., Kalieva, S., Loman, V., Kukushkin, I., Merz, I., Merz, V., Sakenov, S., Varfolomeyev, V., Usmanova, E., Zaibert, V., Arbuckle, B., Belinskiy, A.B., Kalmykov, A., Reinhold, S., Hansen, S., Yudin, A.I., Vybornov, A.A., Epimakhov, A., Berezina, N.S., Roslyakova, N., Kosintsev, P.A., Kuznetsov, P.F., Anthony, D., Kroonen, G.J., Kristiansen, K., Wincker, P., Outram, A & Orlando, L. (2021). The origins and spread of domestic horses from the Western Eurasian steppes. *Nature*, vol. 598, pp. 634–640
- Lieberman-Aiden, E., Berkum, N.L. Van, Williams, L., Ragoczy, T., Telling, A., Amit, I., Lajoie, B.R., Peter, J., Dorschner, M.O., Sandstrom, R., Bernstein, B., Groudine, M., Gnirke, A., Stamatoyannopoulos, J. & Leonid, A. (2009). Comprehensive mapping of long range interactions reveals folding principles of the human genome. *Science*, vol. 326 (October 9), pp. 289–293
- Lindåse, S., Nostell, K., Söder, J. & Bröjer, J. (2017). Relationship Between β -cell Response and Insulin Sensitivity in Horses based on the Oral Sugar Test and the Euglycemic Hyperinsulinemic Clamp. *Journal of Veterinary Internal Medicine*, vol. 31 (5), pp. 1541–1550
- Lindgren, G., Backström, N., Swinburne, J., Hellborg, L., Einarsson, A., Sandberg, K., Cothran, G., Vilà, C., Binns, M. & Ellegren, H. (2004). Limited number of patrilineal lines in horse domestication. *Nature Genetics*, vol. 36 (4), pp. 335–336
- Liu, J., Zhou, Y., Liu, S., Song, X., Yang, X.Z., Fan, Y., Chen, W., Akdemir, Z.C., Yan, Z., Zuo, Y., Du, R., Liu, Z., Yuan, B., Zhao, S., Liu, G., Chen, Y., Zhao, Y., Lin, M., Zhu, Q., Niu, Y., Liu, P., Ikegawa, S., Song, Y.Q., Posey, J.E., Qiu, G., Zhang, F., Wu, Z., Lupski, J.R. & Wu, N. (2018). The coexistence of copy number variations (CNVs) and single nucleotide

- polymorphisms (SNPs) at a locus can result in distorted calculations of the significance in associating SNPs to disease. *Human Genetics*, vol. 137 (6–7), pp. 553–567 Springer Verlag.
- Lucidi, P., Bacco, G., Sticco, M., Mazzoleni, G., Benvenuti, M., Bernabò, N. & Trentini, R. (2013). Assessment of motor laterality in foals and young horses (*Equus caballus*) through an analysis of derailment at trot. *Physiology and Behavior*, vol. 109 (1), pp. 8–13 Elsevier Inc.
- Maeda, S., Miyauchi, T., Iemitsu, M., Tanabe, T., Irukayama-Tomobe, Y., Goto, K., Yamaguchi, I. & Matsuda, M. (2002). Involvement of endogenous endothelin-1 in exercise-induced redistribution of tissue blood flow: An endothelin receptor antagonist reduces the redistribution. *Circulation*, vol. 106 (17), pp. 2188–2193
- Mantere, T., Kersten, S. & Hoischen, A. (2019). Long-read sequencing emerging in medical genetics. *Frontiers in Genetics*, vol. 10 (MAY), pp. 1–14
- Marchini, J., Cardon, L.R., Phillips, M.S. & Donnelly, P. (2004). The effects of human population structure on large genetic association studies. vol. 36 (5), pp. 512–517
- Martínez-Llorens, J., Ramírez, M., Colomina, M.J., Bagó, J., Molina, A., Cáceres, E. & Gea, J. (2010). Muscle dysfunction and exercise limitation in adolescent idiopathic scoliosis. *Eur Respir J*, vol. 36, pp. 393–400
- Mathesius, U., Imin, N., Natera, S.H.A. & Rolfe, B.G. (2003). Proteomics as a Functional Genomics Tool. *Methods in molecular biology (Clifton, N.J.)*, vol. 236, pp. 395–413 Humana Press.
- Mazzocchi, G., Malendowicz, L.K., Meneghelli, V. & Nussdorfer, G.G. (1992). Endothelin-1 stimulates mitotic activity in the zona glomerulosa of the rat adrenal cortex. *Cytobios*, vol. 69 (277), pp. 91–96
- Mazzocchi, G., Rebuffat, P. & Meneghelli, V. (1990). Effects of Prolonged Infusion With Endothelin-1 on the Function and Morphology of Rat Adrenal Cortex. *Peptides*, vol. 11, pp. 767–772
- McCue, M.E., Bannasch, D.L., Petersen, J.L., Gurr, J., Bailey, E., Binns, M.M., Distl, O., Guérin, G., Hasegawa, T., Hill, E.W., Leeb, T., Lindgren, G., Penedo, M.C.T., Røed, K.H., Ryder, O.A., Swinburne, J.E., Tozaki, T., Valberg, S.J., Vaudin, M., Lindblad-Toh, K., Wade, C.M. & Mickelson, J.R. (2012). A high density SNP array for the domestic horse and extant *Perissodactyla*: Utility for association mapping, genetic diversity, and phylogeny studies. *PLoS Genetics*, vol. 8 (1)
- McGivney, B.A., Browne, J.A., Fonseca, R.G., Katz, L.M., MacHugh, D.E., Whiston, R. & Hill, E.W. (2012). MSTN genotypes in Thoroughbred horses influence skeletal muscle gene expression and racetrack performance. *Animal Genetics*, vol. 43 (6), pp. 810–812
- McGivney, B.A., Han, H., Corduff, L.R., Katz, L.M., Tozaki, T., MacHugh, D.E. & Hill, E.W. (2020). Genomic inbreeding trends, influential sire lines and

- selection in the global Thoroughbred horse population. *Scientific Reports*, vol. 10 (1), pp. 1–12
- McGreevy, P.D. & Thomson, P.C. (2006). Differences in motor laterality between breeds of performance horse. *Applied Animal Behaviour Science*, vol. 99 (1–2), pp. 183–190 Elsevier.
- McKeever, K.H., Hinchcliff, K. V, Schmall, L.M. & Muir 3rd, W.W. (1991). Renal tubular function in horses during submaximal exercise. *Am. J. Physiol*, vol. 261 (Sep), pp. R553-560
- McKeever, K.H. & Hinchcliff, K.W. (1995). Neuroendocrine control of blood volume, blood pressure and cardiovascular function in horses. *Equine Veterinary Journal*, vol. 27 (18 S), pp. 77–81
- McKeever, K.H., Hinchcliff, K.W., Schmall, L.M., Reed, S.M., Lamb, D.R. & Muir, W.W. (1992). Plasma renin activity and aldosterone and vasopressin concentrations during incremental treadmill exercise in horses. *American journal of veterinary research*, vol. 53 (8), pp. 1290–1293
- Mclaren, W., Gil, L., Hunt, S.E., Riat, H.S., Ritchie, G.R.S., Thormann, A., Flicek, P. & Cunningham, F. (2016). The Ensembl Variant Effect Predictor. *Genome Biology*, pp. 1–14 Genome Biology.
- McPherron, A., Lawler, A. & Lee, S.. (1997). Regulation of skeletal muscle mass in mice by a new TGF-beta superfamily member. *Nature*, vol. 387 (May), pp. 83–90
- Meadows, J.R.S. & Lindblad-Toh, K. (2017). Dissecting evolution and disease using comparative vertebrate genomics. *Nature Reviews Genetics*, vol. 18 (10), pp. 624–636 Nature Publishing Group.
- Milenkovic, S.M., Kocijancic, R.I. & Belojevic, G.A. (2004). Left handedness and spine deformities in early adolescence. *Eur. J. Epidemiol*, vol. 19, pp. 969–972
- Müller, H.P., Sogo, J.M. & Schaffner, W. (1989). An enhancer stimulates transcription in Trans when attached to the promoter via a protein bridge. *Cell*, vol. 58 (4), pp. 767–777 Cell Press.
- Nakatani, Y., Horikoshit, M., Brenner, M., Yamamoto, T., Besnard, F., Roedert, R.G. & Freese, E. (1990). A downstream initiation element required for efficient TATA box binding and in vitro function of TFIID. *Nature*, vol. 348 (November), pp. 1–3
- Negro, J.J., Blázquez, M.C., Fernández-Alés, R. & Martín-Vicente, A. (2021). Interspecific sexual selection, a new theory for an old practice: the increase of artificial biodiversity through creation of modern, standardized breeds. *Animal Biodiversity and Conservation*, vol. 1, pp. 109–115
- Nostell, K., Lindåse, S., Hedberg, H. & Bröjer, J. (2019). The effect of insulin infusion on heart rate and systemic blood pressure in horses with equine metabolic syndrome. *Equine Veterinary Journal*, vol. 51 (6), pp. 733–737

- Nostell, K., Lindåse, S., Winqvist, E. & Bröjer, J. (2021). The effect of diet-induced obesity and pasture on blood pressure and serum cortisol in Standardbred mares. *Equine Veterinary Journal*, vol. 53 (May 2021), pp. 542–548
- Nostell, K.E.A., Lindåse, S.S. & Bröjer, J.T. (2016). Blood pressure in Warmblood horses before and during a euglycemic-hyperinsulinemic clamp. *Acta Veterinaria Scandinavica*, vol. 58 (1), pp. 65–70 BioMed Central.
- Outram, K., Stear, N., Bendrey, R., Olsen, S., Kasparov, A., Zaibert, V., Thorpe, N. & Evershed, R. (2009). The Earliest Horse Harnessing and Milking. *Science*, vol. 323 (March), pp. 1332–1335
- Patterson, S.D. & Aebersold, R. (1995). Mass spectrometric approaches for the identification of gel-separated proteins. *Electrophoresis*, vol. 16 (1), pp. 1791–1814 John Wiley & Sons, Ltd.
- Petersen, J.L., Mickelson, J.R., Cothran, E.G., Andersson, L.S., Axelsson, J., Bailey, E., Bannasch, D., Binns, M.M., Borges, A.S., Brama, P., Leeb, T., Lindgren, G., Lohi, H., Lopes, M.S. & McGivney, B.A. (2013a). Genetic Diversity in the Modern Horse Illustrated from Genome-Wide SNP Data. vol. 8 (1)
- Petersen, J.L., Mickelson, J.R., Rendahl, A.K., Valberg, S.J., Andersson, L.S., Axelsson, J., Bailey, E., Bannasch, D., Binns, M.M., Borges, A.S., Brama, P., da Câmara Machado, A., Capomaccio, S., Cappelli, K., Cothran, E.G., Distl, O., Fox-Clipsham, L., Graves, K.T., Guérin, G., Haase, B., Hasegawa, T., Hemmann, K., Hill, E.W., Leeb, T., Lindgren, G., Lohi, H., Lopes, M.S., McGivney, B.A., Mikko, S., Orr, N., Penedo, M.C.T., Piercy, R.J., Raekallio, M., Rieder, S., Røed, K.H., Swinburne, J., Tozaki, T., Vaudin, M., Wade, C.M. & McCue, M.E. (2013b). Genome-Wide Analysis Reveals Selection for Important Traits in Domestic Horse Breeds. *PLoS Genetics*, vol. 9 (1)
- Pober, J.S. & Sessa, W.C. (2007). Evolving functions of endothelial cells in inflammation. *Nature Reviews Immunology*, vol. 7 (10), pp. 803–815
- Pontzer, H. (2007). Effective limb length and the scaling of locomotor cost in terrestrial animals. *Journal of Experimental Biology*, vol. 210 (10), pp. 1752–1761
- Poole, D.C. & Erickson, H.H. (2004). *Heart and vessels: Function during exercise and response to training*. *Equine Sports Medicine and Surgery*. Elsevier Ltd.
- Posthuma, D. Genome-wide association studies. *Nature Reviews Methods Primers*, Springer US.
- Qin, T., An, Y., Liu, C., Wu, J., Dai, R., Liu, D., Li, X., Jiang, L., Wu, D., Tang, X., Song, W., Wang, T. & Zhao, X. (2015). Novel DOCK8 gene mutations lead to absence of protein expression in patients with hyper-IgE syndrome. *Immunologic Research 2015 64:1*, vol. 64 (1), pp. 260–271 Springer.
- Ricard, A., Robert, C., Blouin, C., Baste, F., Torquet, G., Morgenthaler, C., Rivière, J., Mach, N., Mata, X., Schibler, L. & Barrey, E. (2017). Endurance exercise

- ability in the horse: A trait with complex polygenic determinism. *Frontiers in Genetics*, vol. 8 (JUN), pp. 1–15
- Rodríguez-Rodríguez, E., Gil-Mori6n, J. & Negro, J..Feral animal populations: separating threats from opportunities. *European Journal of Wildlife Research*,
- Rosengren, M.K., Sigurđard6ttir, H., Eriksson, S., Naboulsi, R., Jouni, A., Novoa-bravo, M., Albertsd6ttir, E., Kristj6nsson, Þ., Rhodin, M., Viklund, 6., Velie, B.D., Negro, J.J., Sol6, M. & Lindgren, G. (2021). A QTL for conformation of back and croup influences lateral gait quality in Icelandic horses. *BMC Genomics*, 22, 267
- Rosengren Pielberg, G., Golovko, A., Sundstr6m, E., Curik, I., Lennartsson, J., Seltenhammer, M.H., Druml, T., Binns, M., Fitzsimmons, C., Lindgren, G., Sandberg, K., Baumung, R., Vetterlein, M., Str6mberg, S., Grabherr, M., Wade, C., Lindblad-Toh, K., Pont6n, F., Heldin, C.H., S6lkner, J. & Andersson, L. (2008). A cis-acting regulatory mutation causes premature hair graying and susceptibility to melanoma in the horse. *Nature Genetics*, vol. 40 (8), pp. 1004–1009
- Rossi, G.P., Sacchetto, A., Cesari, M. & Pessina, A.C. (1999). Interactions between endothelin-1 and the renin – angiotensin – aldosterone system. *Cardiovascular Research*, vol. 43, pp. 300–307
- Rossi, N.F. (1993). Effect of endothelin-3 on vasopressin release in vitro and water excretion in vivo in Long-Evans rats. *The Journal of Physiology*, vol. 461 (1), pp. 501–511
- Rossi, N.F. (1995). Cation channel mechanisms in ET-3-induced vasopressin secretion by rat hypothalamo-neurohypophysial explants. *American Journal of Physiology - Endocrinology and Metabolism*, vol. 268 (3 Pt 1)
- Rowell, L.. (1993). Human Cardiovascular Control. New York: Oxford University Press, pp. 441–479.
- Roytman, M., Kichaev, G., Gusev, A. & Pasaniuc, B. (2018). Methods for fine-mapping with chromatin and expression data. *PLoS Genetics*, vol. 14 (2), pp. 1–25
- Rus, H., Cudrici, C. & Niculescu, F. (2005). The Role of the Complement System in Innate Immunity. *Immunologic Research*, (410), pp. 103–112
- Rustin, M., Janssens, S., Buys, N. & Gengler, N. (2009). Multi-trait animal model estimation of genetic parameters for linear type and gait traits in the Belgian warmblood horse. *Journal of Animal Breeding and Genetics*, vol. 126 (5), pp. 378–386
- Sahl6n, P., Abdullayev, I., Ramsk6ld, D., Matskova, L., Rilakovic, N., L6tstedt, B., Albert, T.J., Lundeberg, J. & Sandberg, R. (2015). Genome-wide mapping of promoter-anchored interactions with close to single-enhancer resolution. *Genome Biology*, vol. 16 (1), pp. 1–13 Genome Biology.

- Sahlén, P., Spalinskas, R., Asad, S., Mahapatra, K. Das, Höjer, P., Anil, A., Eisfeldt, J., Srivastava, A., Nikamo, P., Mukherjee, A., Kim, K.H., Bergman, O., Ståhle, M., Sonkoly, E., Pivarcsi, A., Wahlgren, C.F., Nordenskjöld, M., Taylan, F., Bradley, M. & Tapia-Páez, I. (2021). Chromatin interactions in differentiating keratinocytes reveal novel atopic dermatitis- and psoriasis-associated genes. *Journal of Allergy and Clinical Immunology*, vol. 147 (5), pp. 1742–1752
- Sakurai, T., Yanagisawa, M., Takuwat, Y., Miyazaki, H., Kimura, S., Goto, K. & Masaki, T. (1990). Cloning of a cDNA encoding a non-isopeptide-selective subtype of the endothelin receptor. *Nature*, vol. 348 (December), pp. 732–735
- Samsonstuen, S., Dolvik, N.I., Olsen, H.F., Lykkjen, S. & Klemetsdal, G. (2020). Inbreeding affects racing performance negatively in the Standardbred trotter. *Acta Agriculturae Scandinavica A: Animal Sciences*, pp. 152–156 Taylor & Francis.
- Sánchez-Guerrero, M.J., Cervantes, I., Molina, A., Gutiérrez, J.P. & Valera, M. (2017). Designing an early selection morphological linear traits index for dressage in the Pura Raza Español horse. *Animal*, vol. 11 (6), pp. 948–957
- Sánchez-Guerrero, M.J., Molina, A., Gómez, M.D., Peña, F. & Valera, M. (2016). Relationship between morphology and performance: Signature of mass-selection in Pura Raza Español horse. *Livestock Science*, vol. 185, pp. 148–155 Elsevier.
- Sanger, F., Nicklen, S. & Coulson, A.R. (1977). DNA sequencing with chain-terminating inhibitors. *Proc Natl Acad Sci U S A*, vol. 74 (12), pp. 5463–5467
- Schaefer, R.J., Schubert, M., Bailey, E., Bannasch, D.L., Barrey, E., Bar-Gal, G.K., Brem, G., Brooks, S.A., Distl, O., Fries, R., Finno, C.J., Gerber, V., Haase, B., Jagannathan, V., Kalbfleisch, T., Leeb, T., Lindgren, G., Lopes, M.S., Mach, N., da Câmara Machado, A., MacLeod, J.N., McCoy, A., Metzger, J., Penedo, C., Polani, S., Rieder, S., Tammen, I., Tetens, J., Thaller, G., Verini-Supplizi, A., Wade, C.M., Wallner, B., Orlando, L., Mickelson, J.R. & McCue, M.E. (2017). Developing a 670k genotyping array to tag ~2M SNPs across 24 horse breeds. *BMC Genomics*, vol. 18 (1), pp. 1–18 BMC Genomics.
- Scoppetta, F., Tartaglia, M., Renzone, G., Avellini, L., Gaiti, A., Scaloni, A. & Chiaradia, E. (2012). Plasma protein changes in horse after prolonged physical exercise: A proteomic study. *Journal of Proteomics*, vol. 75 (14), pp. 4494–4504 Elsevier B.V.
- Shrestha, M., Eriksson, S., Schurink, A., Andersson, L.S., Sundquist, M., Frey, R., Broström, H., Bergström, T., Ducro, B. & Lindgren, G. (2015). Genome-Wide Association Study of Insect Bite Hypersensitivity in Swedish-Born Icelandic Horses. pp. 366–374

- Shrestha, M., Solé, M., Ducro, B.J., Sundquist, M., Thomas, R., Schurink, A., Eriksson, S. & Lindgren, G. (2020). Genome-wide association study for insect bite hypersensitivity susceptibility in horses revealed novel associated loci on chromosome 1. *Journal of Animal Breeding and Genetics*, vol. 137 (2), pp. 223–233
- Slijper, E.J. (1946). *Comparative biologic-anatomical investigations on the vertebral column and spinal musculature of mammals*. K. Ned. Acad. Wetensch.
- Smale, S.T. (2010). Luciferase Assay. *Cold Spring Harbor Protocols*,
- Söder, J., Bröjer, J.T. & Nostell, K.E. (2012). Interday variation and effect of transportation on indirect blood pressure measurements, plasma endothelin-1 and serum cortisol in Standardbred and Icelandic horses. *Acta veterinaria Scandinavica*, vol. 54, p. 37
- Solé, M., Gómez, M.D., Galisteo, A.M., Santos, R. & Valera, M. (2013). Kinematic Characterization of the Menorca Horse at the Walk and the Trot: Influence of Hind Limb Pastern Angle. *Journal of Equine Veterinary Science*, vol. 33 (9), pp. 726–732 Elsevier Inc.
- Sparrow, L.M., Pellatt, E., Yu, S.S., Raichlen, D.A., Pontzer, H. & Rolian, C. (2017). Gait changes in a line of mice artificially selected for longer limbs. *PeerJ*, vol. 2017 (2), p. e3008 PeerJ Inc.
- Sung, C., Arleth, A., Storer, B. & Ohlstein, E. (1994). Angiotensin type 1 receptors mediate smooth muscle proliferation and endothelin biosynthesis in rat vascular smooth muscle. *Pharmacol Exp Ther.*, vol. 1 (Oct 271), pp. 429–437
- Suri, P., Palmer, M.R., Tsepilov, Y.A., Freidin, M.B., Boer, C.G., Yau, M.S., Evans, D.S. & Gelemanovic, A. (2018). Genome-wide meta-analysis of 158,000 individuals of European ancestry identifies three loci associated with chronic back pain. *PLOS ONE*, vol. 48 (7), pp. 1–23
- Svensk Travsport (2019). *ÅRSSTATISTIK 2019*. Available at: <https://www.travsport.se/siteassets/relaterade-dokument/tavling/statistik/arsstatistik-2019.pdf?823>
- Svensk Travsport (2020). *Kallblodsaveln travar på 2020*. Available at: <https://www.travsport.se/arkiv/nyheter/2020/december/kallblodsaveln-travar-pa-2020/> [2021-10-20]
- Svensk Travsport (2021a). *Avelsvärdering av kallblodiga hingstar*. Available at: <https://www.travsport.se/avel-och-uppfodning/avelshingstar/avelsvardering-av-kallblod/> [2021-05-07]
- Svensk Travsport (2021b). *Hästarna*. Available at: <https://www.travsport.se/svensk-travsport/travsporten-i-sverige/hastarna/> [2021-08-23]
- Tassano, E., Accogli, A., Pavanello, M., Bruno, C., Capra, V., Gimelli, G. & Cuoco, C. (2016). Interstitial 9p24.3 deletion involving only DOCK8 and KANK1

- genes in two patients with non-overlapping phenotypic traits. *European Journal of Medical Genetics*, vol. 59 (1), pp. 20–25 Elsevier Masson.
- Todd, E.T., Ho, S.Y.W., Thomson, P.C., Ang, R.A., Velie, B.D. & Hamilton, N.A. (2018). Founder-specific inbreeding depression affects racing performance in Thoroughbred horses. *Scientific Reports*, vol. 8 (1), pp. 1–10
- Tozaki, T., Sato, F., Hill, E.W., Miyake, T., Endo, Y., Kakoi, H., Gawahara, H., Hirota, K. ichi, Nakano, Y., Nambo, Y. & Kurosawa, M. (2011). Sequence variants at the myostatin gene locus influence the body composition of thoroughbred horses. *Journal of Veterinary Medical Science*, vol. 73 (12), pp. 1617–1624
- Tyers, M. & Mann, M. (2003). From genomics to proteomics. *Nature*, vol. 422 (March), pp. 193–197
- Uhlen, H.-E. (2007). *Vår kallblodstravare*. Ulleriks Hb.
- van der Valk, R.A., Vreede, J.A., Crémazzy, F. & Dame, R.T. (2015). Genomic Looping: A Key Principle of Chromatin Organization. *J Mol Microbiol Biotechnol*, pp. 344–359
- Velie, B.D., Fegraeus, K.J., Solé, M., Rosengren, M.K., Røed, K.H., Ihler, C.-F., Strand, E. & Lindgren, G. (2018). A genome-wide association study for harness racing success in the Norwegian-Swedish coldblooded trotter reveals genes for learning and energy metabolism. *BMC Genetics*, vol. 19 (1)
- Velie, B.D., Lillie, M., Fegraeus, K.J., Rosengren, M.K., Solé, M., Wiklund, M., Ihler, C.-F., Strand, E. & Lindgren, G. (2019a). Exploring the genetics of trotting racing ability in horses using a unique Nordic horse model. *BMC Genomics*, vol. 20 (1)
- Velie, B.D., Shrestha, M., François, L., Schurink, A., Tesfayonas, Y.G., Stinckens, A., Blott, S., Ducro, B.J., Mikko, S., Thomas, R., Swinburne, J.E., Sundqvist, M., Eriksson, S., Buys, N. & Lindgren, G. (2016). Using an Inbred Horse Breed in a High Density Genome-Wide Scan for Genetic Risk Factors of Insect Bite Hypersensitivity (IBH). (Barendse, W., ed.) *PLOS ONE*, vol. 11 (4), p. e0152966 Public Library of Science.
- Velie, B.D., Solé, M., Fegraeus, K.J., Rosengren, M.K., Røed, K.H., Ihler, C.-F., Strand, E. & Lindgren, G. (2019b). Genomic measures of inbreeding in the Norwegian-Swedish Coldblooded Trotter and their associations with known QTL for reproduction and health traits. *Genetics Selection Evolution*, vol. 51 (1)
- Viklund, Å., Näsholm, A., Strandberg, E. & Philipsson, J. (2011). Genetic trends for performance of Swedish Warmblood horses. *Livestock Science*, vol. 141 (2–3), pp. 113–122 Elsevier B.V.
- Wade, C. & Freund, B. (1990). Hormonal control of blood volume during and following exercise. In: Gisolfi, C. V & Lamb, D.R. (eds.) *Fluid Homeostasis*

- During Exercise (Perspectives in Exercise Science & Sports Medicine, Vol. 3)*. Camel, Indiana: Benchmark Press, pp. 207–241.
- Wade, C.M., Giulotto, E., Sigurdsson, S., Zoli, M., Gnerre, S., Imsland, F., Lear, T.L., Adelson, D.L., Bailey, E., Bellone, R.R., Blöcker, H., Distl, O., Edgar, R.C., Garber, M., Leeb, T., Mauceli, E., MacLeod, J.N., Penedo, M.C.T., Raison, J.M., Sharpe, T., Vogel, J., Andersson, L., Antczak, D.F., Biagi, T., Binns, M.M., Chowdhary, B.P., Coleman, S.J., Della Valle, G., Fryc, S., Guérin, G., Hasegawa, T., Hill, E.W., Jurka, J., Kiialainen, A., Lindgren, G., Liu, J., Magnani, E., Mickelson, J.R., Murray, J., Nergadze, S.G., Onofrio, R., Pedroni, S., Piras, M.F., Raudsepp, T., Rocchi, M., Røed, K.H., Ryder, O.A., Searle, S., Skow, L., Swinburne, J.E., Syvänen, A.C., Tozaki, T., Valberg, S.J., Vaudin, M., White, J.R., Zody, M.C., Lander, E.S. & Lindblad-Toh, K. (2009). Genome sequence, comparative analysis, and population genetics of the domestic horse. *Science*, vol. 326 (5954), pp. 865–867
- Wallner, B., Vogl, C., Shukla, P., Burgstaller, J.P., Druml, T. & Brem, G. (2013). Identification of Genetic Variation on the Horse Y Chromosome and the Tracing of Male Founder Lineages in Modern Breeds. vol. 8 (4)
- Wang, K., Li, M. & Hakonarson, H. (2010). ANNOVAR : functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Research*, vol. 38 (16), pp. 1–7
- Weatherby, J. (1791). *An Introduction to a General Stud-Book*. London.
- Weirather, J., de Cesare, M., Wang, Y., Piazza, P., Sebastiano, V., Wang, X., Buck, D. & Au, K. (2017). Comprehensive comparison of Pacific Biosciences and Oxford Nanopore Technologies and their applications to transcriptome analysis [version 2 ; referees : 2 approved] Jason L Weirather Referee Status : *F1000Research*, vol. 6
- Weitzberg, E., Hemsén, A., Lundberg, J.M. & Ahlborg, G. (1995). ET-3 is extracted by and induces potent vasoconstriction in human splanchnic and renal vasculatures. *Journal of Applied Physiology*, vol. 79 (4), pp. 1255–1259
- WHO (2021). *Hypertension*. Available at: <https://www.who.int/news-room/fact-sheets/detail/hypertension> [2021-10-23]
- Wold, M.S., Rich, A., Weeks, D. & Lutter, L.C. (2014). Deoxyribonucleic acid (DNA). *Access Science*, McGraw-Hill Education.
- Wong, D., Miles, K. & Sponseller, B. (2006). CONGENITAL SCOLIOSIS IN A QUARTER HORSE FILLY. *Veterinary Radiology & Ultrasound*, vol. 47 (3), pp. 279–282 John Wiley & Sons, Ltd.
- WorldFengur *The Studbook of Origin for the Icelandic horse*. Available at: <https://www.worldfengur.com/> [2021-05-07]
- Wutke, S., Sandoval-Castellanos, E., Benecke, N., Döhle, H.J., Friederich, S., Gonzalez, J., Hofreiter, M., Löugas, L., Magnell, O., Malaspinas, A.S., Morales-Muñiz, A., Orlando, L., Reissmann, M., Trinks, A. & Ludwig, A.

- (2018). Decline of genetic diversity in ancient domestic stallions in Europe. *Science Advances*, vol. 4 (4)
- Yanagisawa, M., Kurihara, H., Kimura, S., Torno be, Y., Kobayashi, M., Mitsui, Y., YazakP, Y., Goto, K. & Masaki, T. (1988). A novel potent vasoconstrictor peptide produced by vascular endothelial cells. *Nature*, vol. 332, pp. 411–415
- Yang, Z. de & Li, M. (2011). There may be a same mechanism of the left-right handedness and left-right convex curve pattern of adolescent idiopathic scoliosis. *Medical Hypotheses*, vol. 76 (2), pp. 274–276 Churchill Livingstone.
- Zhang, L. & Elias, J.E. (2017). Relative Protein Quantification Using Tandem Mass Tag Mass Spectrometry. In: Comai, L., Katz, J., & Mallick, P. (eds.) *Proteomics*. New York, NY.: Humana Press, pp. 185–198.

Popular science summary

Horses are amazing athletes. They have big hearts that can pump up to 400 liters of blood per minute. In a well-trained elite horse, up to 90 percent of the blood can be distributed to the working muscles. By nature, horses need to escape from predators to survive, and they have since domestication about 4000-5500 years ago been selected for desired traits such as strength, speed, and gait ability. The long history of selection and the population structure with many different breeds make the horse a perfect model for studying the genetics behind complex biological processes. Many diseases and conditions that affect humans are also present in horses. Using the horse as a model, we can get more information about the biological mechanisms behind these conditions and thereby improve diagnosis and treatment.

The overall aims of this thesis were to dissect the genetics of athletic performance and conformation in horses as well as investigate the level of inbreeding in Coldblooded trotters. A strong selection in a limited population can result in inbreeding, negatively affecting performance and health. Coldblooded trotters have been strongly selected for harness racing performance for more than sixty years and a few popular stallions dominate the breeding. Traditionally, the level of inbreeding is monitored by using information from pedigree data. However, improved methods to investigate the horse genome open up the possibility of using genomic data to estimate the level of inbreeding. The genetic diversity decreases when there is a selection, and these genomic footprints of selection can be identified using genomic data. In study I, we investigated the level of inbreeding in Coldblooded trotters using both pedigree and genomic data. The results showed that the level of inbreeding had gradually increased during the studied years 2000 to 2009. The inbreeding coefficient based on genomic

data was higher compared to pedigree data. Pedigree data may contain errors, and genomic data give more exact information and can identify genomic regions sensitive for inbreeding. Thus, genomic measures of relationship between potential breeding horses could be introduced when selecting horses for breeding to minimize further increased inbreeding.

As well as elite human athletes, racehorses need to have an ideal genetic setup for athleticism to tolerate training and perform at an elite level to win races. Athletic performance is a complex trait influenced by multiple factors. In studies II-IV we used different genetic mapping methods to identify genetic factors associated with athletic performance in Coldblooded trotters. The results from studies II and III revealed that athletic performance is influenced by several genes and indicated that not only physical characteristics are important for racing performance, but also mental qualities such as learning and intelligence. Horses that have easy to learn and remember the routines likely have easier to adapt to training and better focus on the race.

In study IV, we investigated genetic regulation of athletic performance and exercise related blood pressure in Coldblooded trotters. The intense exercise that horses perform during training and races induces a physical stress response known as “the fight or flight response” to obtain more energy for the working muscles. The stress hormones that are released also affect inflammatory processes and blood pressure. Horses with the unfavorable genetic variants (negatively associated with athletic performance) had higher plasma concentration of the protein Endothelin 1, which are known to constrict blood vessels. In addition, they had a lower concentration of the vasodilator protein Endothelin 3. These horses had higher blood pressure during exercise and needed a longer time to return to resting blood pressure values after the exercise than the horses carrying the favorable genetic variants (associated with elite performance). High blood pressure results in impaired blood flow to the working muscles due to vascular constriction, which leads to increased oxidative stress and inflammation. Horses with the unfavorable genetic variants had higher levels of proteins linked to inflammation and coagulation processes. Biomarkers for inflammatory factors could be introduced to monitor subclinical exercise induced disorders such as overtraining. Previous studies in horses have shown that the inflammatory factors return to resting values between two to 24 hours after

intense exercise. However, there is a risk for chronic inflammation and overtraining if the horses do not recover between the races.

Conformation is essential for health, durability, and performance. It is well known that conformation of back and croup affects riding ability and performance in Icelandic horses. In study V, we investigated the genome in Icelandic horses assessed at breeding field tests to investigate the genetics of conformation of back and croup. A genomic region was detected containing genes linked to body length, scoliosis, and motor laterality in humans. The genomic region affected conformation of back and croup and the quality of the lateral gaits tölt and pace. Horses with the genetic variants associated with a higher score for back and croup had higher scores for tölt and pace. A genetic test for the detected significant genetic markers has been developed in Icelandic horses.

In conclusion, this thesis contributes to increased knowledge about the molecular genetics of athletic performance, biological processes activated by training as well as conformation traits in horses. The results are valuable for trainers and breeders to aid when selecting horses for breeding. The horse can also serve as a model to understand the genetics and biological mechanisms behind complex traits such as athletic performance, blood pressure regulation, inflammation, and locomotion pattern in humans and other mammalian species.

Populärvetenskaplig sammanfattning

Hästar är utmärkta atleter, deras stora hjärtan kan pumpa upp till 400 liter blod per minut. Hos en vältränad elithäst kan upp till 90 procent av blodet som pumpas ut från hjärtat fördelas till de arbetande musklerna. I hästens ursprung ligger förmågan att kunna fly från rovdjur för att överleva. Dessutom har hästen sedan domesticeringen för ungefär 4000-5500 år sedan selekterats för önskvärda egenskaper som styrka, snabbhet och gångartsförmåga. Den långa selektionsprocessen och populationsstrukturen med många olika raser gör hästen till en perfekt modell för att studera genetiken bakom komplexa biologiska processer. Många sjukdomar och tillstånd som påverkar människor förekommer också hos hästar. Med hästen som modell kan vi få mer information om de biologiska mekanismerna bakom dessa tillstånd och därigenom förbättra diagnostik och behandling.

Det övergripande målet med denna avhandling var att undersöka den genetiska bakgrunden till atletisk prestation och exteriör hos hästar samt undersöka inavelsgraden hos kallblodstravare. Ett för starkt urval i en begränsad population kan resultera i inavel, vilket påverkar prestation och hälsa negativt. Kallblodstravare har sedan mer än sextio år starkt selekterats för prestation på travbanan och ett fåtal populära hingstar dominerar aveln. Inavelsgraden övervakas genom att använda information från stamtavlor. Förbättrade metoder för att undersöka hästgenomet öppnar upp möjligheten att använda molekylär genetisk data för att uppskatta inavelsgraden. Vid selektion minskar den genetiska variationen och dessa genomiska fotavtryck av selektion kan identifieras med hjälp av molekylär genetisk data. I den första studien undersökte vi inavelsgraden hos kallblodstravare genom att använda både information från stamtavlor och molekylär genetisk data. Våra resultat visade att inavelsgraden gradvis ökat under de studerade åren 2000-2009 och att uppskattningen av inavelsgraden baserad på molekylär genetisk

data var högre jämfört med information från stamtavlor. Stamtavlor kan innehålla fel och molekylär genetisk data ger en mera exakt information om inavelsgraden och kan att identifiera regioner i arvsmassan som är känsliga för inavel. Därför skulle genomiska mått på inavelsgrad kunna användas för att förhindra fortsatt ökad inavel.

Liksom elitidrottare måste tävlingshästar ha optimala genetiska förutsättningar för att tolerera hård träning och prestera på elitnivå för att vinna tävlingslopp. Atletisk prestation är en komplex egenskap som påverkas av många faktorer. I studie två till fyra använde vi olika genetiska kartläggningsmetoder för att identifiera genetiska faktorer som påverkar atletisk prestation hos kallblodstravare. Resultaten från studie två och tre visade att atletisk prestation påverkas av många gener och att inte bara fysiska egenskaper är viktiga utan också mentala egenskaper som exempelvis minne och inläring. Hästar som har lätt att lära sig rutinerna för träning och tävling har sannolikt lättare att anpassa sig och fokusera på tävlingsloppen.

I den fjärde studien undersökte vi genetisk reglering av atletisk prestation och blodtryck hos kallblodstravare. Det intensiva arbetet som hästar utför under träning och tävlingslopp framkallar en fysisk stressreaktion som kallas "fight or flight response" för att få mer energi till de arbetande musklerna. Stresshormonerna som frigörs påverkar även inflammatoriska processer samt blodtrycket. Hästarna som bar på de ogynnsamma genetiska varianterna (negativt associerad med atletisk prestation) hade högre koncentration av proteinet Endotelin 1 som är känt för att dra ihop blodkärlen. Dessutom hade de en lägre koncentration av det kärilvidgande proteinet Endotelin 3. Dessa hästar hade även högre blodtryck under träning och behövde längre tid för att återgå till vilovärden efter träningen jämfört med hästarna som bar på de gynnsamma genetiska varianterna (förknippad med elitprestation). Högt blodtryck ger ett försämrat blodflöde till de arbetande musklerna på grund av att kärlen är sammandragna, vilket leder till ökad oxidativ stress och inflammation. Hästarna med de ogynnsamma genetiska varianterna hade högre nivåer av proteiner kopplade till inflammation och koagulation (blodets levringsförmåga). Biomarkörer för inflammatoriska faktorer skulle kunna utformas för att övervaka träningsrelaterade tillstånd såsom överträning. Tidigare studier hos häst har visat att de inflammatoriska faktorerna återgår till vilovärden mellan två till 24 timmar efter intensiv

träning. Det finns dock risk för kronisk inflammation och överträning om hästarna inte återhämtar sig mellan träningspassen och loppen.

Ryggen och korssets exteriör är viktigt för hälsa, hållbarhet och prestation. Det är välkänt att exteriör av rygg och kors påverkar prestationsförmågan hos islandshästar. I den femte studien undersökte vi arvsmassan hos islandshästar som bedömts vid avelsbedömning för att identifiera genetiska bakgrunden till exteriör av rygg och kors. En region hittades i arvsmassan som innehöll gener associerade med kroppslängd, skolios och motorik hos människa. Den genetiska regionen påverkade inte bara exteriör av rygg och kors utan också kvaliteten av de laterala gångarterna passgång och tölt. Hästar med de genetiska varianterna förknippade med högre poäng för rygg och kors hade även högre poäng för tölt och passgång. Ett gentest för de identifierade genetiska markörerna har utvecklats på islandshästar.

Sammanfattningsvis bidrar denna avhandling till ökad kunskap om genetiken bakom atletisk prestation, biologiska processer aktiverade av träning samt exteriör hos hästar. Resultaten är värdefulla för tränare för att anpassa träningen samt för uppfödare som hjälp när hästar ska väljas för avel. Hästen kan också fungera som en modell för att förstå genetiken och de biologiska mekanismerna bakom komplexa egenskaper som atletisk prestation, blodtrycksreglering, inflammation och rörelsemönster hos människor och andra däggdjur.

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RESEARCH ARTICLE

Open Access

Genomic measures of inbreeding in the Norwegian–Swedish Coldblooded Trotter and their associations with known QTL for reproduction and health traits

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Abstract

Background: Since the 1950s, the Norwegian–Swedish Coldblooded trotter (NSCT) has been intensively selected for harness racing performance. As a result, the racing performance of the NSCT has improved remarkably; however, this improved racing performance has also been accompanied by a gradual increase in inbreeding level. Inbreeding in NSCT has historically been monitored by using traditional methods that are based on pedigree analysis, but with recent advancements in genomics, the NSCT industry has shown interest in adopting molecular approaches for the selection and maintenance of this breed. Consequently, the aims of the current study were to estimate genomic-based inbreeding coefficients, i.e. the proportion of runs of homozygosity (ROH), for a sample of NSCT individuals using high-density genotyping array data, and subsequently to compare the resulting rate of genomic-based F_{ROH} to that of pedigree-based F_{PED} coefficients within the breed.

Results: A total of 566 raced NSCT were available for analyses. Average F_{ROH} ranged from 1.78 to 13.95%. Correlations between F_{ROH} and F_{PED} were significant ($P < 0.001$) and ranged from 0.27 to 0.56, with F_{PED} and F_{ROH} from 2000 to 2009 increasing by 1.48 and 3.15%, respectively. Comparisons of ROH between individuals yielded 1403 regions that were present in at least 95% of the sampled horses. The average percentage of a single chromosome covered in ROH ranged from 9.84 to 18.82% with chromosome 31 and 18 showing, respectively, the largest and smallest amount of homozygosity.

Conclusions: Genomic inbreeding coefficients were higher than pedigree inbreeding coefficients with both methods showing a gradual increase in inbreeding level in the NSCT breed between 2000 and 2009. Opportunities exist for the NSCT industry to develop programs that provide breeders with easily interpretable feedback on regions of the genome that are suboptimal from the perspective of genetic merit or that are sensitive to inbreeding within the population. The use of molecular data to identify genomic regions that may contribute to inbreeding depression in the NSCT will likely prove to be a valuable tool for the preservation of its genetic diversity in the long term.

Background

In recent years, there has been a rapid increase in the intensity of selection in many livestock breeding programs with the growing use of elite animals, which

ultimately reduces the effective population size (N_e) of some breeds [1–6]. Consequently, a small N_e not only reduces genetic variability, but it also increases the effects of inbreeding (F) and genetic drift, and potentially alters the patterns of runs of homozygosity (ROH) in the long term [3–8]. While such alterations may not necessarily be of concern for large and highly diverse populations, increased homozygosity at loci with a heterozygous

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advantage in small native populations reduces furthermore their genetic diversity [7]. Small populations can be particularly vulnerable to inbreeding depression since mating between relatives often decreases individual fitness and can significantly reduce population growth [7, 9]. Moreover, selection programs, while driving favorable alleles to fixation, also allow deleterious alleles to hitchhike along with favorable mutations. In addition to this, more intense selection resulting from combining genomic selection with embryo biotechnologies (e.g. artificial insemination) not only increases rates of genetic gain, but can also increase levels of inbreeding [4, 10].

The Norwegian–Swedish Coldblooded trotter (NSCT) is a domestic breed of horse in Norway and Sweden and is one of the few remaining descendants of the original Nordic coldblooded horse [11]. Since the 1950s, the breed has been intensively selected for harness racing performance with estimated breeding values produced annually since the 1980s [12, 13]. As a result, a remarkable improvement in the racing performance of NSCT has occurred during the last half-century. However, this improved racing performance has also been accompanied by a gradual increase in pedigree-based F levels [14]. Although today NSCT is considered as a relatively healthy breed, the NSCT breeding industry is well aware that increased levels of inbreeding are widely known to increase the expression of recessive deleterious alleles that are linked to genetic diseases. Historically, inbreeding in NSCT has been monitored by using traditional methods that are based on pedigree analysis [14, 15]. While informative, the NSCT industry understands that this classical metric likely underestimates inbreeding within the breed and does not account for the fact that homozygosity at some regions may, in principal, be more or less desirable than at other regions. Two animals that have the same level of inbreeding, may display drastically different unfavorable effects of inbreeding. Even with an extensive and complete pedigree, realized inbreeding levels will likely differ from pedigree-based F levels due to recombination and Mendelian sampling, which is then compounded by the fact that, although the base animals in a pedigree are considered unrelated, they are more often than not, related.

Consequently, the NSCT industry has actively supported a shift towards using genomic data for F calculations in the breed, thus allowing for diversity across the entire genome as well as at specific regions to be evaluated and monitored, and providing not only a more accurate assessment of inbreeding within the breed, but also a much more detailed assessment. As such, the aims of the current study were to provide genomic-based F coefficient estimates (F_{ROH}) for a sample of NSCT using a high-density genotyping array and to compare the

rate of F_{ROH} to that of classical pedigree-based F (F_{PED}) within the breed. Common ROH within the breed were also assessed for overlaps with previously characterized quantitative trait loci (QTL) for health and reproduction traits in the horse, thus providing a first look at genomic regions and traits that may warrant industry intervention in the future.

Methods

Pedigree data

Complete pedigree information on all raced and unraced NSCT were provided by the trotter associations in both Norway and Sweden (Det Norske Travselvskap and Svensk Travsport). The pedigree consisted of 112,195 individuals with a median pedigree depth of 15 generations.

Collection of samples

In total, 566 individuals born between 1 January 2000 and 31 December 2009 were selected for this study based on the following criteria: (1) each horse had to have participated in at least one race during its lifetime; this restriction was implemented to allow for a broader use of the data in future analyses that will explore racing performance traits within the breed; (2) hair and/or blood samples had to be readily accessible from the pedigree registration authorities in either Norway (Department of Basic Sciences and Aquatic Medicine, Norwegian University of Life Sciences) or Sweden (Animal Genetics Laboratory, Swedish University of Agricultural Sciences); and (3) a sufficient amount of sample material had to be available to ensure high DNA quality standards.

DNA isolation

DNA was extracted from hair roots using a standard procedure of hair preparation. Briefly, 186 μL of Chelex 100 Resin (Bio-Rad Laboratories, Hercules, CA) and 14 μL of proteinase K (20 mg/mL; Merck KgaA, Darmstadt, Germany) were added to each sample. This mix was incubated at 56 °C for 2 h and proteinase K was inactivated for 10 min at 95 °C. For DNA preparation from blood, DNA from 350- μL blood samples were extracted by using the Qiasymphony instrument and the Qiasymphony DSP DNA mini kit (Qiagen, Hilden, Germany).

Genotyping and quality control

Prior to quality control (QC), the dataset consisted of individuals that were genotyped with the 670K Axiom equine genotyping array ($n=473$) and the 670K+ Axiom equine genotyping array ($n=93$). Data from the two arrays were subsequently merged based on SNP name, chromosome number and position, which yielded a combined SNP dataset of 611,888 SNPs for 566 horses (SNPs

located on chromosomes X and Y were excluded during this process). Then, QC was performed with the PLINK v1.07 software. SNPs were screened based on minor allele frequency (MAF > 0.01), Hardy–Weinberg equilibrium ($p > 0.0001$), and genotyping rate (> 0.95) with data that did not conform to these criteria and individuals with missing genotypes (> 15%) being removed. Descriptive data for the sample of horses used in the analyses are in Table 1.

Inbreeding coefficient and runs of homozygosity

Inbreeding coefficients (F_{PED}) were calculated based on the complete pedigree of the breed using the Contribution, Inbreeding (F), Coancestry v1.0 software, which uses a modified algorithm of Sargolzaei et al. [16] to compute inbreeding coefficients that is a fast and accurate tool for F_{PED} calculations.

Inconsistency between ROH-defining criteria in various industries and breeds has been shown to convolute the comparison of studies over time and across population samples [1–6, 17–22]. Since the criteria to define a ROH continue to remain ambiguous, in our study, we applied a wide range of ROH-defining criteria. Runs of homozygosity were defined in PLINK v1.07 using the sliding windows approach through the *homozyg* command. The details of each applied threshold setting are in Table S1 (see Additional file 1: Table S1). Genomic inbreeding coefficients (F_{ROH}) were estimated for each

threshold setting by dividing the summed length of all ROH (per individual) by the length of the genome (2,242,879,462 bp) covered with SNPs. Pearson correlation coefficients between F_{PED} and all F_{ROH} were determined using the statistical software R [23]. Paired t-tests between all F_{ROH} were also performed.

To better identify population-wide ROH in the breed, custom scripts in R were applied to ROH data from the threshold setting that resulted in the highest correlation between F_{PED} and F_{ROH} . These scripts were used to determine which regions of the genome were shared in at least 95% of individuals in the sample [23]. Ultimately, we chose the threshold setting that resulted in the highest correlation between F_{PED} and F_{ROH} since not only did it allow the capture of longer ROH that would subsequently be more beneficial when evaluating previously associated QTL, but it also yielded a more conservative estimate of inbreeding within the breed (i.e. an estimate that was more likely to be skewed upwards than downwards). Homozygous regions that were present in at least 95% of the sampled NSCT were then compared to previously reported QTL for reproduction and health traits in the horse (downloaded from the horse QTL database; [24]) using bed file comparisons in BEDOPS [25].

Results

After QC, 360,977 autosomal SNPs and 566 horses were available for analyses. Summary statistics, stratified by country of birth, for F_{PED} are in Table 2. F_{PED} and F_{ROH} of Norwegian born horses were higher than those of Swedish born horses, although the highest F_{PED} estimate was found for a Swedish born horse. Median F_{PED} and F_{ROH} for the entire cohort of sampled horses, stratified by year, are shown in Fig. 1. Inbreeding in the NSCT population during the 2000–2009 period increased by 1.48 and 3.15% based on F_{PED} and F_{ROH} estimates, respectively. Average F_{ROH} (%) ranged from 1.78 to 13.95% (see Additional file 1: Table S1). Correlations between F_{PED} and all F_{ROH} estimates were significant ($P < 0.001$) and ranged from 0.27 to 0.56 (see Additional file 2: Table S2) and Fig. 2. The threshold settings as defined below resulted in the highest correlation ($R = 0.5629$) between F_{PED} and F_{ROH} :

- Size of the sliding window in SNPs: 50 SNPs.
- Minimum length in kb that a run must have to be called as a ROH: 500.
- Minimum number of SNPs that a run must have to be called as a ROH: 100.
- Number of heterozygous SNPs allowed in a ROH: 1.
- Number of missing calls allowed in a ROH: 5.

Table 1 Descriptive data on the genotyped horses

	Number
Sex	
Intact males	56
Females	222
Geldings	288
Country of birth	
Norway	265
Sweden	301
Year of birth	
2000	25
2001	60
2002	72
2003	53
2004	55
2005	40
2006	60
2007	70
2008	63
2009	68
Total	566

Table 2 Descriptive results, stratified by country of birth, for average inbreeding coefficient (F_{PED}) and average genomic inbreeding coefficient (F_{ROH}) for a sample of raced Norwegian–Swedish Coldblooded trotters born between 1 January 2000 and 31 December 2009

	Min	25th percentile	Median	Mean	75th percentile	Max
Country of birth Norway						
F_{PED} (%)	0.96	5.18	6.18	6.59	7.38	14.35
F_{ROH} (%) ^a	1.98	8.86	10.12	9.60	11.76	14.39
Country of birth Sweden						
F_{PED} (%)	1.19	4.50	5.42	5.81	6.86	17.04
F_{ROH} (%) ^a	1.61	7.39	9.03	8.69	10.98	13.56

^a Results based on the F_{ROH} across all threshold settings

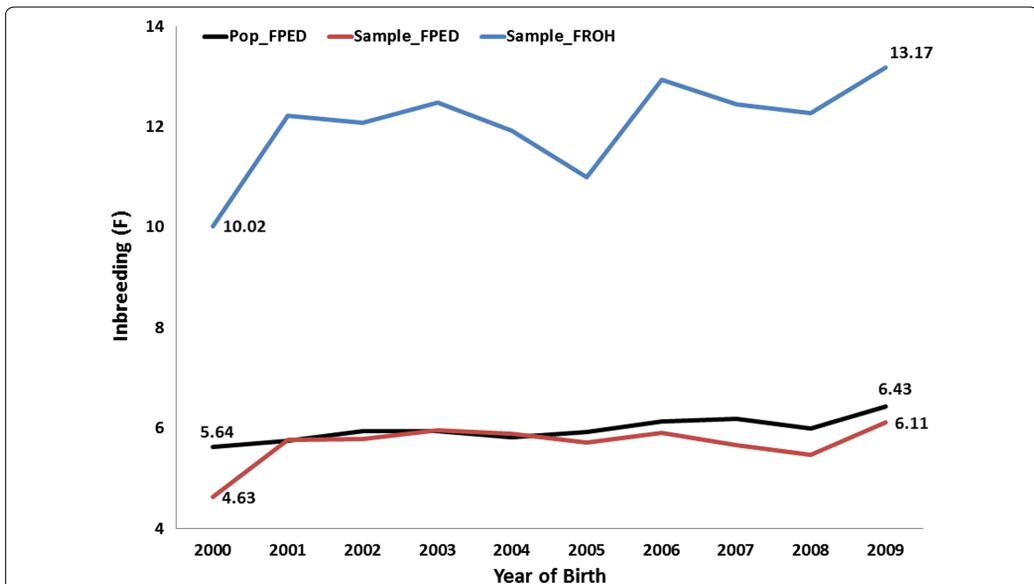


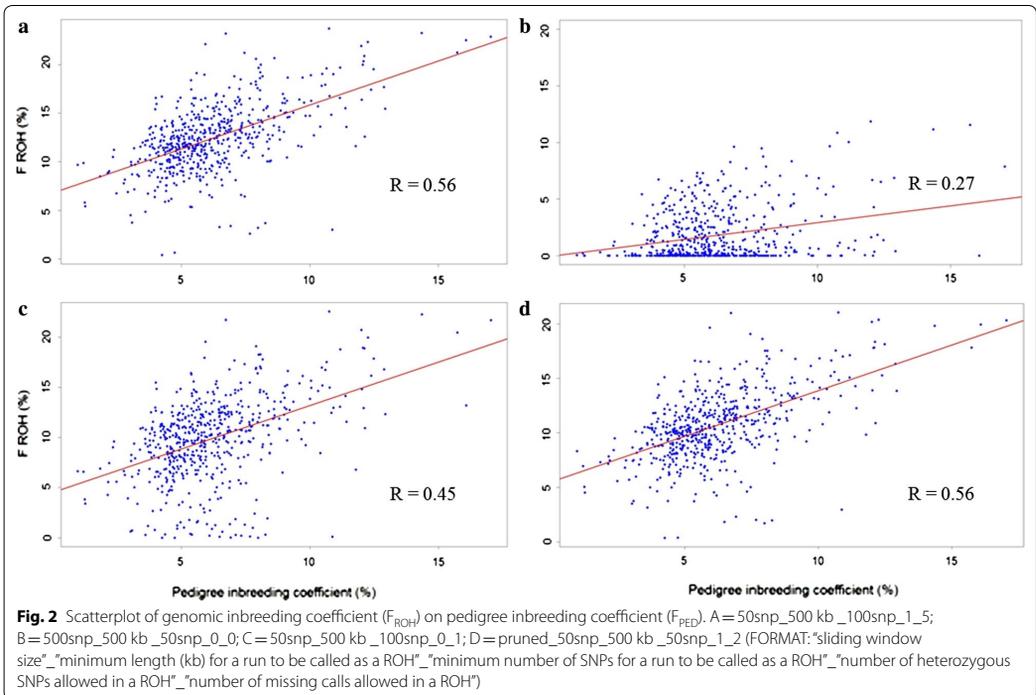
Fig. 1 Median inbreeding levels in the Norwegian–Swedish Coldblooded trotter for horses born between 2000 and 2009. Pop_FPED = pedigree inbreeding for the population ($n = 14,547$); Sample_FPED = pedigree inbreeding and Sample_FROH = genomic inbreeding for the sample of individuals studied here ($n = 566$)

- Pruned for linkage disequilibrium: No.
- Minimum density to consider a ROH: 1 SNP per 50 kb.
- Maximum gap allowed between two SNPs: 100 kb.

Whereas the above settings resulted in the highest correlation between F_{PED} and F_{ROH} , a similarly strong correlation ($R = 0.5594$) was obtained from the analysis of the pruned data with the same threshold settings except that the minimum number SNPs that a run must have to be called as a ROH was set to 50 SNPs.

Paired t test between all F_{ROH} yielded significant differences for most of the F_{ROH} threshold settings with only 35 (1.49%) comparisons resulting in no significant difference (see Additional file 3: Table S3). Variations in sliding window size, minimum length in kb and minimum number of SNPs of a run to be called as a ROH clearly altered F_{ROH} . The influence of different threshold settings on ROH length and ultimately on F_{ROH} is illustrated in Figures S1 and S2 (see Additional file 4: Figure S1 and file 5 Figure S2).

By applying the threshold settings that resulted in the highest correlation between F_{PED} and F_{ROH} , the average



percentage of a single chromosome covered in ROH ranged from 9.84 to 18.82% (Table 3 Column D). Comparisons of ROH between individuals yielded 1403 regions that were present in at least 95% of the sampled horses (Fig. 3). The length of these regions ranged from 1 bp to 935 kb and overlapped with 35 previously characterized QTL for reproduction and health traits (see Additional file 6: Table S4). A visual representation of overlapping regions is in Fig. 3 with a brief description of each overlapped QTL in Table 4. QTL associated with osteochondrosis accounted for 48.6% of the overlapped QTL with only one of the 35 QTL being associated with fertility (QTL 103450, located on *Equus caballus* chromosome (ECA) 1).

Discussion

As expected based on previous studies in other species, the realized F_{ROH} in the NSCT population tended to be slightly higher than the F_{PED} estimates [1–7, 17, 26]. However, in our study, applying strict threshold settings regarding the number of heterozygous SNPs or missing calls allowed in a ROH significantly reduced correlations between F_{PED} and F_{ROH} , and drastically altered the ability to capture longer ROH. Since size and frequency

of ROH provide evidence for relatedness within and between populations, as well as details on distant and recent ancestry, the ability to capture consistently long ROH is essential for the integration of genomic data into breeding evaluation and preservation protocols for the NSCT breed [4–6, 18–20]. Shorter ROH (<1 Mb) tended to be more easily detected regardless of the ROH criteria applied, but longer ROH (>10 Mb) were more difficult to capture when no heterozygous SNPs or missing calls were allowed in a ROH and at least 100 SNPs were required for a run to be called as a ROH. Although this seems logical since a true ROH does not include any heterozygous SNPs, the high-density equine genotyping array contains more than 670,000 SNPs. Even a genotyping error rate of only 1% could yield 6700 possibly incorrectly genotyped SNPs. Since these incorrectly genotyped SNPs, which are likely attributable to poor sample quality in the current study, tend to be randomly scattered across the entire genome, individual horses can be disproportionately affected simply by chance.

Nevertheless, regardless of the ROH threshold settings applied, F_{ROH} in the NSCT breed appears to have steadily increased between 2000 and 2009. While the overall inbreeding level within the breed is slightly

Table 3 Average percentage of the genome, stratified by chromosome, covered by runs of homozygosity (ROH)

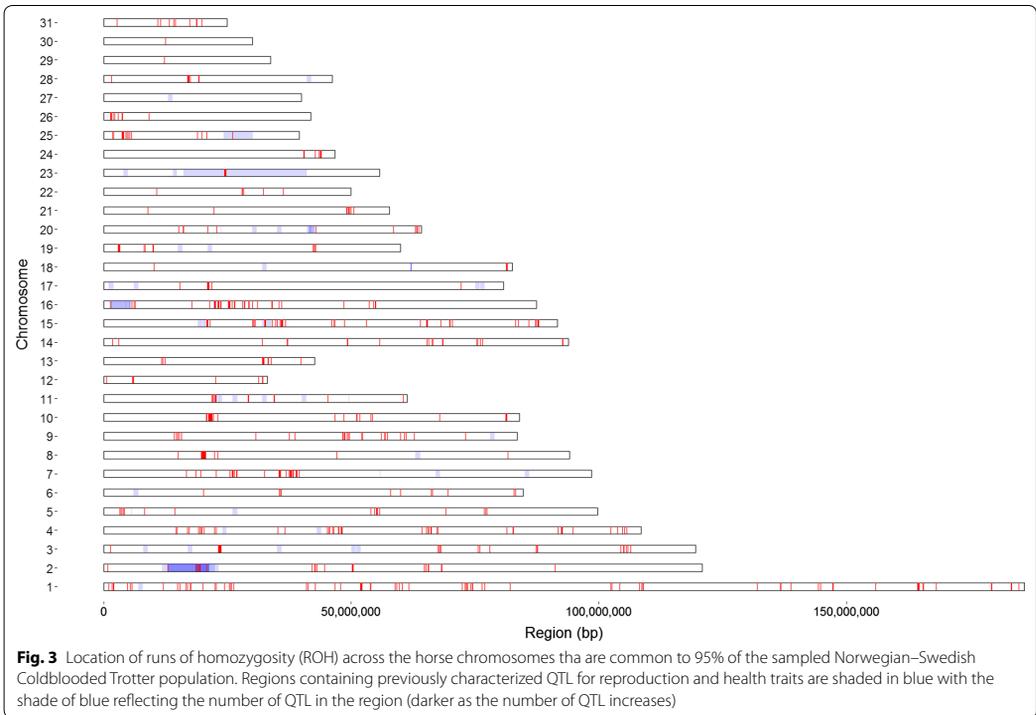
Chromosome	Average ROH (%)							
	A	B	C	D	E	F	G	H
1	10.08	12.67	14.89	15.05	13.59	13.58	7.35	11.34
2	9.40	11.67	13.89	14.08	12.02	12.02	8.62	10.74
3	9.18	11.07	12.66	12.74	12.27	12.24	10.02	9.11
4	10.09	12.27	13.82	13.99	13.16	13.17	11.50	10.61
5	10.38	12.28	14.28	14.46	13.06	13.05	10.62	11.10
6	8.30	10.34	12.58	12.69	10.57	10.57	8.21	10.23
7	10.41	12.23	14.29	14.48	13.12	13.07	9.53	9.58
8	8.61	10.07	11.02	11.05	10.43	10.45	11.73	8.93
9	9.25	10.88	12.90	13.05	11.67	11.65	10.15	8.94
10	9.90	12.09	14.11	14.29	13.05	13.05	11.04	9.87
11	9.29	11.00	12.81	12.84	11.73	11.70	11.76	8.21
12	11.10	12.76	14.74	14.79	12.77	12.73	15.28	11.02
13	12.96	14.70	15.64	15.76	14.43	14.43	14.28	13.06
14	8.96	10.81	12.24	12.34	11.34	11.31	9.85	9.81
15	10.40	12.08	13.36	13.52	12.66	12.68	11.43	10.96
16	13.10	15.72	17.43	17.58	16.45	16.45	12.72	14.17
17	9.19	10.88	11.94	12.01	10.65	10.63	12.94	10.72
18	7.23	8.74	9.78	9.84	8.80	8.81	9.56	8.30
19	9.95	12.08	13.04	13.19	11.51	11.53	13.29	11.15
20	8.11	9.66	11.44	11.54	9.69	9.69	8.43	9.91
21	9.86	11.69	12.67	12.68	11.36	11.36	13.65	11.23
22	11.34	13.04	13.89	13.96	12.34	12.32	16.26	12.59
23	8.29	9.58	10.42	10.58	9.16	9.22	14.82	8.78
24	10.13	11.70	12.87	12.89	11.20	11.19	15.51	11.28
25	12.33	13.44	13.87	13.88	12.24	12.45	19.26	12.93
26	12.48	14.68	15.46	15.56	12.97	12.98	16.99	14.06
27	11.11	12.65	13.64	13.59	12.01	12.00	18.84	12.35
28	10.02	11.60	12.39	12.44	10.91	10.97	14.89	11.07
29	11.28	12.96	14.24	14.40	12.45	12.43	17.81	12.05
30	11.43	13.11	14.24	14.48	11.08	11.08	23.12	12.27
31	14.51	17.03	18.55	18.82	16.25	16.25	22.79	16.81

A = 50snp_500 kb _100snp_0_0; B = 50snp_500 kb _100snp_0_2; C = 50snp_500 kb _100snp_1_2; D = 50snp_500 kb _100snp_1_5; E = 50snp_500 kb _15snp_0_1; F = 50snp_500 kb _50snp_0_1; G = 500snp_500 kb _50snp_0_1; H = pruned_50snp_500 kb _50snp_0_1 (FORMAT: "sliding window size"_"minimum length (kb) for a run to be called as a ROH"_"minimum number of SNPs for a run to be called as a ROH"_"number of heterozygous SNPs allowed in a ROH"_"number of missing calls allowed in a ROH")

underestimated based on classical metrics, the upward trend of inbreeding level revealed by the F_{PED} calculations is clearly supported by the F_{ROH} estimates and likely warrants additional exploration by the NSCT breeding industry—particularly in relation to the difference in inbreeding levels between Norwegian born horses and Swedish born horses (Table 2). Furthermore, it is important to note the difference in F_{PED} between the entire population and the sample of individuals used in our study (Fig. 1). Generally speaking, inbreeding is expected to increase by 1% per generation (i.e. 7–9 years in the NSCT). This 1% increase in inbreeding level is clearly seen in the F_{PED} values for the whole population, but is

not so obvious for the sample of individuals analyzed here, for which a ~1.5% increase was observed instead of the expected 1% over the same time period. Consequently, since the sample of individuals used in our study included only raced horses, although, not certain, it is plausible that the population of raced NSCT is perhaps slightly more inbred than the unraced population.

Although NSCT is not currently considered an at risk breed, it represents unique Norwegian and Swedish genetic resources and is present on the department of agriculture’s list of horse breeds that should be preserved [27]. The NSCT industry has historically been at the forefront regarding the application of emerging



genetic technologies in racehorses, and is currently providing F_{PED} estimates, as well as estimated breeding values (EBV) for breeders and owners to use as part of their criteria for determining sire/dam pairing [28, 29]. While this information has undoubtedly proved valuable over the last half-century, genomic information provides the opportunity to manage NSCT breeding more effectively - particularly if it is used to produce genomic EBV. In addition, the use of genomic information to determine both inbreeding levels and relationships between individuals is also likely to have a knock-on effect on performance, increasing the accuracy of the industry's current EBV and therefore increasing the industry's ability to improve the performance and health of their horses.

As with other species and breeds, opportunities exist for the NSCT industry to develop software programs that provide breeders with easily interpretable feedback on regions of the genome that are suboptimal from the perspective of genetic merit or that are sensitive to inbreeding within the population. Overall, 1403 common ROH regions were identified within the sample of raced horses used here. There were few overlaps with known QTL for health and reproduction traits, which indicates that perhaps only a small percentage of these regions

may warrant concern, at this time [24]. Whereas multiple ROH regions ($n=17$) contained QTL that are associated with osteochondrosis (OC) [30–33], it is possible that homozygosity in these regions may be optimal rather than detrimental when one considers the widely heralded robustness of the breed and that only raced horses were evaluated in our study. It is likely that both the draught horse origins of NSCT and the breeding industry's emphasis on continued production of robust, tractable horses through artificial selection, have resulted in the breed displaying a strong resistance to the development of OC with increasing homozygosity in specific areas of the genome over time. A similar observation can also be made for the common ROH that overlap with QTL associated with recurrent exertional rhabdomyolysis (RER), which is another condition rarely seen in NSCT [34]. However, additional research is required to confirm this.

Increased inbreeding within a population also tends to impact fertility traits unfavorably; however, only one of the common ROH regions overlapped with a known QTL related to reproduction [35], which suggests that, at present, poor fertility may not be a major concern in the NSCT breed. Nevertheless, it is strongly recommended that future genomic studies in this breed

Table 4 Previously reported QTL for reproduction and health trait in the horse that overlap with common (> 95% of the sample) ROH regions in the Norwegian–Swedish Coldblooded Trotter

QTL ID	Symbol	Trait name	Chr	Start position (bp)	End position (bp)
103450	MOTSCT	Number of motile sperm	1	53958169	53958209
32144	OSTEO	Osteochondrosis	2	11816213	21391792
32142	OSTEO	Osteochondrosis	2	12910010	21391792
32146	OSTEO	Osteochondrosis	2	12910010	21391792
32145	OSTD	Osteochondrosis dissecans	2	13028376	22500086
32143	OSTEO	Osteochondrosis	2	18664801	19717761
32147	OSTEO	Osteochondrosis	2	18664801	23235964
29325	OSTEO	Osteochondrosis	3	105163057	105163097
29326	OSTEO	Osteochondrosis	3	105546982	105547022
29327	OSTEO	Osteochondrosis	3	105830605	105830645
29306	SUSBITE	Insect bite hypersensitivity	4	43000811	43945687
37902	OSTD	Osteochondrosis dissecans	5	77424966	77425006
29287	SUSBITE	Insect bite hypersensitivity	11	22016942	22985500
29268	RHOD	Rhodococcus equi infection	14	3055253	3055293
29315	SUSBITE	Insect bite hypersensitivity	15	20012397	20994475
29316	SUSBITE	Insect bite hypersensitivity	15	32000266	32987009
29035	GPT	Guttural pouch tympany	15	53093059	53093099
29111	GPT	Guttural pouch tympany	15	65298904	65298944
29067	GPT	Guttural pouch tympany	15	78013499	78013539
28922	OSTEO	Osteochondrosis	16	1299549	5389006
29002	OSTD	Osteochondrosis dissecans	16	1299549	5389006
28933	OSTEO	Osteochondrosis	16	5228939	5496903
28999	OSTD	Osteochondrosis dissecans	16	5228939	5496903
28927	OSTD	Osteochondrosis dissecans	16	22275834	22702331
28937	OSTEO	Osteochondrosis	16	22275834	22702331
29006	OSTEO	Osteochondrosis	16	22275834	22702331
29338	RER	Recurrent exertional rhabdomyolysis	16	29314251	29314291
29277	RER	Recurrent exertional rhabdomyolysis	16	29349222	29349262
29337	RER	Recurrent exertional rhabdomyolysis	16	29349222	29349262
29320	SUSBITE	Insect bite hypersensitivity	19	21037979	21977304
29298	SUSBITE	Insect bite hypersensitivity	20	41031989	41982509
37895	SUSBITE	Insect bite hypersensitivity	20	41530793	42603867
37896	SUSBITE	Insect bite hypersensitivity	20	41530793	42603867
28920	SARRESI	Equine sarcoids	23	16126529	41049320
28921	SARRESI	Equine sarcoids	25	24227654	30109054

should consider the inclusion of data on fertility traits, since it will likely prove to be highly beneficial in subsequent efforts to preserve the breed's genetic variability in the long term [5, 36].

Conclusions

In the current study, both F_{PED} and F_{ROH} were calculated for a sample of raced NSCT with F_{ROH} resulting in higher inbreeding coefficients, and both methods showing a gradual increase in inbreeding between 2000 and 2009. Stricter ROH threshold criteria regarding the

number of heterozygous SNPs and missing calls allowed in a ROH significantly reduced correlations between F_{PED} and F_{ROH} and noticeably altered the chances of capturing long ROH. While the exact reasons behind this decrease in correlations are not known with certainty, the established associations between classical F estimates and recent inbreeding within a pedigree (characterized by long ROH) in other species provide some insight. Since retaining genetic variation is important to allow populations to adapt to changing environments, the integration of genomic data into their EBV and the use of molecular

data to identify both genomic regions contributing to inbreeding depression and pedigree errors will likely prove invaluable as the NSCT industry moves forward in its conservation and selection efforts.

Additional files

Additional file 1: Table S1. Threshold settings used to define runs of homozygosity in PLINK and the corresponding average F_{ROH} for a sample of raced Norwegian-Swedish Coldblooded trotters born between 1 January 2000 and 31 December 2009.

Additional file 2: Table S2. Correlation matrix between F_{PED} and all F_{ROH} estimates.

Additional file 3: Table S3. Results of the paired t-test (P-values) between F_{PED} and all F_{ROH} estimates.

Additional file 4: Figure S1. Histograms for run of homozygosity (ROH) lengths based on four different threshold combinations in PLINK v 1.07. A = 50snp_500kb_100snp_0_0; B = 50snp_500kb_100snp_0_2; C = 50snp_500kb_100snp_1_2; D = 50snp_500kb_100snp_1_5 (FORMAT: "sliding window size"_"minimum length (kb) to be called as homozygous"_"minimum number of SNPs to be called as homozygous"_"number of heterozygotes allowed"_"number of missing calls allowed").

Additional file 5: Figure S2. Histograms for run of homozygosity (ROH) lengths based on varying window size thresholds in PLINK v 1.07. A = 50snp_500kb_15snp_0_1; B = 50snp_500kb_50snp_0_1; C = 500snp_500kb_50snp_0_1; D = pruned_50snp_500kb_50snp_0_1 (FORMAT: "sliding window size"_"minimum length (kb) for a run to be called as a ROH"_"minimum number of SNPs for a run to be called as a ROH"_"number of heterozygous SNPs allowed in a ROH"_"number of missing calls allowed in a ROH").

Additional file 6: Table S4. Homozygous regions of the genome that are shared by at least 95% of the sample of Norwegian-Swedish Coldblooded Trotters (n = 566).

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Authors' contributions

BDV, KJF, CI, ES, and GL conceived and designed the experiments; KJF, MKR, and KHR contributed to sampling. GL and ES contributed the reagents and MKR extracted the DNA; BDV analyzed the data and drafted the manuscript; KJF, MS, CI, ES, and GL discussed and contributed to data analysis. All authors read and approved the final manuscript.

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Availability of data and materials

The data that support the findings of this study are available from the Swedish Trotter Association (Stockholm, Sweden) and the Norwegian Trotter Association (Oslo, Norway), but restrictions apply to the availability of these data, which were used under license for the current study, and so are not publicly available. However, data are available from the authors upon reasonable request and with permission of the Swedish Trotter Association (Stockholm, Sweden) and the Norwegian Trotter Association (Oslo, Norway).

Ethics approval and consent to participate

All experimental procedures and sample collection methods were approved by the Ethics Committee for Animal Experiments in Uppsala, Sweden [Number: C 121/14]. Samples used in this study were already available at either the Animal Genetics Laboratory at SLU in Uppsala, Sweden or the Department of Basic Sciences and Aquatic Medicine at the Norwegian University of Life Sciences in Oslo, Norway, since they had been previously used for parentage testing. Permission to use the samples was granted from the Swedish Trotting Association and the Norwegian Trotting Association (the owners of the samples per the rules/guidelines of the industry).

Consent for publication

Not applicable.

Competing interests

The authors have the following interests: GL is a co-inventor on a granted patent concerning commercial testing of the DMRT3 mutation: A method to predict the pattern of locomotion in horses. PCT EP 12,747,875.8. European patent registration date: 2011-05-05, US patent registration date: 2011-08-03. There are no further patents, products in development, or marketed products to declare.

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References

- Solé M, Gori A, Faux P, Bertrand A, Farnir F, Gautier M, et al. Age-based partitioning of individual genomic inbreeding levels in Belgian Blue cattle. *Genet Sel Evol*. 2017;49:92.
- Luan T, Yu X, Dolezal M, Bagnato A, Meuwissen THE. Genomic prediction based on runs of homozygosity. *Genet Sel Evol*. 2014;46:64.
- Pryce JE, Haile-Mariam M, Goddard ME, Hayes BJ. Identification of genomic regions associated with inbreeding depression in Holstein and Jersey dairy cattle. *Genet Sel Evol*. 2014;46:71.
- Howard JT, Pryce JE, Baes C, Maltecca C. Inbreeding in the genomics era: inbreeding, inbreeding depression, and management of genomic variability. *J Dairy Sci*. 2017;100:6009–24.
- Peripolli E, Munari DP, Silva MVGB, Lima ALF, Irgang R, Baldi F. Runs of homozygosity: current knowledge and applications in livestock. *Anim Genet*. 2017;48:255–71.
- Kim ES, Sonstegard TS, Van Tassel CP, Wiggans G, Rothschild MF. The relationship between runs of homozygosity and inbreeding in Jersey cattle under selection. *PLoS One*. 2015;10:e0129967.
- Kardos M, Akesson M, Fountain T, Flagstad O, Loberg O, Olason P, et al. Genomic consequences of intensive inbreeding in an isolated wolf population. *Nat Ecol Evol*. 2018;2:124–31.
- Ayroles JF, Hughes KA, Rowe KC, Reedy MM, Rodriguez-Zas SL, Druvich JM, et al. A genome-wide assessment of inbreeding depression: gene number, function, and mode of action. *Conserv Biol*. 2009;23:920–30.
- Charlesworth D, Willis JH. The genetics of inbreeding depression. *Nat Rev Genet*. 2009;10:783–96.
- Ponsart C, Le Bourhis D, Knijn H, Fritz S, Guyader-Joly C, Otter T, et al. Reproductive technologies and genomic selection in dairy cattle. *Reprod Fertil Dev*. 2013;26:12–21.
- Jäderkvist Fegraeus K, Velie BD, Axelsson J, Ang R, Hamilton NA, Andersson L, et al. A potential regulatory region near the EDN3 gene may control both harness racing performance and coat color variation in horses. *Physiol Rep*. 2018;6:e13700.
- Klemetsdal G. Norwegian trotter breeding and estimation of breeding values. In: State of breeding evaluation in trotters. In Proceedings of the

- European Federation of Animal Science Symposium on the Commission on Horse Production: 27 June–1 July 1988; Helsinki; 1989. pp 95–105.
13. Arnason T, Bendroth M, Philipsson J, Henriksson K, Darenius A. Genetic evaluation of Swedish trotters. In State of breeding evaluation in trotters. In Proceedings of the European Federation of Animal Science Symposium on the Commission on Horse Production: 27 June–1 July 1988; Helsinki; 1989. pp 106–30.
 14. Svensk Travsport. 2018. <https://www.travsport.se/wicket/bookmarkable/se.atg.web.travsport.page.polopoly.ArticlePage?6&cid=2.255>. Accessed 1 Sept 2018.
 15. Klemetsdal G. The effect of inbreeding on racing performance in Norwegian cold-blooded trotters. *Genet Sel Evol*. 1998;30:351–66.
 16. Sargolzaei M, Iwaisaki H, Colleau JJ, CFC, A tool for monitoring genetic diversity. In: Proceedings of the 8th World Congress on Genetics Applied to Livestock Production: 13–18 August 2006; Belo Horizonte; 2006.
 17. Forneris NS, Steibel JP, Legarra A, Vitezica ZG, Bates RO, Ernst CW, et al. A comparison of methods to estimate genomic relationships using pedigree and markers in livestock populations. *J Anim Breed Genet*. 2016;133:452–62.
 18. Druml T, Neuditschko M, Grilz-Seiger G, Horna M, Ricard A, Mesaric M, et al. Population networks associated with runs of homozygosity reveal new insights into the breeding history of the Haflinger horse. *J Hered*. 2018;109:384–92.
 19. Keller MC, Visscher PM, Goddard ME. Quantification of inbreeding due to distant ancestors and its detection using dense single nucleotide polymorphism data. *Genetics*. 2011;189:237–49.
 20. Ferencakovic M, Solkner J, Curik I. Estimating autozygosity from high-throughput information: effects of SNP density and genotyping errors. *Genet Sel Evol*. 2013;45:42.
 21. Metzger J, Karwath M, Tonda R, Beltran S, Agueda L, Gut M, et al. Runs of homozygosity reveal signatures of positive selection for reproduction traits in breed and non-breed horses. *BMC Genomics*. 2015;16:764.
 22. Kaminski S, Hering DM, Jaworski Z, Zabolewicz T, Rusc A. Assessment of genomic inbreeding in Polish Konik horses. *Pol J Vet Sci*. 2017;20:603–5.
 23. R Development Core Team: R-A Language and environment for statistical computing; 2018 <https://www.r-project.org/>. Accessed 1 April 2018.
 24. Hu ZL, Park CA, Reecy JM. Developmental progress and current status of the Animal QTLdb. *Nucleic Acids Res*. 2016;44:D827–33.
 25. Neph S, Kuehn MS, Reynolds AP, Haugen E, Thurman RE, Johnson AK, et al. BEDOPS: high-performance genomic feature operations. *Bioinformatics*. 2012;28:1919–20.
 26. Kardos M, Luikart G, Allendorf FW. Measuring individual inbreeding in the age of genomics: marker-based measures are better than pedigrees. *Heredity* (Edinb). 2015;115:63–72.
 27. Department of Agriculture. Delmål för husdjursgenetiska resurser åren 2010 till 2020; 2009. <http://www.jordbruksverket.se/download/18.2958036f1211efb32978000308/rapport%2B070511.pdf>. Accessed 1 Sept 2018.
 28. Svensk Travsport; 2018 https://www.travsport.se/artikel/kvalificerings-_&_premielopp. Accessed 1 Sept 2018.
 29. Det Norske Travelskap; 2018 <https://www.travsport.no>. Accessed 1 Sept 2018.
 30. Dierks C, Komm K, Lampe V, Distl O. Fine mapping of a quantitative trait locus for osteochondrosis on horse chromosome 2. *Anim Genet*. 2010;41:S87–90.
 31. Orr N, Hill EW, Govindarajan P, Conroy J, van Grevenhof EM, Ducro BJ, et al. Genome-wide association study of osteochondrosis in the tarsocrural joint of Dutch Warmblood horses identifies susceptibility loci on chromosomes 3 and 10. *Anim Genet*. 2013;44:408–12.
 32. Lykkjen S, Dolvik NI, McCue ME, Rendahl AK, Mickelson JR, Roed KH. Genome-wide association analysis of osteochondrosis of the tibiotarsal joint in Norwegian Standardbred trotters. *Anim Genet*. 2010;41:S111–20.
 33. Lampe V, Dierks C, Distl O. Refinement of a quantitative gene locus on equine chromosome 16 responsible for osteochondrosis in Hanoverian warmblood horses. *Animal*. 2009;3:1224–31.
 34. Fritz KL, McCue ME, Valberg SJ, Rendahl AK, Mickelson JR. Genetic mapping of recurrent exertional rhabdomyolysis in a population of North American Thoroughbreds. *Anim Genet*. 2012;43:730–8.
 35. Gottschalk M, Metzger J, Martinsson G, Sieme H, Distl O. Genome-wide association study for semen quality traits in German Warmblood stallions. *Anim Reprod Sci*. 2016;171:81–6.
 36. de Simoni Gouveia JJ, da Silva MVGB, Paiva SR, de Oliveira SMP. Identification of selection signatures in livestock species. *Genet Mol Biol*. 2014;37:330–42.

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RESEARCH ARTICLE

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Exploring the genetics of trotting racing ability in horses using a unique Nordic horse model

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Abstract

Background: Horses have been strongly selected for speed, strength, and endurance-exercise traits since the onset of domestication. As a result, highly specialized horse breeds have developed with many modern horse breeds often representing closed populations with high phenotypic and genetic uniformity. However, a great deal of variation still exists between breeds, making the horse particularly well suited for genetic studies of athleticism. To identify genomic regions associated with athleticism as it pertains to trotting racing ability in the horse, the current study applies a pooled sequence analysis approach using a unique Nordic horse model.

Results: Pooled sequence data from three Nordic horse populations were used for F_{ST} analysis. After strict filtering, F_{ST} analysis yielded 580 differentiated regions for trotting racing ability. Candidate regions on equine chromosomes 7 and 11 contained the largest number of SNPs ($n = 214$ and 147 , respectively). GO analyses identified multiple genes related to intelligence, energy metabolism, and skeletal development as potential candidate genes. However, only one candidate region for trotting racing ability overlapped a known racing ability QTL.

Conclusions: Not unexpected for genomic investigations of complex traits, the current study identified hundreds of candidate regions contributing to trotting racing ability in the horse. Likely resulting from the cumulative effects of many variants across the genome, racing ability continues to demonstrate its polygenic nature with candidate regions implicating genes influencing both musculature and neurological development.

Keywords: Athleticism, Conformation, Genomic, Performance, Racehorse

Background

As genomics improves and enables the design of more targeted studies relating genotypes to phenotypes, the opportunity for non-model organisms continues to expand - facilitating greater opportunities to gain novel insight into the mechanisms regulating biological homeostasis and health [1, 2]. Genomic studies of natural model species, domestic species in particular, give a complimentary view of genotype-phenotype relationships compared with

the knowledge gained from the study of humans and experimental organisms [2]. Since the onset of domestication, horses have been strongly selected for, among other things, speed, strength, and endurance-exercise traits [3]. This diverse and, at times, divergent selection has ultimately led to the development of highly specialized horse breeds. Within the last 400 years, breed specialization has focused primarily on preserving and improving traits related to aesthetics and performance [3]. As a result, most horse breeds today are closed populations with high phenotypic and genetic uniformity within breed. However, a great deal of variation continues to exist among breeds [3]. This variation, combined with breed specialization, has made the horse particularly well suited for genetic studies of locomotion patterns and provides a unique opportunity for genetic studies of

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athleticism [1–5]. Generally speaking, athleticism describes the physical qualities that are characteristic of athletes and typically refers to traits such as strength, fitness, and agility. Many modern day horse breeds exemplify some if not all of these traits, with shared selective pressures within breeds (e.g. health, fertility traits, conformation) and divergent selection between breeds (e.g. speed vs strength) yielding a wide range of athletic phenotypes [3, 6].

Here we apply a genomic approach to investigate athletic phenotypes associated with trotting racing ability (TRA) using a unique Nordic horse model consisting of the Norwegian-Swedish Coldblooded trotter (NSCT), the North Swedish Draught horse (NSD), and the Standardbred trotter (SB) (Fig. 1). Although both the NSCT and the NSD are horse breeds derived from the original North-Swedish horse, a small, heavy horse traditionally used in agriculture and forestry work, selection for traits beneficial to agricultural work only continues in the NSD [7–11]. Since the 1960s, the NSCT has been intensively selected for harness racing performance and is now considered a true racing breed [10–13]. As a result, a remarkable improvement in the racing performance of NSCTs has occurred during the last half-century. However, it is also well established that some degree of cross-breeding occurred between NSCT and SBs, a significantly faster breed of horse from a different gene pool, before obligatory paternity testing was introduced in Sweden in 1969 [10–13]. Consequently, the improvement in NSCT racing performance may be partially explained by a marked increase of favorable genetic variants originating from SBs. It is this specific attempt at gaining a competitive racing advantage that makes the NSCT ideal for genomic studies investigating TRA phenotypes.

Despite a dispersed history of crossbreeding with SBs, the relationship between the NSCT and NSD remains closer than either of the breeds with the SB [10]. While both the NSCT and the SB are selected for racing performance, the Norwegian and Swedish breed organizations have remained highly committed to preserving the historical work-horse appearance of the NSCT breed [7, 8]. As a result, both NSCTs and NSDs can be classified as heavy horse breeds, with NSCTs sometimes referred to as “draft trotters” (Fig. 1) [14]. Any lingering genetic similarities between the NSCT and the SB are therefore highly likely to be associated with favorable traits for TRA. Our aim was to identify these similarities using pooled whole genome sequence data from a carefully selected sample of NSCTs, NSDs, and SBs.

Results

A summary of the population statistics for the three populations is provided in Tables 1 and 2.

F_{ST} analyses

After clustering of windows located less than 0.1 Mb apart and filtering out regions containing less than 2 SNPs or

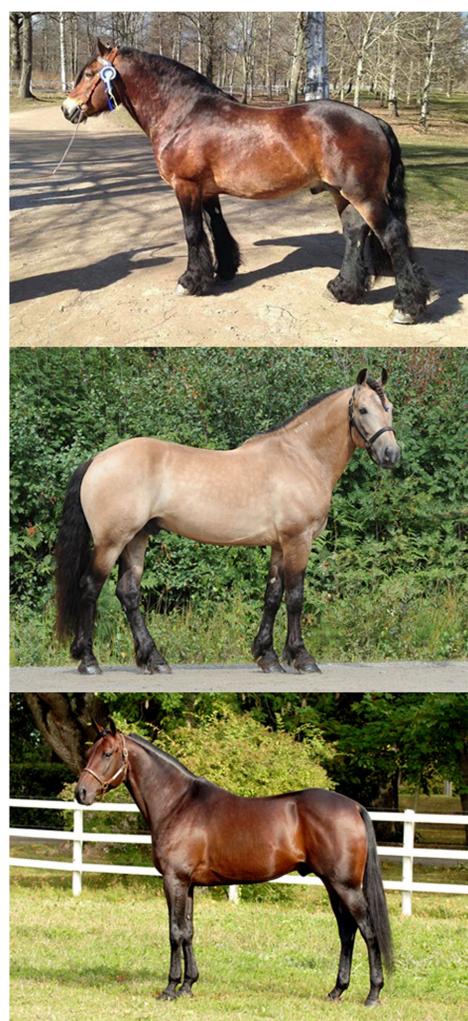


Fig. 1 Breed classifications. Top panel = North Swedish Draught Horse; middle panel = Norwegian-Swedish Coldblooded trotter; bottom panel = Standardbred trotter. *Images used in the figure are privately owned and thus were not taken from previously published sources requiring written permission for use

windows measuring only 1000 bp, 580 differentiated regions were retained (Fig. 2; Additional file 1). Candidate regions ranged in length from 1.5 kb to 773.5 kb (mean/median length: 80/56 kb; total cumulative length: 46.389 Mb).

Candidate regions on *Equus caballus* chromosome (ECA) 7 and 11 contained the largest number of SNPs at 214 (length: 151 kb) and 147 (length: 683.5 kb), respectively. Multiple regions on ECA 11, 13, and 15 were greater than

Table 1 Pooled population samples and genome information

Breed	Number of individuals ^a	Coverage	Nucleotide diversity (π) (%) ^b
Norwegian-Swedish Coldblooded trotter	18	43.98	0.171
North Swedish Draught	25	35.96	0.175
Standardbred trotter	22	56.26	0.159

^aMale/Female ratio: Norwegian-Swedish Coldblooded trotter = 2/1; North Swedish Draught = 25/0; Standardbred trotter = 7/4

^bNucleotide diversity calculated over 5000 bp windows, averaged across the genome (%)

500 kb in length (Fig. 2; Additional file 1). From GO analysis, candidate regions identified contained 271 candidate genes associated with known molecular functions, 519 with known biological processes, and 124 with known pathways (Figs. 3 and 4; Additional files 2 and 3). One candidate region for TRA overlapped with a previously characterized QTL for racing ability [15]. This region was located on ECA1 and overlapped a QTL for racing speed (Fig. 2; Table 3).

Discussion

For all intents and purposes, racehorses are professional athletes. Like professional human athletes, racehorses must not only endure the day to day physical demands of their sport, but they must also have a genetic capacity for athleticism relative to their sport in order to ultimately achieve success (i.e. win). However, unlike in humans, racehorses have been carefully selected and bred for centuries, resulting in alleles with subtle effects on athleticism being enriched over time. As a result, racehorses in particular provide a unique opportunity to identify genes and subsequently the molecular mechanisms underpinning athletic ability. Rarely found outside of the Nordic countries, the NSCT is perhaps one of the most unique types of racehorse in the modern era – originating not from historic racing breeds such as the Thoroughbred or Standardbred, but instead tracing its lineage back to the original North Swedish horse [10, 11]. Using whole-genome re-sequencing of pooled DNA from a carefully selected group of NSCTs, NSDs, and SBs, we capitalized on this unusual ancestry of the NSCT and identified 580 candidate regions for TRA in the horse.

Only one previously characterized QTL for racing ability was overlapped by the TRA candidate regions in the current study [15]. A SNP in the first intron of the sortilin related VPS10 domain containing receptor 3 (*SORCS3*) gene, previously associated with racing speed in endurance horses, was overlapped by a 107.5 kb candidate region on

ECA1 [15]. The gene encodes for a type-I transmembrane receptor protein that is a member of family of receptors with known pleiotropic functions [16]. Genetic variation in *SORCS3* has been associated with Alzheimer’s disease in humans with more recent studies suggesting that additive epistatic effects of genetic variants within the gene may be important [17–19]. Furthermore, variation in *SORCS3* has been associated with attention deficit hyperactivity disorder and *SORCS3* knockout mice display defects in spatial learning and memory, as well as increased fear extinction [20, 21]. Although the function of this gene in horses remains unknown, as the transcript is generally expressed at high levels in the brain, it could perhaps alter an individual’s perception of athletic competition (e.g. altered feedback loop in response to exercise, reduced fear extinction) [20].

Interestingly, while a mere 1.9% (46.39 Mb) of the genome was covered by TRA candidate regions, the regions on ECA11 not only accounted for 11.2% of all TRA candidate regions identified, but ECA11 candidate regions also contained some of the highest number of SNPs ($n = 100+$) and the largest sweeps (> 50 kb in length). Furthermore, ECA11 had the highest concentration of candidate TRA regions when compared to the other equine chromosomes. However, despite the density of this TRA signal, no QTLs for racing ability have been mapped to ECA11 [22].

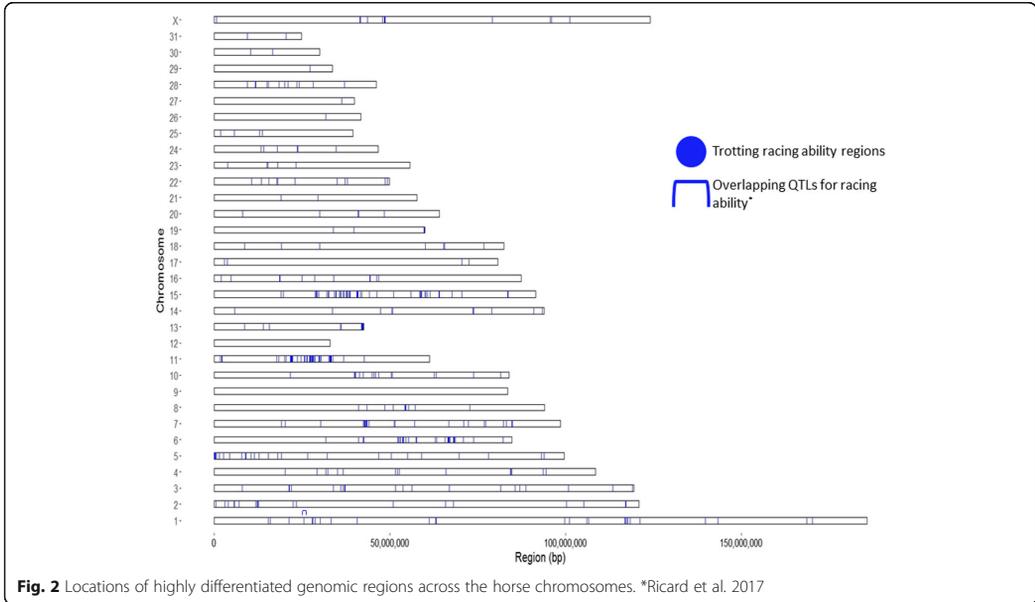
Consequently, regions on ECA11 have previously been associated with size (i.e. height and mass), which, given the prudent design of the current study, is particularly interesting [3, 23, 24]. Although similar in height, NSCTs, NSDs, and SBs differ in their physique. SBs tend to be leaner and more refined in their appearance compared with NSCTs, while NSCTs tend to be leaner and more refined than NSDs (Fig. 1). In order for a region to have been considered as a candidate region for TRA in the current study, the region had to be highly similar between NSCTs and SBs yet decidedly different from NSDs. It is possible that strict adherence to include only top performing NSCTs in the sequenced pool may have skewed the NSCT pool towards lighter framed horses; thereby demonstrating what conceivably is a competitive racing advantage for lighter horses. Perhaps even more interesting to note is that no previously reported QTLs for growth or conformation traits overlapped any of the candidate regions for TRA on ECA11; however, a recent study in American Quarter

Table 2 Differentiation between breeds

	NSD	NSCT	SB
NSD		0.07159	0.11077
NSCT	0.05755		0.10819
SB	0.08823	0.08555	

Mean F_{ST} above the diagonal, median F_{ST} below the diagonal

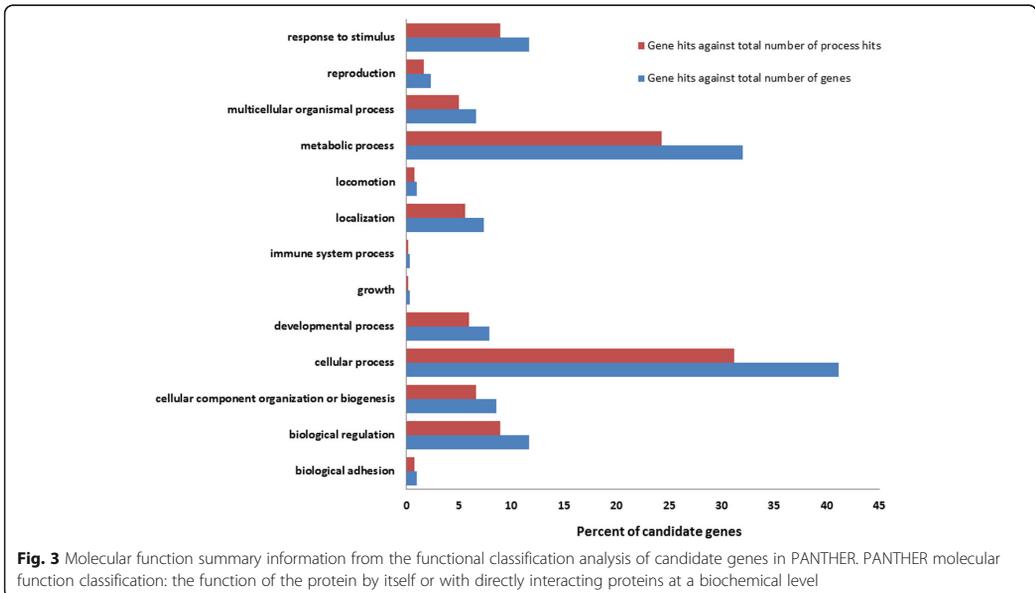
NSD North Swedish Draught Horse, NSCT Norwegian-Swedish Coldblooded Trotter, SB Standardbred trotter

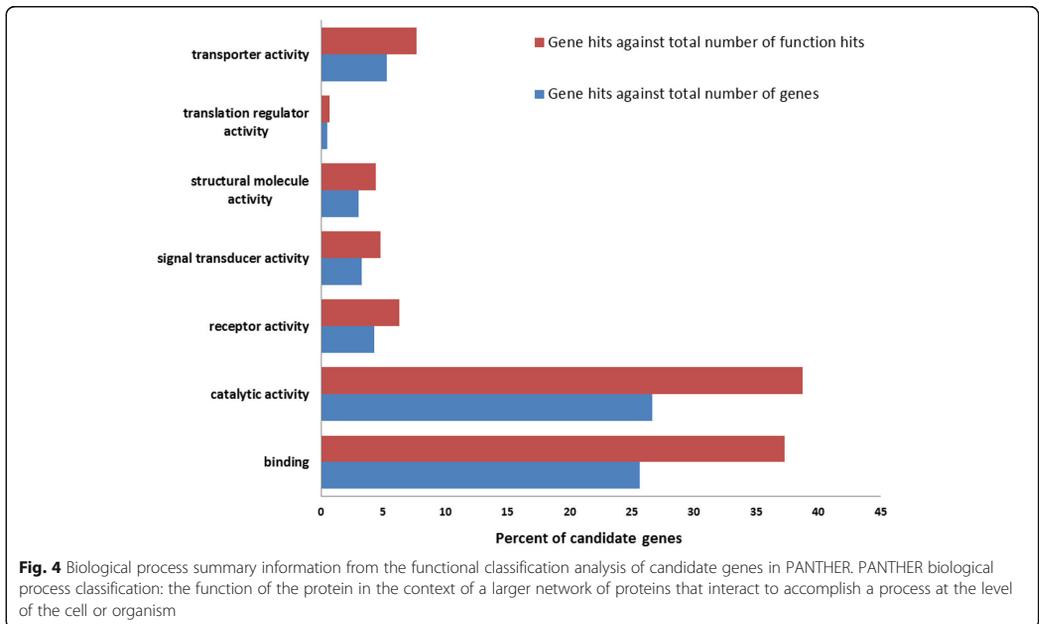


horses also suggested ECA11 as potentially important for racing ability [25].

ECA11 also contained the candidate region with the second largest number of SNPs ($n = 147$) in the study. The region, located ECA11:32,874,000-33,557,500, encompasses 9

genes, (RAD51C, PPM1E, ENSECAG0000003590, ENSECAG00000015244, GDPD1, YPEL2, SKA2, PRR11) one of which is tripartite motif containing 37 (*TRIM37*). Mutations in *TRIM37* are associated with Mulibrey Nanism in humans, an extremely rare autosomal recessive disorder





characterized by profound growth delays and abnormalities of the muscles, liver, brain, and eyes [26, 27]. Instinctively this would further support the candidate region being associated with body size; however, even if this is the case, it does not necessarily mean the region is solely associated with body size and shape. It is highly plausible that haplotypes associated with body size differ by multiple substitutions with pleiotropic functional effects. Mutations that impact underlying mechanisms for muscle, ligament, and tendon development would certainly influence TRA – limiting racing ability in some instances, while enhancing racing ability in others [28–30]. Moreover, a large conserved haplotype containing tripartite motif containing 13 (*TRIM13*), a gene located on ECA17, has previously been suggested as having selective importance in the Thoroughbred [3]. *TRIM37*, while located on a different chromosome, is part of the same gene family as *TRIM13*.

Conclusions

This study identified hundreds of candidate genomic regions contributing to TRA in the horse, a result not unexpected for investigations into the genomics of such a

complex trait. The trait is undoubtedly polygenic, resulting from the cumulative effects of many variants across the genome. Candidates for TRA implicated both genes influencing musculature and conformation, as well as genes involved in neurological development, further suggesting that racing ability may not solely be a product of physical characteristics, but also mental characteristics. This study identified a strong racing ability signal on ECA11 that will be particularly interesting for follow-up.

Methods

Animals

Genomic DNA samples from 18 NSCTs, 25 NSDs, and 22 SBs were prepared from blood samples and pooled in equimolar ratios prior to library construction (Table 1). Each horse was selected based on strict breed specific criteria. All trotting horses racing in Norway and Sweden have breeding values estimated annually. For NSCTs, estimated breeding values (EBVs) are estimated using an animal model that includes the combined effect of country, sex, and birth year. EBVs are subsequently based on racing performance results (i.e. racing status

Table 3 QTLs overlapped by trotting racing ability candidate regions

QTL ID	ECA	Start Position	End Position	Trait Name	Study
138,687	1	25,715,314	25,715,354	racing speed	Ricard et al. 2017

ECA Equus caballus chromosome

and earnings) occurring between 3 and 6 years of age [31]. For inclusion in the current study, NSCT horses were required to have an estimated breeding value of at least 115 and sire/progeny ratios were restricted to reflect the larger population as accurately as possible (Additional file 4) [7, 8].

For EBVs in SBs, the animal model includes genetic base group and a combination of sex and birth year with the evaluation based on racing performance results occurring between 2 and 5 years of age [31]. For inclusion in the current study, Standardbreds were also required to have estimated breeding values of at least 115 and both SBs and NSDs were not allowed to have a common ancestor within three generations (i.e. no shared sires, dams, grandsires, granddams) [7, 9]. An EBV requirement for NSDs was not possible as EBVs are not calculated for this breed.

Pool sequencing, genome alignments, variant calling, and population analyses

Genome sequencing library construction and sequencing was carried out by SciLifeLab (Uppsala, Sweden) using two lanes on the Illumina HiSeq2500 (150 bp paired-end). Sequencing libraries were prepared from 100 ng DNA using the TruSeq Nano DNA sample preparation kit targeting an insert size of 350 bp. Reads were aligned to the *Equus caballus* genome (EquCab2.70) using BWA (v0.7.15) [32]. Duplicates were marked with Picard (v1.118; <http://broadinstitute.github.io/picard/>) and GATK was used for realignment around indels [33]. Samtools (v1.8 [34, 35]) was used to generate the mpileup files needed for Popoolation (v1.2.2) and PoPoolation2 (v1.201) [36]. Nucleotide diversity (π) was calculated across 5000 bp windows for each population pool using Popoolation [36]. PoPoolation2 was used to calculate F_{ST} over 1000 bp sliding windows with 50% overlap between the selected population samples using the Karlsson et al. method [36, 37]. Minimum count was set at 3, minimum coverage at 10, maximum coverage at 100, and minimum coverage fraction at 1.

Differentiated regions

Given the close relationship between NSCTs and NSDs, candidate regions for athletic traits were defined as genomic regions where F_{ST} values were relatively high between NSCTs and NSDs, but low between NSCTs and SBs. As such, stringent F_{ST} cutoffs ($> 95\%$ percentile, $F_{ST} = 0.179$ NSCT vs. NSD; $< 5\%$ percentile, $F_{ST} = 0.013$ NSCT vs. SB) were used when defining candidate regions. Windows with F_{ST} values that met these criteria were clustered into candidate sweep regions when they were less than 0.1 Mb from one another (custom R scripts) [38]. Clusters containing only a single 1000 bp window or less than 2 SNPs were excluded. Candidate gene screening was subsequently carried

out using the bioinformatics database Ensembl (<http://www.ensembl.org/>). Candidate regions from the F_{ST} analyses were used to generate a list of annotated genes using the Ensembl Biomart function. The resulting list of candidate genes was then piped into the PANTHER Classification system in order to obtain an overview of the molecular functions and biological processes affected by the candidate genes [39, 40]. Previously reported racing ability QTLs in the horse (downloaded from the horse QTL database; [22]) were also compared to differentiated regions to determine overlaps using bed file comparisons in BEDOPS [41].

Additional files

Additional file 1: Summary information for trotting racing ability candidate sweep regions. (XLSX 53 kb)

Additional file 2: Functional classification gene list from PANTHER analysis of trotting racing ability candidate sweep regions. (XLSX 42 kb)

Additional file 3: Pathway summary information from the functional classification analysis of trotting racing ability candidate sweep regions. (XLSX 12 kb)

Additional file 4: Pedigree breakdown for the 18 Norwegian-Swedish Coldblooded trotters (NSCT) included in the NSCT pool. (XLSX 11 kb)

Abbreviations

EBVs: Estimated breeding values; ECA: *Equus caballus* chromosome; GDDP1: Glycerophosphodiester phosphodiesterase domain containing 1; NSCT: Norwegian-Swedish Coldblooded trotter; NSD: North Swedish Draught horse; PPM1E: Protein phosphatase Mg²⁺/Mn²⁺ dependent 1E; PRR11: Proline rich 11; RAD51C: RAD51 paralog C; SB: Standardbred trotter; SKA2: Spindle and kinetochore associated complex subunit 2; SORCS3: Sortilin related VPS10 domain containing receptor 3; TRA: Trotting racing ability; TRIM13: Tripartite motif containing 13; TRIM37: Tripartite motif containing 37; YPEL2: Yippee like 2

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Availability of data and materials

The data that support the findings of this study are available from the Swedish Trotting Association (Stockholm, Sweden) and the Norwegian Trotting Association (Oslo, Norway), but restrictions apply to the availability of these data, which were used under license for the current study, and so are not publicly available. However, data are available from the authors upon reasonable request and with permission of the Swedish Trotting Association (Stockholm, Sweden) and the Norwegian Trotting Association (Oslo, Norway).

Authors' contributions

BDV, KJF, and GL conceived and designed the experiments; BDV, KJF, MKR, MW, and ES contributed to sampling. GL and ES contributed the reagents and KJF and MKR extracted the DNA; BDV and ML analyzed the data and drafted the manuscript; KJF, MKR, MW, ES, MS, CFI, ML, and GL discussed and contributed to data analysis; All authors read and approved the final manuscript.

Ethics approval and consent to participate

All experimental procedures and sample collection methods were approved by the Ethics Committee for Animal Experiments in Uppsala, Sweden [Number: C 121/14]. Samples used in the study were already available at either the Animal Genetics Laboratory at SLU in Uppsala, Sweden or the Department of Basic Sciences and Aquatic Medicine at the Norwegian University of Life Sciences in Oslo, Norway as they previously had been used for parentage testing. Permission to use the samples was granted from the Swedish Trotting Association and the Norwegian Trotting Association (the owners of the samples per the rules/guidelines of the industry).

Consent for publication

Not applicable.

Competing interests

The authors have the following interest: GL is a co-inventor on a granted patent concerning commercial testing of the DMRT3 mutation: A method to predict the pattern of locomotion in horses. PCT EP 12747875.8. European patent registration date: 2011-05-05, US patent registration date: 2011-08-03. There are no further patents, products in development, or marketed products to declare.

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References

- Meadows JRS, Lindblad-Toh K. Dissecting evolution and disease using comparative vertebrate genomics. *Nat Rev Genet.* 2017;18:624–36.
- Andersson L. Domestic animals as models for biomedical research. *Ups J Med Sci.* 2016;121:1–11.
- Petersen JL, Mickelson JR, Rendahl AK, Valberg SJ, Andersson LS, Axelsson J, et al. Genome-wide analysis reveals selection for important traits in domestic horse breeds. *PLoS Genet.* 2013;9:e1003211.
- Andersson L. How selective sweeps in domestic animals provide new insight into biological mechanisms. *J Intern Med.* 2012;271:1–14.
- Andersson LS, Larhammar M, Memic F, Wootz H, Schwochow D, Rubin CJ, et al. Mutations in *DMRT3* affect locomotion in horses and spinal circuit function in mice. *Nature.* 2012;30:642–6.
- Petersen JL, Mickelson JR, Cothran EG, Andersson LS, Axelsson J, Bailey E, et al. Genetic diversity in the modern horse illustrated from genome-wide SNP data. *PLoS One.* 2013;8:e54997.
- Svensk Travsport:Uppfödning. <https://www.travsport.se/artikel/uppfoedning> (2018). Accessed 31 Aug 2018.
- Det Norske Travselvskap: Næring og Avl. <https://www.travsport.no> (2018). Accessed 31 Aug 2018.
- Föreningen Nordsvenska Hästen. <http://www.nordsvensken.org/> (2018). Accessed 31 Aug 2018.
- Jäderkvist Fegraeus K, Velie BD, Axelsson J, Ang R, Hamilton NA, Andersson L, et al. A potential regulatory region near the *EDN3* gene may control both harness racing performance and coat color variation in horses. *Physiol Rep.* 2018;6:e13700.
- Bohin O, Rönningsen K. Inbreeding and relationship within the north-Swedish horse. *Acta Agric Scand.* 1975;25:121–5.
- Árnason T. Trends and asymptotic limits for racing speed in Standardbred trotters. *Livest Prod Sci.* 2001;72:135–45.
- Árnason T, Bendroth M, Philipsson J, Henriksson K, Darenius A. Genetic evaluations of Swedish trotters-state of breeding evaluation in trotters. Proceedings of the European Federation of Animal Science symposium of the commission on horse production. Wageningen, the Netherlands: Pudoc; 1989. p. 106–29.
- Pedigree Online All Breed Database. <https://www.allbreedpedigree.com/> (2018). Accessed 31 Aug 2018.
- Ricard A, Robert C, Blouin C, Baste F, Torquet G, Morgenthaler C, Rivière J, Mach N, Mata X, Schibler L, Barrey E. Endurance exercise ability in the horses: a trait with complex polygenic determinism. *Front Genet.* 2017;8:89.
- Hermeijer G. The Vps10p-domain receptor family. *Cell Mol Life Sci.* 2009;66:2677–89.
- Reitz C, Tosto G, Vardarajan B, Rogava E, Ghani M, Rogers RS, et al. Independent and epistatic effects of variants in *VPS10-d* receptors on Alzheimer disease risk and processing of the amyloid precursor protein (APP). *Transl Psychiatry.* 2013;3:e256.
- Reitz C. The role of the retromer complex in aging-related neurodegeneration: a molecular and genomic review. *Mol Gen Genomics.* 2015;290:413–27.
- Binzer S, Stenager E, Binzer M, Kyvik KO, Hillert J, Imrell K. Genetic analysis of the isolated Faroer Islands reveals *SORCS3* as a potential multiple sclerosis risk gene. *Mult Scler.* 2016;22:733–40.
- Breiderhoff T, Christiansen GB, Pallesen LT, Vaegter C, Nykjaer A, Holm MM, et al. Sortilin-related receptor *SORCS3* is a postsynaptic modulator of synaptic depression and fear extinction. *PLoS One.* 2013;8:e75006.
- Lionel AC, Crosbie J, Barbosa N, Goodale T, Thiruvahindrapuram B, Rickaby J, et al. Rare copy number variation discovery and cross-disorder comparisons identify risk genes for ADHD. *Sci Transl Med.* 2011;3:95ra75.
- Hu Z, Park CA, Reecy JM. Developmental progress and current status of the animal QTLdb. *Nucleic Acids Res.* 2016;44:D827–33.
- Frischknecht M, Jagannathan V, Plattet P, Neuditschko M, Signer-Hasler H, Bachmann I, et al. A non-synonymous *HMG2* variant decreases height in Shetland ponies and other small horses. *PLoS One.* 2015;10:e0140749.
- Metzger J, Schrimpf R, Philipp U, Distl O. Expression levels of *LCORL* are associated with body size in horses. *PLoS One.* 2013;8:e56497.
- Avila F, Mickelson JR, Schaefer RJ, McCue ME. Genome-wide signatures of selection reveal genes associated with performance in American quarter horse subpopulations. *Front Genet.* 2018;9:249.
- Tang SL, Gao YL, Wen-Zhong H. Knockdown of *TRIM37* suppresses the proliferation, migration and invasion of glioma cells through the inactivation of *PI3K/Akt* signaling pathway. *Biomed Pharmacother.* 2018;99:59–64.
- Jobic F, Morin G, Vincent-Delorme C, Cadet E, Cabry R, Mathieu-Dramard M, et al. New intragenic rearrangements in non-Finnish mulibrey nanism. *Am J Med Genet A.* 2017;173:2782–8.
- Mosher DS, Quigon P, Bustamante CD, Sutter NB, Mellersh CS, Parker HG, Ostrander EA. A mutation in the myostatin gene increases muscle mass and enhances racing performance in heterozygote dogs. *PLoS Genet.* 2007;3:e79.
- Hill EW, McGivney BA, Gu J, Whiston R, MacHugh DE. A genome-wide SNP-association study confirms a sequence variant (g.66493737C>T) in the equine myostatin (*MSTN*) gene as the most powerful predictor of optimum racing distance for thoroughbred racehorses. *BMC Genomics.* 2010;11:552.
- McGivney BA, Browne JA, Fonseca RG, Katz LM, MacHugh DE, Whiston R, Hill EW. *MSTN* genotypes in thoroughbred horses influence skeletal muscle gene expression and race track performance. *Anim Genet.* 2012;43:810–2.
- Árnason T. Genetic evaluation of Swedish Standardbred trotters for racing performance traits and racing status. *J Anim Breed Genet.* 1999;116:387–98.
- Li H, Durbin R. Fast and accurate short read alignment with burrows-wheeler transform. *Bioinformatics.* 2009;25:1754–60.
- McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernysky A, et al. The genome analysis toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.* 2010;20:1297–303.
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, et al. Genome project data P: the sequence alignment/map format and SAMtools. *Bioinformatics.* 2009;25:2078–9.
- Li H. A statistical framework for SNP calling, mutation discovery, association mapping and population genetical parameter estimation from sequencing data. *Bioinformatics.* 2011;27:2987–93.
- Kofler R, Orozco-terWengel P, De Maio N, Pandey RV, Nolte V, Futschik A, et al. PoPoolation: a toolbox for population genetic analysis of next generation sequencing data from pooled individuals. *PLoS One.* 2011;6:e15925.

37. Karlsson EK, Baranowska I, Wade CM, Salmon Hillbertz NHC, Zody MC, Anderson N, et al. Efficient mapping of mendelian traits in dogs through genome-wide association. *Nat. Genet.* 2007;39:1321–8.
38. R Development Core Team: R-A Language and Environment for Statistical Computing. <https://www.r-project.org/>. (2018). Accessed 13 Jan 2018.
39. Thomas PD, Campbell MJ, Kejariwal A, Mi HY, Karlak B, Daverman R, et al. PANTHER: a library of protein families and subfamilies indexed by function. *Genome Res.* 2003;13:2129–41.
40. Mi H, Poudel S, Muruganujan A, Casagrande JT, Thomas PD. PANTHER version 10: expanded protein families and functions, and analysis tools. *Nucleic Acids Res.* 2016;44D336–42.
41. Neph S, Kuehn MS, Reynolds AP, Haugen E, Thurman RE, Johnson AK, et al. BEDOPS: high-performance genomic feature operations. *Bioinformatics.* 2012;28:1919–20.

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RESEARCH ARTICLE

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A genome-wide association study for harness racing success in the Norwegian-Swedish coldblooded trotter reveals genes for learning and energy metabolism

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Abstract

Background: Although harness racing is of high economic importance to the global equine industry, significant genomic resources have yet to be applied to mapping harness racing success. To identify genomic regions associated with harness racing success, the current study performs genome-wide association analyses with three racing performance traits in the Norwegian-Swedish Coldblooded Trotter using the 670 K Axiom Equine Genotyping Array.

Results: Following quality control, 613 horses and 359,635 SNPs were retained for further analysis. After strict Bonferroni correction, nine genome-wide significant SNPs were identified for career earnings. No genome-wide significant SNPs were identified for number of gallops or best km time. However, four suggestive genome-wide significant SNPs were identified for number of gallops, while 19 were identified for best km time. Multiple genes related to intelligence, energy metabolism, and immune function were identified as potential candidate genes for harness racing success.

Conclusions: Apart from the physiological requirements needed for a harness racing horse to be successful, the results of the current study also advocate learning ability and memory as important elements for harness racing success. Further exploration into the mental capacity required for a horse to achieve racing success is likely warranted.

Keywords: Genetic, Horse, Performance, Racehorse

Background

Regardless of horseracing discipline, speed, or perhaps more appropriately, unparalleled speed, is the “holy grail” of almost every horse owner, trainer, and breeder. However, speed alone does not necessarily equate to success on the racecourse. The manner in which a horse demonstrates speed is critical to its racing success [1–3]. For example, while the ability to gallop fast may result in a champion Thoroughbred (TB) or Quarter Horse (QH), the same ability in a Standardbred (SB) or Coldblooded Trotter (CT) is of little value. In SB and CT racing, horses

undoubtedly require speed, but galloping, a four-beat gait, results in disqualification [1–3]. Thus, speed in these breeds must be demonstrated at trot, a contralateral two-beat gait, or pace, an ipsilateral two-beat gait [1–4]. Consequently, racing success in SBs, CTs, and other harness racing breeds depends not on an individual's capacity for speed, but on an individual's capacity for speed in a specific gait.

To date, significant genomic resources have been applied in studies attempting to map speed and racing success in TBs and QHs [4–18]. While these studies have proven to be of great value for gallop racing breeds, their applicability to harness racing breeds has been limited [19–21]. In fact, a genomic study exploring locomotion pattern in Icelandic horses is arguably the most influential

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study to date when it comes to racing success in harness racing breeds [22]. The study discovered that a premature stop codon in the doublesex and mab-3 related transcription factor 3 (*DMRT3*) gene had a major effect on the pattern of locomotion in the horse, resulting in some horses possessing the ability to trot or pace at high speed without transitioning into gallop [22]. Follow-up studies were then able to demonstrate that for many harness racing breeds the *DMRT3* mutation was associated with racing success (e.g. increased earned prize money) [20, 23–26]. However, the mutation appears to only account for between 0 and 6.3% of the phenotypic variation in traits widely used to evaluate racing success [24, 26]. Considering that heritability estimates for some of these traits are as high as 0.38, the likelihood of other genes playing a significant role in a harness racing horse's success is high [20, 23, 27–38]. Nevertheless, genome-wide association (GWA) studies with harness racing performance are lacking [23].

Despite the fact that harness racing success is of high economic importance to the global equine industry, genome scans for performance appear to predominantly target Thoroughbreds and sport horses (e.g. Warmbloods) [4–6, 8–14, 39–41]. Most studies involving harness racing breeds tend to focus on detecting genes underlying certain conditions and diseases [42–47]. While disease studies are undoubtedly important for improved animal well-being, a greater awareness of the genes, and by extension the underlying biological mechanisms, involved in racing success are also likely to prove highly valuable. A deeper understanding of the biology underpinning success in a racehorse ultimately can lead to more targeted medical treatments as well as a heightened awareness of the physical limitations (e.g. lack of speed at trot) some horses will inevitably possess.

Motivated by these facts, we conducted a GWA study to identify genomic regions and genes associated with harness racing success using the Norwegian-Swedish Coldblooded Trotter (NSCT). Norwegian-Swedish Coldblooded Trotters are ideally suited for GWA analyses of harness racing performance as their small population is not only likely to correspond with low within breed genetic variation, but the limited region in which NSCTs are eligible to race is also likely to reduce environmental variation. Thus, a more accurate assessment of the relationship between genomic regions and harness racing performance is achievable.

Methods

Data collection

Pedigree information and performance data on all raced and unraced NSCTs born between 1 January 2000 and 31 December 2009 were provided by the trotting associations in both Norway and Sweden (Norsk Rikstoto and Svensk Travsport). Pedigree information included horse

name, horse id, date of birth, country of birth, sex, breeder, sire id, and dam id. Performance data, as of 8 February 2017, was presented per race per horse (i.e. data included individual race records with each record corresponding to a given horse's specific performance in the race). This data included non-competitive premie and qualification races, with each record containing information on horse id, race date, race track, race type, race distance, trainer, owner, driver, finish position, prize money earned, gallop status, and average km time [48, 49].

Sample acquisition

In order to reflect the raced population as accurately as possible, a list of raced individuals was randomly generated from the data described above using the statistical software R [50]. In addition to having raced, two requirements were set for inclusion in the study: 1. Hair and/or blood samples had to be readily accessible from the pedigree registration authorities in either Norway (Department of Basic Sciences and Aquatic Medicine, Norwegian University of Life Sciences) or Sweden (Animal Genetics Laboratory, Swedish University of Agricultural Sciences) 2. Sufficient sample material had to be available to ensure DNA quality standards would likely be achieved. The first 661 horses on the list that met these criteria were included in the study. Average relatedness within the selected group, estimated using the genetic software Contribution, Inbreeding, Coancestry, was 0.16 (interquartile range [IQR] 0.11–0.18) [51].

DNA isolation

Deoxyribonucleic acid was prepared from the hair roots using a standard hair-preparation procedure. Briefly, 186 μ L Chelex 100 Resin (Bio-Rad Laboratories, Hercules, CA) and 14 μ L of proteinase K (20 mg/mL; Merck KGaA, Darmstadt, Germany) were added to the sample. The mix was incubated at 56 °C for 2 h and the proteinase K was inactivated for 10 min at 95 °C. For DNA preparation from blood samples, 350 μ L of blood was used and isolated on the Qiasymphony instrument using the Qiasymphony DSP DNA mini kit (Qiagen, Hilden, Germany). A summary of the final horses selected for genotyping is shown in Table 1.

Genotyping and quality control

Prior to quality control (QC) the data set consisted of individuals genotyped using the 670 K Axiom Equine Genotyping Array ($n = 570$) and the 670 K+ Axiom Equine Genotyping Array ($n = 91$). The data from the two arrays were subsequently merged based on SNP name, chromosome, and position, yielding a combined SNP data set of 611,888 SNPs for 661 horses. Iterative QC was then performed with the GenABEL package in R to remove poorly genotyped and noisy data using the

Table 1 Summary information for the genotyped and the final sample of horses

	Genotyped Horses N	Final GWA Horses N
Sex		
Intact males	70	62
Females	271	247
Geldings	320	304
Country of birth		
Norway	360	312
Sweden	301	301
Year of birth		
2000	45	36
2001	73	66
2002	84	80
2003	69	64
2004	56	49
2005	62	55
2006	70	67
2007	71	70
2008	63	59
2009	68	67
Total	661	613

GWA Genome-wide association

following thresholds: minor allele frequency (MAF) (< 0.5%), missing genotypes per single nucleotide polymorphism (SNP) (> 5%), missing SNPs per sample (> 15%), and Hardy-Weinberg equilibrium (HWE) (first QC $p < 1e^{-10}$; second QC FDR < 0.2) [50].

Phenotyping

Pedigree and performance data for the 661 genotyped horses were structured for analyses using custom scripts written in Perl v5.20.1. Sex classifications were based on the official sex of the horse at the end of the study. Number of career starts was defined as the total number of competitive races for each individual (i.e. premie and qualification races were excluded). Number of gallops (NG) was defined as the total number of competitive races in which the horse was recorded as galloping at some point during the race. Career earnings (CE) were calculated as the total amount of prize money won for each horse as of 8 February 2017. Prize money won in Sweden (SEK) was converted to Norwegian currency (NOK) based on the average exchange rate for the year in which the race occurred [52]. Horses having participated in only premie or qualification races were given a value for career earnings of - 1 NOK in order to distinguish them from horses that had zero career earnings, despite having raced competitively. Best km time (BT),

independent of starting method, was defined as the fastest average km time for a competitive race in which the horse was not recorded as galloping during the race (i.e. races in which a horse galloped, regardless of if the horse was disqualified, were excluded).

Genome-wide association analyses

Genome-wide association (GWA) analyses were performed using the GenABEL package in R [50]. The package was used to compute an autosomal genomic kinship matrix as well as perform standard K-means clustering. K-means clustering with $K = \{1,2,\dots,10\}$ were executed to determine the number of clusters (subpopulations). For each iteration, the sum of within-cluster sum of squares ($\Sigma WCSS$) was calculated and subsequently plotted vs. K. To define the subpopulations, the number of clusters corresponding with the first inflection point ($K = 3$) was chosen [53]. The multidimensional scaling (MDS) plot yielded no apparent outliers and a visualization of the genomic-kinship matrix and subpopulations using MDS can be seen in Fig. 1.

To account for any population stratification, genome-wide association analyses of CE, BT, and NG were performed using a mixed model-structured association approach (“mmscore” function with the “strata” option in GenABEL). Preliminary analyses indicated significant effects of sex, birth year, and number of career starts on all three traits. Country of birth was also shown to influence CE. As a result, sex, birth year, number of career starts, and country of birth were included as co-variants in the final analyses accordingly. Genome-wide significance for each of the analyses was determined by Bonferroni correction ($p < 1.39 \times 10^{-7}$, corrected for total number of SNPs post QC) while a “suggestive” genome-wide significance threshold was also set at 1.0×10^{-5} . Manhattan and quantile-quantile plots were generated in R while the extent of linkage disequilibrium (LD) was estimated by calculating r^2 values between all pairs of SNPs with inter-SNP distances of less than 1 Mb using PLINK v1.09 (<http://zzz.bwh.harvard.edu/plink/>). The effB in the GenABEL result was regarded as the allele substitution effect and the proportion of phenotypic variance explained by each significant SNP was estimated as follows:

$$VAR(\%) = \frac{2pq\beta^2}{S^2} * 100$$

Where p and q are the allele frequencies, β is the estimated allele substitution effect, and S^2 is the sample phenotypic variance. Stepwise regressions were then performed to estimate the total proportion of phenotypic variation explained by the multiple genome-wide associated SNPs (before and after pruning at $r^2 > 0.2$; PLINK

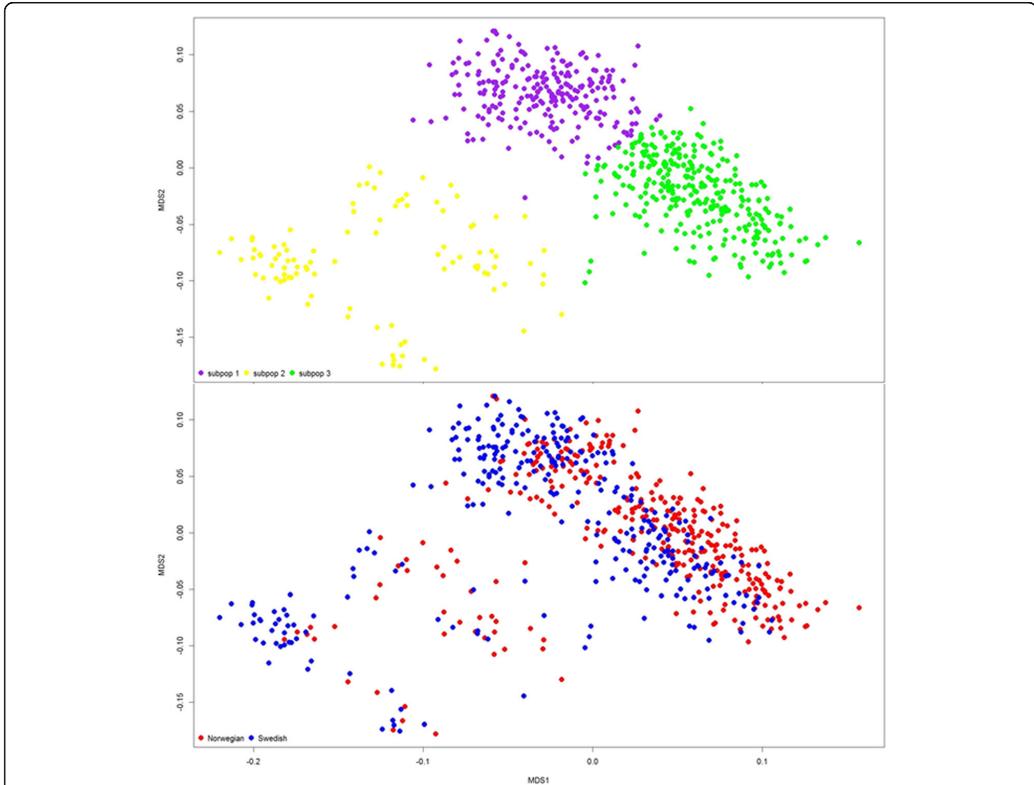


Fig. 1 Visualization of multidimensional scaling, stratified by country of birth and subpopulation, performed on the genomic-kinship matrix

command `-indep-pairwise 100 25 0.2`) for each trait using the `lm` function in R [50]. The bioinformatics database Ensembl (<http://www.ensembl.org/>) was used for candidate gene screening. Genomic coordinates of genome-wide significant and suggestive genome-wide significant SNPs ± 500 kb were used as inputs to generate a list of annotated genes using the Ensembl BiomaRt function. The PANTHER Classification system was then used to obtain an overview of the biological processes, molecular functions, and pathways known to be affected by these genes [54, 55].

Results

Following QC, 359,635 autosomal SNPs and 642 horses were available for association analyses. Of these individuals, 29 had only participated in non-competitive premie and/or qualification races. Initial association analyses with CE, combined with the vast array of reasons known to prevent a horse from competitive racing suggested the exclusion of these horses would help to reduce noise in the final analyses. As a result, only 613 horses,

representing 120 sires (interquartile range [IQR] 1–5) and 547 dams (IQR 1–1), were included in the final GWA analyses (Table 1). Descriptive statistics for CE, BT, and NG in the final sample are presented in Table 2. Both CE and NG were not normally distributed and were subsequently log transformed for the GWA analyses. The extent of LD decayed faster across the final sample of horses with mean r^2 dropping below 0.20 by 3 kb (Additional file 1). The GWA analysis of CE yielded multiple genome-wide significant SNPs ($p < 1.39 \times 10^{-7}$), with the majority of these SNPs residing on *Equus caballus* chromosome (ECA) 6 (Fig. 2; Table 3; Additional file 2). Analyses of both BT and NG failed to result in genome-wide significant SNPs. However, two regions of interest were apparent based on the presence of slight peaks on ECA17 and ECA23 in the resulting Manhattan plots (Fig. 3; Additional file 2). Genome-wide significant SNPs for CE, suggestive genome-wide significant SNPs for BT and NG, and the nearest genes are shown in Table 3. The most significant SNP was detected at the 20,006,740 position on chromosome 28 (AX-104828170, $p = 9.01E-10$)

Table 2 Descriptive statistics of CE, BT, NG for the final sample of horses (*n* = 613)

	Min	25th percentile	Median	Mean	75th percentile	Max
Career earnings (NOK)	0	36,321	124,625	302,506	327,493	4,216,554
Best km time (s)	78.9	86.8	89.3	89.6	92.4	105.7
Number of gallops	0	5	11	14.4	21	123

CE career earnings, BT Best km time, NG number of gallops

and presented an estimated allele substitution effect of -7079.46 NOK. The favourable allele appeared to be the T allele, with each C allele resulting in a negative effect on CE. Despite the fact that the frequency of the T allele was 98.2% and the frequency of the C allele was only 1.8%, the percentage of phenotypic variance explained by this SNP was 3.85%. The 32 SNPs in total were estimated to explain 18.34%, 18.71% and 33.17% of the variation for CE, NG and BT, respectively, in the population studied. Consequently, after LD pruning, only 17 SNPs remained - explaining 14.17%, 18.38% and 33.13% of the variation for CE, NG and BT, respectively (Table 3). Overall, 378, 144, and 23 candidate genes identified were associated with known biological processes, molecular functions, and pathways, respectively (Figs. 4, 5 and 6; Additional file 3).

Discussion

Knowing where, why, and how genes and athletic prowess intersect in a racehorse has long been the goal of countless researchers, veterinarians, breeders, trainers, and owners [4-38]. While great strides in this area have recently been made for gallop racing horses, similar

advancements for harness racing horses have been limited [4-26]. Using the NSCT, the current study explored the genetic background for athletic prowess in a harness racing horse by performing GWA analyses and functional classification for three traits associated with harness racing success. These analyses resulted in a total of 32 SNPs of interest with 9 demonstrating genome-wide significance and 13 residing in genes. Subsequent functional classifications went on to provide further support of the complexity of harness racing success with several candidate genes involved in neurological, metabolic, and musculoskeletal regulation identified. Since a gene can be declared as a candidate gene if at least 1 out of the 4 following characteristics are present: 1) the gene has a known physiological role in the phenotype of interest, 2) the gene affects the trait in question based on studies of knockouts, mutations, or transgenics in other species, 3) The gene is preferentially expressed in organs related to the quantitative trait, or 4) the gene is preferentially expressed during developmental stages related to the phenotype, a large fraction of the genes identified in the current study can plausibly be considered as candidate

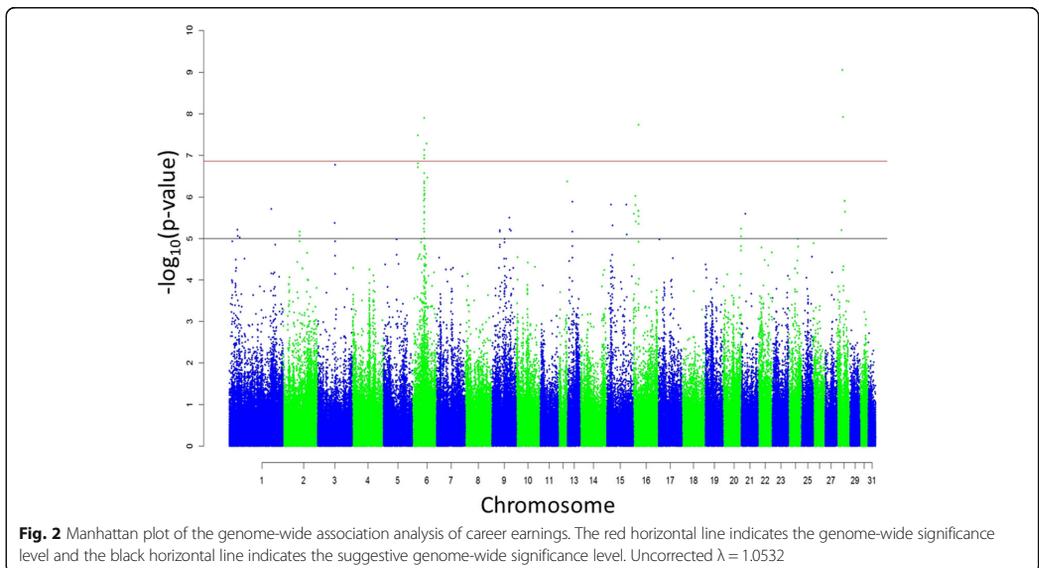


Fig. 2 Manhattan plot of the genome-wide association analysis of career earnings. The red horizontal line indicates the genome-wide significance level and the black horizontal line indicates the suggestive genome-wide significance level. Uncorrected $\lambda = 1.0532$

Table 3 Genome-wide significant SNPs for CE and suggestive genome-wide significant SNPs for BT and NG

SNP	Analysis	Location (EC:A:bp)	MAF	β	SE	Var (%)	Nearest gene	Distance to nearest gene (bp)	Raw p-value
AX-104828170	CE	28:20,006,740	0.018	-3.380	0.552	3.85	<i>NDUFA12</i>	114,732	9.01E-10
AX-103147507	CE	28:22,153,465	0.005	-5.762	1.011	3.27	<i>ENSECAG00000025907</i>	46,196	1.20E-08
AX-104611735	CE	6:47,132,529	0.015	-3.407	0.599	3.16	<i>PDE3A</i>	0	1.28E-08
AX-104865129	CE	16:18,366,321	0.008	-4.488	0.798	3.17	<i>ENSECAG00000003087/PROK2</i>	138,149/138,791	1.87E-08
AX-104494389	CE	6:20,020,914	0.006	-4.557	0.825	3.32	<i>INPP5D</i>	0	3.36E-08
AX-103248294	CE	6:49,512,490	0.016	-3.166	0.582	3.11	<i>SOX5</i>	0	5.28E-08
AX-103090138	CE	6:42,063,985	0.018	-3.007	0.559	2.46	<i>PLBD1</i>	3191	7.48E-08
AX-104711589	CE	6:41,462,481	0.012	-3.497	0.657	2.59	<i>GRIN2B</i>	0	9.99E-08
AX-104307051	CE	6:41,329,519	0.012	-3.474	0.657	2.67	<i>GRIN2B</i>	0	1.21E-07
AX-104144838	NG	23:23,333,501	0.278	-0.214	0.046	1.51	<i>ENSECAG000000023609</i>	187,643	3.18E-06
AX-104568609	NG	1:159,285,045	0.004	-1.343	0.293	1.14	<i>ENSECAG00000003696/ENSECAG000000022264</i>	19,813/25,804	4.52E-06
AX-102982528	NG	23:23,324,996	0.412	-0.186	0.041	1.27	<i>ENSECAG000000023609</i>	179,138	6.74E-06
AX-103734745	NG	29:24,530,437	0.453	-0.176	0.040	1.29	<i>ENSECAG000000004576</i>	944,126	8.67E-06
AX-104373992	BT	1:162,993,722	0.016	3.289	0.655	1.79	<i>ENSECAG000000023062/ENSECAG000000008721</i>	32,728/51,254	5.10E-07
AX-103261370	BT	23:22,522,071	0.409	-0.910	0.183	0.87	<i>DOCK8</i>	0	6.49E-07
AX-104219924	BT	17:19,525,955	0.279	0.946	0.197	2.24	<i>WDFY2</i>	0	1.57E-06
AX-104634248	BT	23:21,857,316	0.108	1.346	0.284	1.21	<i>PIPSK1B/FAM122A</i>	14,893/18,794	2.20E-06
AX-103287280	BT	23:21,064,571	0.117	1.270	0.268	1.49	<i>PTAR1</i>	43,300	2.22E-06
AX-104645782	BT	17:19,318,167	0.258	0.951	0.201	2.21	<i>ATP7B</i>	0	2.32E-06
AX-103762427	BT	23:22,461,979	0.154	-1.156	0.245	1.37	<i>DOCK8</i>	0	2.45E-06
AX-103530176	BT	23:22,464,604	0.156	-1.139	0.246	1.25	<i>DOCK8</i>	0	3.49E-06
AX-103445942	BT	1:151,919,692	0.012	3.600	0.777	1.39	<i>ENSECAG000000012236/ENSECAG000000013533</i>	74,605/78,356	3.63E-06
AX-104538418	BT	17:21,083,126	0.279	0.905	0.196	2.29	<i>KCNRG</i>	0	3.99E-06
AX-104268231	BT	23:21,689,609	0.385	-0.823	0.179	0.79	<i>PIPSK1B</i>	0	4.25E-06
AX-102964033	BT	23:22,423,197	0.121	-1.259	0.275	0.95	<i>DOCK8</i>	5699	4.59E-06
AX-104117851	BT	7:65,266,179	0.010	3.724	0.821	1.76	<i>ENSECAG000000007398</i>	162,661	5.78E-06
AX-104642194	BT	31:14,300,483	0.006	4.612	1.021	1.37	<i>MTRF1L/FBXO5</i>	108/14,491	6.31E-06
AX-103166989	BT	23:22,496,787	0.116	-1.270	0.283	0.99	<i>DOCK8</i>	0	7.07E-06
AX-103803214	BT	2:21,466,714	0.049	1.725	0.386	1.82	<i>AGO1</i>	2494	7.72E-06
AX-104450418	BT	2:21,311,680	0.087	1.351	0.304	1.93	<i>TEKT2/ADPRHL2</i>	10,587/14,440	9.00E-06
AX-103305676	BT	25:26,866,219	0.019	2.590	0.584	1.67	<i>OR1L3</i>	492	9.11E-06
AX-104591507	BT	17:20,813,164	0.249	0.898	0.203	1.84	<i>KCNRG</i>	227,34	9.36E-06

CE career earnings, BT Best km time, NG number of gallop, MAF Minor allele frequency, β Estimated allele substitution effect, Var (%) Percentage of phenotypic variance explained

Red line = Bonferroni threshold ($P < 1.39 \times 10^{-7}$)

genes [56]. As a result, the following discussion prioritizes genes that contained variants with significant or suggestive associations with our traits of interest. The rationale for this prioritization is simply that for associated variants that reside outside of annotated genes, it is in general more difficult to determine which gene(s) the variants act on.

Glutamate ionotropic receptor NMDA type subunit 2B (GRIN2B)

Two genome-wide significant SNPs associated with career earnings were located in the *GRIN2B* gene, a gene also identified in a previous study exploring pacing ability in Icelandic horses [57]. The gene has been shown to be involved in neural regulations in humans and

laboratory species with mutations in the gene having been associated with neurodevelopmental disorders [58–60]. Considered to be an important factor for learning and memory, one can only speculate as to its association with career earnings in a harness racing horse. However, horses with a greater capacity to learn and adapt to the highly variable nuances of harness racing would conceivably be more likely to achieve racing success. On the other hand, the gene’s association with attention-deficit/hyperactivity disorder in humans suggests that perhaps certain horses lack the ability to focus on racing and training, thereby preventing or at least hindering their racing performance [61].

ATPase copper transporting beta (*ATP7B*)

A single suggestive genome-wide significant SNP associated with best time was located in the *ATP7B* gene on ECA17. The gene encodes a protein that functions as a monomer, exporting copper out of cells. Excess copper

can cause serious toxicity with the process of excess copper disposal relying heavily on *ATP7B* [62, 63]. Over 500 mutations have been identified in the gene, 380 of which are considered to be disease causing mutations [64]. Elevated levels of copper in the body often result in muscle stiffness with acute muscle stiffness prior to a race having the potential to affect individual performance and chronic muscle stiffness likely to impact a horse’s conditioning, trainability, and overall capacity for speed [62, 63].

Potassium channel regulator (*KCNRG*)

The *KCNRG* gene on ECA17 was also identified by two SNPs demonstrating suggestive genome-wide significance for best time. The gene encodes a protein which regulates the activity of voltage-gated potassium channels with a study using songbirds suggesting potassium channels to be lineage specific [64, 65]. The same study also revealed that apart from broad expression in the

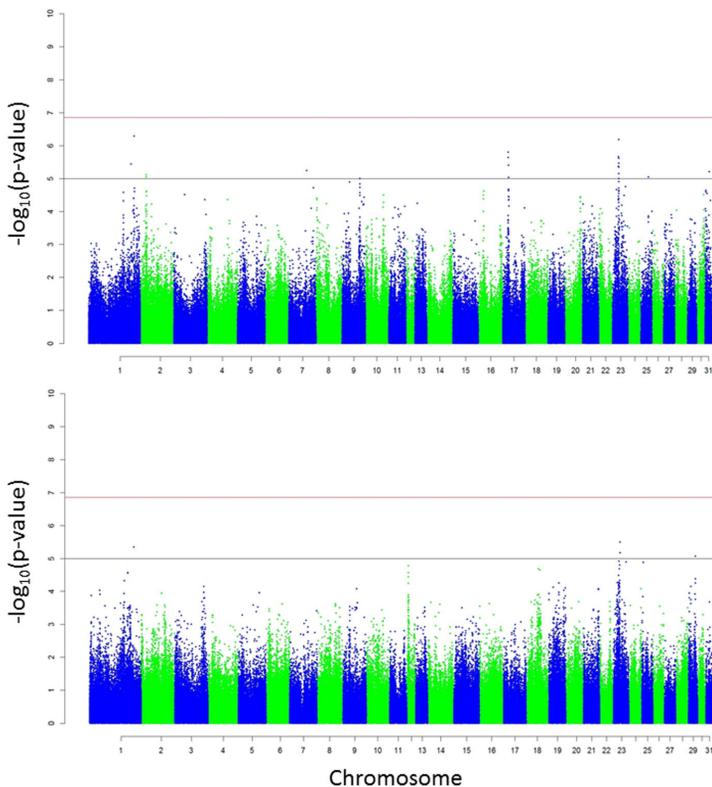


Fig. 3 Manhattan plots of the genome-wide association analyses of best km time and number of gallops. The red horizontal lines indicate the genome-wide significance levels and the black horizontal lines indicate the suggestive genome-wide significance levels. Top panel: Best km time analysis, uncorrected $\lambda = 1.0902$. Bottom panel: Number of gallops analysis, uncorrected $\lambda = 1.0256$

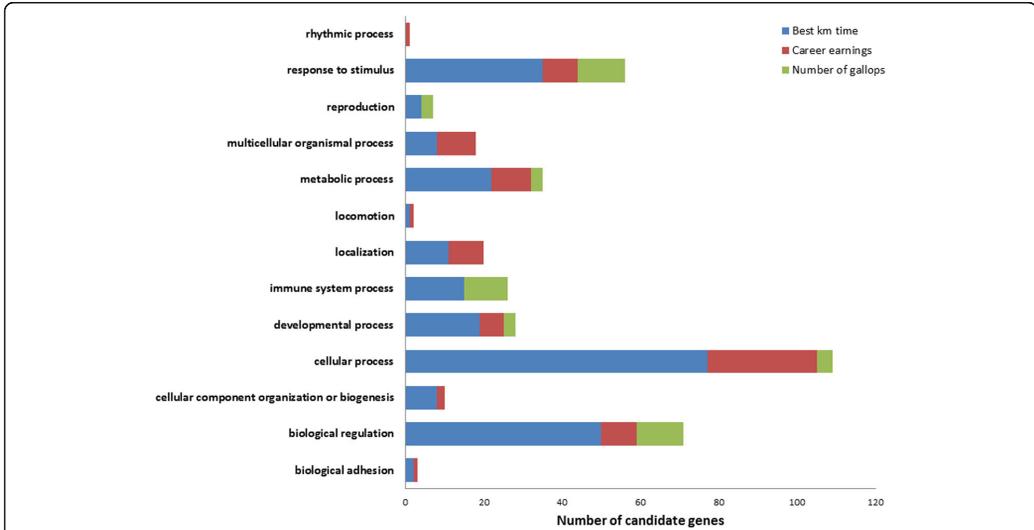


Fig. 4 Biological process summary information from the functional classification analysis of candidate genes in PANTHER. PANTHER biological process classification: the function of the protein in the context of a larger network of proteins that interact to accomplish a process at the level of the cell or organism

brain a subset of potassium channel genes are selectively expressed, with the authors hypothesizing that the *KCNRG* gene may be associated with learning [65]. Although no previous studies of racehorse performance have specifically identified *KCNRG* as important for racing success, a large

conserved haplotype on ECA17 has been advocated to have selective importance in Thoroughbreds and closely related breeds [4, 66].

Also of note is the role voltage-gate potassium channels play in Hyperkalemic periodic paralysis (HYPP), a

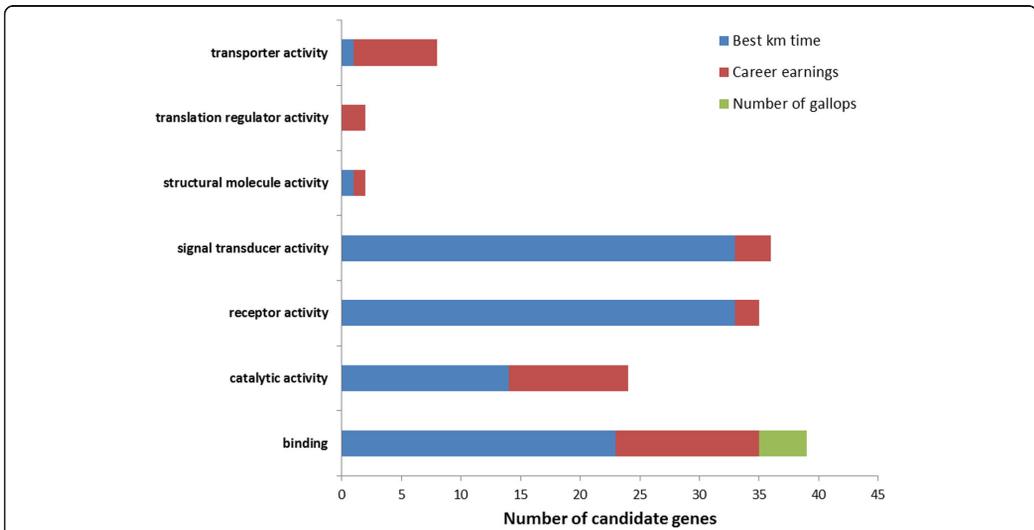
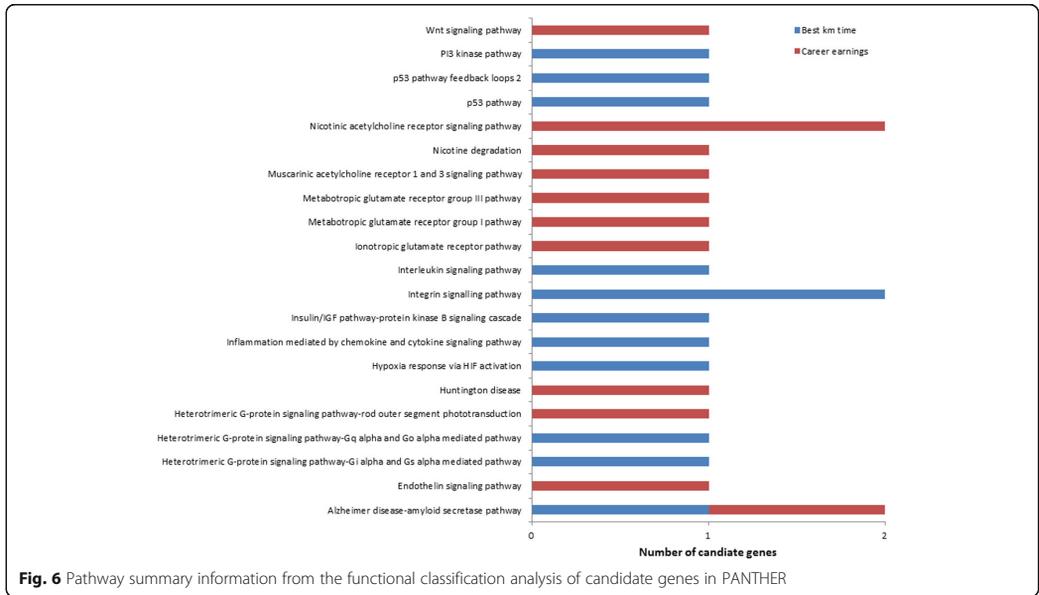


Fig. 5 Molecular function summary information from the functional classification analysis of candidate genes in PANTHER. PANTHER molecular function classification: the function of the protein by itself or with directly interacting proteins at a biochemical level



genetic disorder predominantly seen in Quarter Horses. The condition, caused by a mutation in the sodium voltage-gated channel alpha subunit 4 (*SCN4A*) gene, manifests intermittently with clinical signs ranging from muscle fasciculation to signs of paresis [67–70]. Hyperkalemia, a term used to describe abnormally high levels of potassium in the blood, is often seen during or immediately after an attack. Voltage-gated potassium channels are thought to remain open, allowing continual potassium efflux, thereby promoting an open sodium channel configuration. As a result, an HYPP attack can be triggered or an already occurring attack can increase in severity [67, 70–72]. Horses with HYPP also tend to possess hypertrophic muscles; however, they have been reported as having a reduced tolerance to exercise with relatively more lactate being produced during exercise [73, 74].

Phosphatidylinositol-4-phosphate 5-kinase type 1 beta (*PIP5K1B*)

A single suggestive genome-wide significant SNP associated with best time was also located in the *PIP5K1B* gene on ECA23. Three widely expressed isoforms of *PIP5K1* are responsible for the regulation of the major pools of cellular phosphatidylinositols in mammalian tissues, with *PIP5K1B* negatively regulated in response to oxidative stress [75, 76]. Neurite outgrowth, a critical process for neuronal development, has also been shown as negatively regulated by *PIP5K1A* [77]. Since the

current study is the first to suggest an association between *PIP5K1B* and racing success, understanding the roles *PIP5K1* isoforms have on a horse’s capacity for speed remains a task for future studies. However, genes that influence cell differentiation processes, such as endocytosis, assuredly contribute in some way or another to the physical limitations and overall performance of any racehorse.

Dedicator of cytokinesis 8 (*DOCK8*)

Perhaps the most obvious candidate gene for harness racing success in the current study was *DOCK8*. Five suggestive genome-wide significant SNPs indicated the importance of *DOCK8* to a horse’s best time, with 4 of the SNPs located in the gene. Mutations in *DOCK8* result in a form of hyper-IgE syndrome; however, loss or mutations of *DOCK8* have also been associated with intelligence and motor retardation [78–83]. While the importance of intelligence in a racehorse has been briefly discussed above, in the case of *DOCK8* the significance of the gene may lie with its link to motor skills. *DOCK8* is not only located on ECA23, the same chromosome as *DMRT3*, but multiple studies have hypothesized some sort of commonality or overlap between *DMRT*-(1,2,3) gene effects and *DOCK8* [22, 82, 84]. Despite the established association between *DMRT3* and harness racing performance, additional research of *DMRT3* in horses strongly suggest that the mutation is unlikely to be the single cause of gaiting ability [22, 53, 85–88]. Therefore,

it is conceivable that *DOCK8* also significantly contributes to gaiting ability, ultimately playing some role in a harness racing horse's propensity to exhibit speed at trot or pace.

Phosphodiesterase 3A (*PDE3A*)

The protein encoded by the *PDE3A* gene, a gene on ECA6 in which a single genome-wide significant SNP associated with career earnings is located, plays a critical role in cardiovascular function [89–91]. The encoded protein regulates vascular smooth muscle contraction and relaxation and has been linked to familial hypertension, cardiovascular disease, and fertility [89–92]. Healthy cardiovascular function is important for racing success as the act of racing undeniably requires a higher than resting-level of oxygen to support the horse's increased muscle activity. A mutation in the *PDE3A* gene that ultimately alters cardiovascular function could potentially prevent a horse from meeting the higher metabolic demands of racing, thus decreasing his/her chances of winning and limiting his/her career earnings. On the contrary, an advantageous mutation in the gene could allow some horses to perform at an even greater cardiovascular level, increasing their likelihood of winning races and earning more prize money.

Inositol polyphosphate-5-phosphatase D (*INPP5D*) & SRY-box 5 (*SOX5*)

Also identified by single genome-wide significant SNPs on ECA6 were the *INPP5D* and the *SOX5* genes. The *INPP5D* gene is an important regulator of immune cell signaling, while the *SOX5* gene is involved in embryonic development and has been associated with multiple human diseases and disorders [93–99]. Moreover, both genes have been suggested as important in B cell activity indicating that their association with career earnings in the current study may be rooted in the immune response of a horse [95, 100]. However, mutations in *SOX5* have also been theorized to disrupt neuronal development and function [101, 102].

Other candidate genes

Regions on ECA1, ECA7, and ECA16 have also previously been described as important for endurance performance traits, while regions on ECA14 and ECA18 associated with gallop racing in other studies do not appear to play a significant role in harness racing [4–8, 10–15, 19–21, 39, 66]. This likely suggest a greater demand for endurance in harness racing compared to gallop racing and is perhaps a sign of the different physiological demands for speed in trot versus speed in gallop. Candidate genes for harness racing success in the current study were also identified on ECA1, ECA2, ECA6, ECA7, ECA16, ECA17,

ECA23, ECA25, ECA28, ECA29, and ECA31. However, it is important to note that the MAF threshold applied in the current study is slightly lower than is generally accepted. Although this may have inadvertently resulted in some SNP associations being simply by chance, it is also plausible that the lower MAF threshold allowed for the capture of candidate genes/regions that are perhaps the difference between an elite horse and a very, very good horse. Racing performance is undoubtedly complex and the unique history of the NSCT, being a blend of draught horse and racehorse, means that rare variants cannot be ruled out purely because they are rare – particularly when one considers the rarity of an elite racehorse. While not all candidate variants/genes are discussed above, the results of the current analyses clearly suggest that different molecular and cellular events mediate adaptive processes in the neuromusculoskeletal system in response to exercise. High intensity exercise (e.g. racing) is known to be associated with significant physiological adaptations in the neuromuscular system in equine athletes with prolonged and intense exercise potentially resulting in oxidative damage to cellular constituents [103]. Moreover, the importance of the central nervous system (CNS) as a critical “central governing” factor in sporting performance has been previously documented in endurance horses with exercise shown to induce several biological processes that regulate neurological functions that help to maintain good mental health [104]. Our results add to this line of thought, providing further evidence that genes involved in neural regulations (e.g. *GRIN2B*) likely play an important role in controlling the fundamental biological processes underlying adaptation to equine athletic performance.

Conclusions

After strict Bonferroni correction, 9 genome-wide significant and 23 suggestive genome-wide significant SNPs associated with harness racing success were identified. These SNPs were located on ECA1, ECA2, ECA6, ECA7, ECA16, ECA17, ECA23, ECA25, ECA28, ECA29, and ECA31 with eight genes (*GRIN2B*, *DOCK8*, *ATP7B*, *KCNRG*, *PIPSK1B*, *PDE3A*, *INPP5D*, *SOX5*) suggested as strong candidate genes for harness racing success. Apart from the physical attributes required to achieve racing success, multiple candidate genes identified in the current study also advocate learning ability and memory as critical to success. However, further analyses of these genes based on additional genetic and functional studies are required to explore this notion in greater detail. Moreover, future studies should also consider a validation study with an independent population as well as sequencing of candidate genes to better identify causal alleles.

Additional files

Additional file 1: Extent of linkage disequilibrium of the final sample of horses. Pairwise r^2 was calculated between each SNP within 1 Mb. (TIFF 15316 kb)

Additional file 2: QQ plots for earnings, best km time, and number of gallops analyses. Top panel - corrected QQ plot for earnings analysis (Uncorrected $\lambda = 1.0532$); Middle panel - corrected QQ plot for best km time analysis ($\lambda = 1.0902$); Bottom panel - corrected QQ plot for number of gallops analysis ($\lambda = 1.0256$). (TIF 111 kb)

Additional file 3: Functional classification gene list from PANTHER analysis. (XLSX 27 kb)

Abbreviations

ATP7B: ATPase copper transporting beta; BT: Best km time; CE: Career earnings; CT: Coldblooded trotter; DMRT3: Doublesex and mab-3 related transcription factor 3; DOCK8: Dedicator of cytokinesis; ECA: *Equus caballus* chromosome; GRIN2B: Glutamate ionotropic receptor NMDA type subunit 2B; GWA: Genome-wide association; INPP5D: Inositol polyphosphate-5-phosphatase D; KCNRG: Potassium channel regulator; NG: Number of gallops; NSCT: Norwegian-Swedish Coldblooded Trotter; PDE3A: Phosphodiesterase 3A; PIP5K1B: Phosphatidylinositol-4-phosphate 5-kinase type 1 beta; QH: Quarter horse; SB: Standardbred trotter; SOX5: SRY-box 5; TB: Thoroughbred

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Availability of data and materials

The data that support the findings of this study are available from the Swedish Trotting Association (Stockholm, Sweden) and the Norwegian Trotting Association (Oslo, Norway), but restrictions apply to the availability of these data, which were used under license for the current study, and so are not publicly available. However, data are available from the authors upon reasonable request and with permission of the Swedish Trotting Association (Stockholm, Sweden) and the Norwegian Trotting Association (Oslo, Norway).

Authors' contributions

BDV, KJF, CI, ES, and GL conceived and designed the experiments; KJF, MKR, and KHR contributed to sampling. GL and ES contributed the reagents and MKR extracted the DNA; BDV and MS analyzed the data and drafted the manuscript; KJF, MS, CI, ES, and GL discussed and contributed to data analysis; All authors read and approved the final manuscript.

Ethics approval and consent to participate

All experimental procedures and sample collection methods were approved by the Ethics Committee for Animal Experiments in Uppsala, Sweden [Number: C 121/14]. Samples used in the study were already available at either the Animal Genetics Laboratory at SLU in Uppsala, Sweden or the Department of Basic Sciences and Aquatic Medicine at the Norwegian University of Life Sciences in Oslo, Norway as they previously had been used for parentage testing. Permission to use the samples was granted from the Swedish Trotting Association and the Norwegian Trotting Association (the owners of the samples per the rules/guidelines of the industry).

Consent for publication

Not applicable.

Competing interests

The authors have the following interest: GL is a co-inventor on a granted patent concerning commercial testing of the DMRT3 mutation: A method to predict the pattern of locomotion in horses. PCT EP 1274875.8. European patent registration date: 2011-05-05, US patent registration date: 2011-08-03. There are no further patents, products in development, or marketed products to declare.

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References

1. United States Trotting Association. Charter, Bylaws, Rules and regulations. In: Rule book. United States trotting association. 2018. <http://www.ustrotting.com/pdf/USTARuleBook.pdf>. Accessed 10 Jan 2018.
2. Harness Racing Australia. Australian harness racing rules. Harness racing Australia Inc. 2017. <http://www.harness.org.au/rules/AHRRules.pdf>. Accessed 10 Jan 2018.
3. Union Européenne du Trot. International Agreement On Trotting Races. Union Européenne du Trot. 2018. http://www.uet-trot.eu/images/pdf-uet/en/publications/international_agreement_on_trotting_races.pdf. Accessed 10 Jan 2018.
4. Petersen JL, Mickelson JR, Rendahl AK, Valberg SJ, Andersson LS, Axelsson J, et al. Genome-wide analysis reveals selection for important traits in domestic horse breeds. *PLoS Genet*. 2013;9:e1003211.
5. Hill EW, McGivney BA, Gu J, Whiston R, MacHugh DE. A genome-wide SNP-association study confirms a sequence variant (g.66493737C>T) in the equine myostatin (*MSTN*) gene as the most powerful predictor of optimum racing distance for thoroughbred racehorses. *BMC Genomics*. 2010;11:552.
6. Petersen JL, Valberg SJ, Mickelson JR, McCue ME. Haplotype diversity in the equine myostatin gene with focus on variants associated with race distance propensity and muscle fiber type proportions. *Anim Genet*. 2014;45:827–35.
7. Tozaki T, Hill EW, Hirota K, Kakoi H, Gamahara H, Miyake T, Sugita S, Hasegawa T, Ishida N, Nakano Y, Kurosawa M. A cohort study of racing performance in Japanese thoroughbred racehorses using genome information on ECA18. *Anim Genet*. 2011;43:42–52.
8. Shin DH, Lee JW, Park JE, Choi IY, Oh HS, Kim HJ, Kim H. Multiple genes related to muscle identified through a joint analysis of a two-stage genome-wide association study for racing performance of 1,156 thoroughbreds. *Asian Australas J Anim Sci*. 2015;28:771–81.
9. Moon S, Lee JW, Shin D, Shin KY, Kim J, Choi IY, Kim J, Kim H. A genome-wide scan for selective sweeps in racing horses. *Asian Australas J Anim Sci*. 2015;28:1525–31.
10. Tozaki T, Miyake T, Kakoi H, Gawahara H, Sugita S, Hasegawa T, Ishida N, Hirota K, Nakano Y. A genome-wide association study for racing performances in thoroughbreds clarifies a candidate region near the *MSTN* gene. *Anim Genet*. 2010;41:28–35.
11. Hill EW, Gu J, Eivers SS, Fonseca RG, McGivney BA, Govindarajan P, Orr N, Katz LM, MacHugh DE. A sequence polymorphism in *MSTN* predicts sprinting ability and racing stamina in thoroughbred horses. *PLoS One*. 2010;5:e8645.
12. Binns MM, Boehler DA, Lambert DH. Identification of the myostatin locus (*MSTN*) as having a major effect on optimum racing distance in the thoroughbred horse in the USA. *Anim Genet*. 2010;41:154–8.
13. Hill EW, Fonseca RG, McGivney BA, Gu J, MacHugh DE, Katz LM. *MSTN* genotype (g.66493737C/T) association with speed indices in thoroughbred racehorses. *J Appl Physiol*. 2012;286–90.
14. Tozaki T, Sato F, Hill EW, Miyake T, Endo Y, Kakoi H, Gawahara H, Hirota K, Nakano Y, Nambu Y, Kurosawa M. Sequence variants at the myostatin gene

- locus influence the body composition of thoroughbred horses. *J Vet Med Sci.* 2011;73:1617–24.
15. McGivney BA, Browne JA, Fonseca RG, Katz LM, MacHugh DE, Whiston R, Hill EW. MSTN genotypes in thoroughbred horses influence skeletal muscle gene expression and racetrack performance. *Anim Genet.* 2012;43:810–2.
 16. Gu J, MacHugh DE, McGivney BA, Park SDE, Katz LM, Hill EW. Association of sequence variants in CKM (creatine kinase, muscle) and COX412 (cytochrome c oxidase, subunit 4, isoform 2) genes with racing performance in thoroughbred horses. *Equine Vet J.* 2010;42:569–75.
 17. Hill EW, Gu J, McGivney BA, MacHugh DE. Targets of selection in the thoroughbred genome contain exercise-relevant gene SNPs associated with elite racecourse performance. *Anim Genet.* 2010;41:56–63.
 18. Meira CT, Fortes MRS, Farah MM, Porto-Neto LR, Kelly M, Moore SS, Pereira GL, Chardulo LAL, Curi RA. Speed index in the racing quarter horse: a genome-wide association study. *J Equine Vet Sci.* 2014;34:1263–8.
 19. Dall'Olio S, Fontanesi L, Nanni Costa L, Tassinari M, Minieri L, Falaschini A. Analysis of horse myostatin gene and identification of single nucleotide polymorphisms in breeds of different morphological types. *J Biomed Biotechnol.* 2010;542945.
 20. Negro Rama S, Valera M, Membrillo A, Gómez MD, Solé M, Menendez-Buxadera A, Anaya G, Molina A. Quantitative analysis of short- and long-distance racing performance in young and adult horses and association analysis with functional candidate genes in Spanish trotter horses. *J Anim Breed Genet.* 2016;133:347–56.
 21. Vellie BD, Bas Conn L, Petäjästö K, Røed KH, Ihler CF, Strand E, Fegraeus KJ, Lindgren G. The importance of MSTN for harness racing performance in the Norwegian-Swedish coldblooded trotter and the Finnhorse. *Proc 10th World Congr Genet Appl Livest Prod.* 2018;11:36.
 22. Andersson LS, Larhammar M, Memic F, Wootz H, Schwchow D, Rubin CJ, et al. Mutations in *DMRT3* affect locomotion in horses and spinal circuit function in mice. *Nature.* 2012;30:642–6.
 23. Ricard A. Does heterozygosity at the *DMRT3* gene make French trotters better racers? *Genet Sel Evol.* 2015;47:10.
 24. Jäderkvist K, Andersson LS, Johansson AM, Arnason T, Mikko S, Eriksson S, Anderson L, Lindgren L. The *DMRT3* "gait keeper" mutation affects performance of Nordic and Standardbred trotters. *J Anim Sci.* 2014;92:4279–86.
 25. Jäderkvist Fegraeus K, Lawrence C, Petäjästö K, Johansson MK, Wiklund M, Olsson C, Andersson L, Andersson LS, Røed KH, Ihler CF, Strand E, Lindgren G, Vellie BD. Lack of significant associations with early career performance suggest no link between the *DMRT3* "gait keeper" mutation and precocity in coldblooded trotters. *PLoS One.* 2017;12:e01777351.
 26. Jäderkvist Fegraeus K, Johansson L, Mäenpää M, Mykkänen A, Andersson LS, Vellie BD, Andersson L, Arnason T, Lindgren G. Different *DMRT3* genotypes are best adapted for harness racing and riding in Finnhorses. *J Hered.* 2015;106:734–40.
 27. Solé M, Valera M, Gómez MD, Sölkner J, Molina A, Mészáros G. Heritability and factors associated with number of harness race starts in the Spanish trotter horse population. *Equine Vet J.* 2017;49:288–93.
 28. Thiruvankadan AK, Kandasamy N, Panneseelvam S. Inheritance of racing performance of trotter horses: an overview. *Livest Sci.* 2009;124:163–81.
 29. Saastamoinen MT, Ojala MJ. Estimates of genetic and phenotypic parameters for racing performance in young trotters. *Acta Agr Scand.* 1991;41:427–36.
 30. Saastamoinen MT, Nylander A. Genetic and phenotypic parameters for age at starting to race and racing performance during early career in trotters. *Livest Prod Sci.* 1996;45:63–8.
 31. Suontama M, van der Werf JH, Juga J, Ojala M. Genetic parameters for racing records in trotters using linear and generalized linear models. *J Anim Sci.* 2012;90:2921–30.
 32. Olsen HF, Klemetsdal G, Odegård J, Arnason T. Validation of alternative models in genetic evaluation of racing performance in North Swedish and Norwegian cold-blooded trotters. *J Anim Breed Genet.* 2012;129:164–70.
 33. Gómez MD, Menendez-Buxadera A, Valera M, Molina A. Estimation of genetic parameters for racing speed at different distances in young and adult Spanish trotter horses using the random regression model. *J Anim Breed Genet.* 2010;127:385–94.
 34. Thiruvankadan AK, Kandasamy N, Panneseelvam S. Inheritance of racing performance of thoroughbred horses. *Livest Sci.* 2009;121:308–26.
 35. Vellie BD, Hamilton NA, Wade CM. Heritability of racing performance in the Australian thoroughbred racing population. *Anim Genet.* 2015;46:23–9.
 36. Tozaki T, Miyake T, Kakoi H, Gawahara H, Hirota K, Nakano Y, Kurosawa M. Heritability estimates for racing performance in Japanese thoroughbred racehorses using linear and non-linear model analyses. *J Anim Breed Genet.* 2012;129:402–8.
 37. Vellie BD, Hamilton NA, Wade CM. Performance selection for thoroughbreds racing in Hong Kong. *Equine Vet J.* 2015;47:43–47.
 38. Sharman P, Wilson AJ. Racehorses are getting faster. *Biol Lett.* 2015;11:20150310.
 39. Ricard A, Robert C, Blouin C, Baste F, Torquet G, Morgenthaler C, Rivière J, Mach N, Mata X, Schibler L, Barrey E. Endurance exercise ability in the horses: a trait with complex polygenic determinism. *Front Genet.* 2017;8:89.
 40. Frischnecht M, Signer-Hasler H, Leeb T, Rieder S, Neuditschko M. Genome-wide association studies based on sequence-derived genotypes reveal new QTL associated with conformation and performance traits in the Franches-Montagnes horse breed. *Anim Genet.* 2016;47:227–9.
 41. Schröder W, Klostermann A, Stock KF, Distl O. A genome-wide association study for quantitative trait loci of show-jumping in Hanoverian warmblood horses. *Anim Genet.* 2012;43:392–400.
 42. Lykkjen S, Dolvik NI, McCue ME, Rendahl AK, Mickelson JR, Røed KH. Equine developmental orthopaedic diseases—a genome-wide association study of first phalanx plantar osteochondral fragments in Standardbred trotters. *Anim Genet.* 2013;44:766–9.
 43. Teyssède S, Dupuis MC, Guérin G, Schibler L, Denoix JM, Elsen JM, Ricard A. Genome-wide association studies for osteochondrosis in French trotter horses. *J Anim Sci.* 2012;90:45–53.
 44. McCoy AM, Beeson SK, Splan RK, Lykkjen S, Ralston SL, Mickelson JR, McCue ME. Identification and validation of risk loci for osteochondrosis in Standardbreds. *BMC Genomics.* 2016;17:41.
 45. Lykkjen S, Dolvik NI, McCue ME, Rendahl AK, Mickelson JR, Røed KH. Genome-wide association analysis of osteochondrosis of the tibiotarsal joint in Norwegian Standardbred trotters. *Anim Genet.* 2010;41:111–20.
 46. Dupuis MC, Zhang Z, Druet T, Denoix JM, Charlier C, Lekeux P, Georges M. Results of a haplotype-based GWAS for recurrent laryngeal neuropathy in the horse. *Mamm Genome.* 2011;22:613–20.
 47. Dupuis MC, Zhang Z, Durkin K, Charlier C, Lekeux P, Georges M. Detection of copy number variants in the horse genome and examination of their association with recurrent laryngeal neuropathy. *Anim Genet.* 2013;44:206–8.
 48. Svensk Travsport: Kvalificerings- & premiellopp. <https://www.travsport.se/artikel/kvalificerings-&premiellopp> (2017). Accessed 1 Nov 2017.
 49. Det Norske Travskapsk: Lover & Reglement. <https://www.travsport.no> (2017). Accessed 1 Nov 2017.
 50. R Development Core Team: R-A Language and Environment for Statistical Computing. <https://www.r-project.org/> (2018). Accessed 13 Jan 2018.
 51. Sargolzaei M, Iwaisaki H, Colleau JJ. CFC, a tool for monitoring genetic diversity. In: Proceedings of the 8th world congress on genetics applied to livestock production, vol. 8. Brazil: Belo Horizonte; 2006. p. 27–8.
 52. OFX: Historical Exchange Rates. <https://www.ofx.com/en-au/forex-news/historical-exchange-rates/> (2017). Accessed 1 Nov 2017.
 53. Everitt BS, Hothorn TA. Handbook of statistical analyses using R: Chapman & Hall/CRC; 2010.
 54. Mi H, Huang X, Muruganujan A, Tang H, Mills C, Kang D, Thomas PD. PANTHER version 11: expanded annotation data from gene ontology and Reactome pathways, and data analysis tool enhancements. *Nucl Acids Res.* 2016; <https://doi.org/10.1093/nar/gkw1138>.
 55. Mi H, Muruganujan A, Casagrande JT, Thomas PD. Large-scale gene function analysis with the PANTHER classification system. *Nat Protoc.* 2013;8:1551–66.
 56. Weller JI. Quantitative trait loci analysis in animals. 2nd ed. Oxfordshire: CAB; 2009.
 57. Jäderkvist Fegraeus K, Hirschberg I, Arnason T, Andersson L, Vellie BD, Andersson LS, Lindgren G. To pace or not to pace: a pilot study of four- and five-gaited Icelandic horses homozygous for the *DMRT3* "Gait Keeper" mutation. *Anim Genet.* 2017;48:694–7.
 58. Tang YP, Shimizu E, Dube GR, Rampon C, Kerchner GA, Zhuo M, Liu G, Tsien JZ. Genetic enhancement of learning and memory in mice. *Nature.* 1999;401:63–9.
 59. Zamanillo D, Sprengel R, Hvalby O, Jensen V, Burnashev N, Rozov A, et al. Importance of AMPA receptors for hippocampal synaptic plasticity but not for spatial learning. *Science.* 1999;284:1805–11.
 60. Hu C, Chen W, Myers SJ, Yuan H, Traynelis SF. Human GRIN2B variants in neurodevelopmental disorders. *J Pharmacol Sci.* 2016;132:115–21.

61. Kim JJ, Yoo JH, Kim D, Jeong B, Kim BN. The effects of GRIN2B and DRD4 gene variants on local functional connectivity in attention-deficit/hyperactivity disorder. *Brain Imaging Behav.* 2017; <https://doi.org/10.1007/s11682-017-9690-2>.
62. Concilli M, Iacobacci S, Chesì G, Carissimo A, Polishchuk R. A systems biology approach reveals new endoplasmic reticulum-associated targets for the correction of the ATP7B mutant causing Wilson disease. *Metalomics.* 2016;8:920–30.
63. Kieffer DA, Medici V. Wilson disease: at the crossroads between genetics and epigenetics—a review of the evidence. *Liver Res.* 2017;1:121–30.
64. Usman H, Mathew MK. Potassium channel regulator KCNRG regulates surface expression of shaker-type potassium channels. *Biochem Biophys Res Commun.* 2010;391:1301–5.
65. Lovell PV, Carleton JB, Mello CV. Genomics analysis of potassium channel genes in songbirds reveals molecular specializations of brain circuits for the maintenance and production of learned vocalizations. *BMC Genomics.* 2013;14:470.
66. Gu J, Orr N, Park SD, Katz LM, Sulimova G, MacHugh DE, Hill EW. A genome scan for positive selection in thoroughbred horses. *PLoS One.* 2009;4:e5767.
67. Naylor JM. Hyperkalemic periodic paralysis. *Vet Clin North Am Equine Pract.* 1997;13:129–44.
68. Bowling AT, Byrns G, Spier S. Evidence for a single pedigree source of the hyperkalemic periodic paralysis susceptibility gene in quarter horses. *Anim Genet.* 1996;27:279–81.
69. Naylor JM, Robinson JA, Bertone J. Familial incidence of hyperkalemic periodic paralysis in quarter horses. *J Am Vet Med Assoc.* 1992;200:340–3.
70. Hinchcliff KW, Kaneps AJ, Geor RJ. Musculoskeletal System. In: *Equine sports medicine and surgery*; Saunders Elsevier; 2014. p. 109–43.
71. Cannon SC, Brown RH Jr, Corey DP. A sodium channel defect in hyperkalemic periodic paralysis: potassium-induced failure of inactivation. *Neuron.* 1991;6:619–26.
72. Ricker K, Camacho LM, Grafe P, Lehmann-Horn F, Rüdell R. Adynamia episodica hereditaria: what causes the weakness? *Muscle Nerve.* 1989;12:883–91.
73. Steele D, Naylor JM. Hyperkalemic periodic paralysis, plasma lactate and exercise tolerance. *J Equine Vet Sci.* 1996;20:933–7.
74. Maxson-Sage A, Parente EJ, Beech J, Lindborg S, May LL, Teleis DC. Effect of high-intensity exercise on arterial blood gas tensions and upper airway and cardiac function in clinically normal quarter horses and horses heterozygous and homozygous for hyperkalemic periodic paralysis. *Am J Vet Res.* 1998;59:615–8.
75. van den Bout I, Jones DR, Shah ZH, Halstead JR, Keune WJ, Mohammed S, D'Santos CS, Divecha N. Collaboration of AMPK and PKC to induce phosphorylation of Ser413 on PIP5K1B resulting in decreased kinase activity and reduced PtdIns(4,5)P2 synthesis in response to oxidative stress and energy restriction. *Biochem J.* 2013;455:347–58.
76. Szalinski CM, Guerriero CJ, Ruiz WG, Docter BE, Ribaizi Y, Pastor-Soler NM, Apodaca G, Puthenveedu MA, Weisz OA. PIP5K1 β selectively modulates apical endocytosis in polarized renal epithelial cells. *PLoS One.* 2013;8:e53790.
77. Liu T, Lee SY. Phosphatidylinositol 4-phosphate 5-kinase α negatively regulates nerve growth factor-induced neurite outgrowth in PC12 cells. *Exp Mol Med.* 2013;45:e16.
78. Qin T, An Y, Liu C, Wu J, Dai R, Liu D, Li X, Jiang L, Wu D, Tang X, Song W, Wang T, Zhao X. Novel DOKC8 gene mutations lead to absence of protein expression in patients with hyper-IgE syndrome. *Immunol Res.* 2016;64:260–71.
79. Biggs CM, Lu HY, Turvey SE. Monogenic immune disorders and severe atopic disease. *Nat Genet.* 2017;49:1162–3.
80. Burbank AJ, Shah SN, Montgomery M, Peden D, Tarrant TK, Weimer ET. Clinically focused exome sequencing identifies an homozygous mutation that confers DOKC8 deficiency. *Pediatr Allergy Immunol.* 2016;27:96–8.
81. Zhang Q, Davis JC, Lamborn IT, Freeman AF, Jing H, Favreau AJ, Matthews HF, Davis J, Turner ML, Uzel G, Holland SM, Su HC. Combined immunodeficiency associated with DOKC8 mutations. *N Engl J Med.* 2009;361:2046–55.
82. Pedurupilly CR, Misceo D, Gamage TH, Dissanyayake VH, Frengen E. Postzygotic breakage of a dicentric chromosome results in mosaicism for a telocentric 9p marker chromosome in a boy with developmental delay. *Gene.* 2014;533:403–10.
83. Griggs BL, Ladd S, Saul RA, DuPont BR, Srivastava AK. Dedicator of cytokinesis 8 is disrupted in two patients with mental retardation and developmental disabilities. *Genomics.* 2008;91:195–202.
84. Tassano E, Accogli A, Pavanello M, Bruno C, Capra V, Gimelli G, Cuoco C. Interstitial 9p24.3 deletion involving only DOKC8 and KANK1 genes in two patients with non-overlapping phenotypic traits. *Eur J Med Genet.* 2016;59:20–5.
85. Jäderkvist K, Kangas N, Andersson LS, Lindgren G. Gaitedness is associated with the DMRT3 'Gait keeper' mutation in Morgan and American curly horses. *Anim Genet.* 2014;45:908–9.
86. Jäderkvist K, Holm N, Inmsland F, Armanon T, Andersson L, Andersson LS, Lindgren G. The effect of the DMRT3 'Gait keeper' mutation on gait traits and gaits in Standardbred and Icelandic horses. *Livest Sci.* 2015;176:33–9.
87. Regatieri IC, Eberth JE, Sarver F, Lear TL, Bailey E. Comparison of DMRT3 genotypes among American Saddlebred horses with reference to gait. *Anim Genet.* 2016;47:603–5.
88. Promerová M, Andersson LS, Juras R, Penedo MC, Reissmann M, Tozaki T, et al. Worldwide frequency distribution of the 'Gait keeper' mutation in the DMRT3 gene. *Anim Genet.* 2014;45:274–82.
89. Vasan RS, Glazer NL, Felix JF, Lieb W, Wild PS, Felix SB, et al. Genetic variants associated with cardiac structure and function: a meta-analysis and replication of genome-wide association data. *JAMA.* 2009;302:168–78.
90. Netherton SJ, Jimmo SL, Palmer D, Tilley DG, Dunkerley HA, Raymond DR, Russell JC, Absher PM, Sage EH, Vernon RB, Maurice DH. Altered phosphodiesterase 3-mediated cAMP hydrolysis contributes to a hypermotile phenotype in obese JCRLA-cp rat aortic vascular smooth muscle cells: implications for diabetes-associated cardiovascular disease. *Diabetes.* 2002;51:1194–200.
91. Boda H, Uchida H, Takaiso N, Ouchi Y, Fujita N, Kuno A, Hata T, Nagatani A, Funamoto Y, Miyata M, Yoshikawa T, Kurahashi H, Inagaki H. A PDE3A mutation in familial hypertension and brachydactyly syndrome. *J Hum Genet.* 2016;61:701–3.
92. Campos-Chillon F, Farmerie TA, Bouma GJ, Clay CM, Carnevale EM. Effects of aging on gene expression and mitochondrial DNA in the equine oocyte and follicle cells. *Reprod Fertil Dev.* 2015;27:925–33.
93. Hamilton MJ, Halvorsen EC, LePard NE, Bosiljic M, Ho VW, Lam V, Banath J, Benneweth KL, Krystal G. SHIP represses lung inflammation and inhibits mammary tumor metastasis in BALB/c mice. *Oncotarget.* 2016;7:3677–91.
94. Somasundaram R, Fernandes S, Deuring JJ, de Haar C, Kuipers EJ, Vogelaar L, Middleton FA, van der Woude CJ, Peppelenbosch MP, Kerr WG, Fuhler GM. Analysis of SHIP1 expression and activity in Crohn's disease patients. *PLoS One.* 2017;12:e0182308.
95. Pauls SD, Ray A, Hou S, Vaughan AT, Cragg MS, Marshall AJ. FcyRIIB-independent mechanisms controlling membrane localization of the inhibitory phosphatase SHIP in human B cells. *J Immunol.* 2016;197:1587–96.
96. Eijgelsheim M, Newton-Cheh C, Sotoodehnia N, de Bakker PI, Müller M, Morrison AC, et al. Genome-wide association analysis identifies multiple loci related to resting heart rate. *Hum Mol Genet.* 2010;19:3885–94.
97. Pfeuffer A, van Noord C, Marcianti KD, Arking DE, Larson MG, Smith AV, et al. Genome-wide association study of PR interval. *Nat Genet.* 2010;42:153–9.
98. Della-Morte D, Beecham A, Rundek T, Wang L, McClendon MS, Slifer S, Blanton SH, Di Tullio MR, Sacco RL. A follow-up study for left ventricular mass on chromosome 12p11 identifies potential candidate genes. *BMC Med Genet.* 2011;12:100.
99. Hersh CP, Silverman EK, Gascon J, Bhattacharya S, Klanderma BJ, Litonjua AA, Lefebvre V, Sparrow D, Reilly JJ, Anderson WH, Lomas DA, Mariani TJ. SOX5 is a candidate gene for chronic obstructive pulmonary disease susceptibility and is necessary for lung development. *Am J Respir Crit Care Med.* 2011;183:1482–9.
100. Rakhmanov M, Sic H, Kienzler AK, Fischer B, Rizzi M, Seidl M, et al. High levels of SOX5 decrease proliferative capacity of human B cells, but permit plasmablast differentiation. *PLoS One.* 2014;9:e100328.
101. Li A, Hooli B, Mullin K, Tate RE, Bubnys A, Kirchner R, Chapman B, Hofmann O, Hide W, Tanzi RE. Silencing of the *Drosophila* ortholog of SOX5 leads to abnormal neuronal development and behavioral impairment. *Hum Mol Genet.* 2017;26:1472–82.
102. Lamb AN, Rosenfeld JA, Neill NJ, Talkowski ME, Blumenthal I, Girirajan S, et al. Haploinsufficiency of SOX5 at 12p12.1 is associated with developmental delays with prominent language delay, behavior problems, and mild dysmorphic features. *Hum Mutat.* 2012;33:728–40.
103. Powers SK, Jackson MJ. Exercise-induced oxidative stress: cellular mechanisms and impact on muscle force production. *Physiol Rev.* 2008;88:1243–76.
104. Lee HJ, Baek SS. Role of exercise on molecular mechanisms in the regulation of antidepressant effects. *J Exerc Rehabil.* 2017;13:617–20.

RESEARCH ARTICLE

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A QTL for conformation of back and croup influences lateral gait quality in Icelandic horses

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Abstract

Background: The back plays a vital role in horse locomotion, where the spine functions as a spring during the stride cycle. A complex interaction between the spine and the muscles of the back contribute to locomotion soundness, gait ability, and performance of riding and racehorses. Conformation is commonly used to select horses for breeding and performance in multiple horse breeds, where the back and croup conformation plays a significant role. The conformation of back and croup plays an important role on riding ability in Icelandic horses. However, the genes behind this trait are still unknown. Therefore, the aim of this study was to identify genomic regions associated with conformation of back and croup in Icelandic horses and to investigate their effects on riding ability. One hundred seventy-seven assessed Icelandic horses were included in the study. A genome-wide association analysis was performed using the 670 K+ Axiom Equine Genotyping Array, and the effects of different haplotypes in the top associated region were estimated for riding ability and additional conformation traits assessed during breeding field tests.

Results: A suggestive quantitative trait loci (QTL) for the score of back and croup was detected on *Equus caballus* (ECA) 22 (p -value = 2.67×10^{-7}). Haplotype analysis revealed two opposite haplotypes, which resulted in higher and lower scores of the back and croup, respectively (p -value < 0.001). Horses with the favorable haplotype were more inclined to have a well-balanced backline with an uphill conformation and had, on average, higher scores for the lateral gaits tölt (p -value = 0.02) and pace (p -value = 0.004). This genomic region harbors three genes: *C20orf85*, *ANKRD60* and *LOC100056167*. *ANKRD60* is associated with body height in humans. *C20orf85* and *ANKRD60* are potentially linked to adolescent idiopathic scoliosis in humans.

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Conclusions: Our results show that the detected QTL for conformation of back and croup is of importance for quality of lateral gaits in Icelandic horses. These findings could result in a genetic test to aid in the selection of breeding horses, thus they are of major interest for horse breeders. The results may also offer a gateway to comparative functional genomics by potentially linking both motor laterality and back inclination in horses with scoliosis in humans.

Keywords: Back, Backline, Conformation, Croup, High-density genome scan, Icelandic horse, Lateral gait quality, Novel QTL

Background

Associations of body measurements with locomotor health and sports performance have been reported in many different breeds, including Icelandic horses [1–11]. Discriminant analyses have shown that several morphological features distinguish with high accuracy between low-class and high-class Icelandic horses with respect to different riding ability traits [3]. The most important features for gait ability in Icelandic horses are the height of the horse at front compared to hind (uphill conformation) with well-balanced backline, croup proportions and width of chest [1, 3]. The analyses also indicated the disadvantage of a forward inclination in the back or a sway back [3]. Conformation of the back and croup thus play a major role on riding ability in Icelandic horses.

The Icelandic horse official breeding goal promotes five-gaited horses with a functional and aesthetically pleasing conformation [12]. Zoometric measurements and subjective scores for conformation and riding ability traits are recorded at breeding field tests. Genetic correlations between conformation of back and croup, and gait qualities have been estimated as moderate to high (0.19–0.54) [1]. Furthermore, moderate heritabilities (0.29–0.31) have been estimated for the subjectively scored back and croup trait [1, 13] and the objectively measured zoometric traits pertaining to conformation of back and croup (0.20–0.25) [3]. For the subjectively scored riding ability traits, the heritability estimates range from 0.18 (walk) to 0.60 (pace) [1, 13].

Despite conformation traits being moderately heritable in the Icelandic horse, only mutations in the Myostatin gene have previously been associated with conformation traits, i.e. estimated breeding values of neck, withers and shoulders [14]. In other horse breeds, as well as other species, many different genes have been shown to influence body size. *LCORL*, *NCAPG* and *HMGA2* are major genes known to regulate body size in mammals including humans, cattle, sheep, dogs and horses [15–23]. These genes, along with other genes such as *ZFAT* and *LASPI*, affect not only the body size of the horse but more specifically the height at withers [15, 24, 25]. Three novel missense variants located in the *ADAMTS17*, *OSTN* and *GHI* genes explained 61% of the variance of

withers height in Shetland pony-related breeds [26]. Other additional quantitative trait loci have also shown significant associations with morphometric angular measurements, with regions on chromosomes ECA28 and ECA29 associated with poll angle in horses [27]. However, the genes behind many other conformation traits are still unknown.

Considering the heritability of conformation of back and croup and its genetic correlation with riding ability, we hypothesized that major genetic factors of importance for back and croup also influence gait quality in Icelandic horses. Therefore, the aim of this study was to identify genomic regions associated with conformation of back and croup in Icelandic horses and investigate their effects on riding ability traits assessed at breeding field tests.

Results

Genome-wide association analysis for conformation of back and croup

In total, 383,896 SNPs (373,041 autosomal and 10,855 X chromosomal) and 177 horses passed QC and were included in the GWA analysis. Thirteen SNPs located on ECA22: 45347522–45,662,708 reached the suggestive threshold ($p < 1.0 \times 10^{-5}$) of which ten were in LD ($r^2 \geq 0.8$) (Fig. 1). Additionally, one single SNP reached the suggestive threshold on ECA12 (Fig. 1). A summary of the GWA results for the 50 top SNPs is presented in Additional file 1.

Haplotype analysis

The haplotype analysis revealed two opposite haplotypes which resulted in higher and lower scores for back and croup (p -value < 0.001) (Table 1). Thirty-four horses were homozygous for the haplotype associated with a higher score and 28 horses homozygous for the haplotype associated with a lower score of back and croup. Five different haplotypes were estimated (Table 1). Haplotypes determined to be too rare to estimate their specific regression coefficients were pooled into a separate group with a frequency of 0.07 (results not presented).

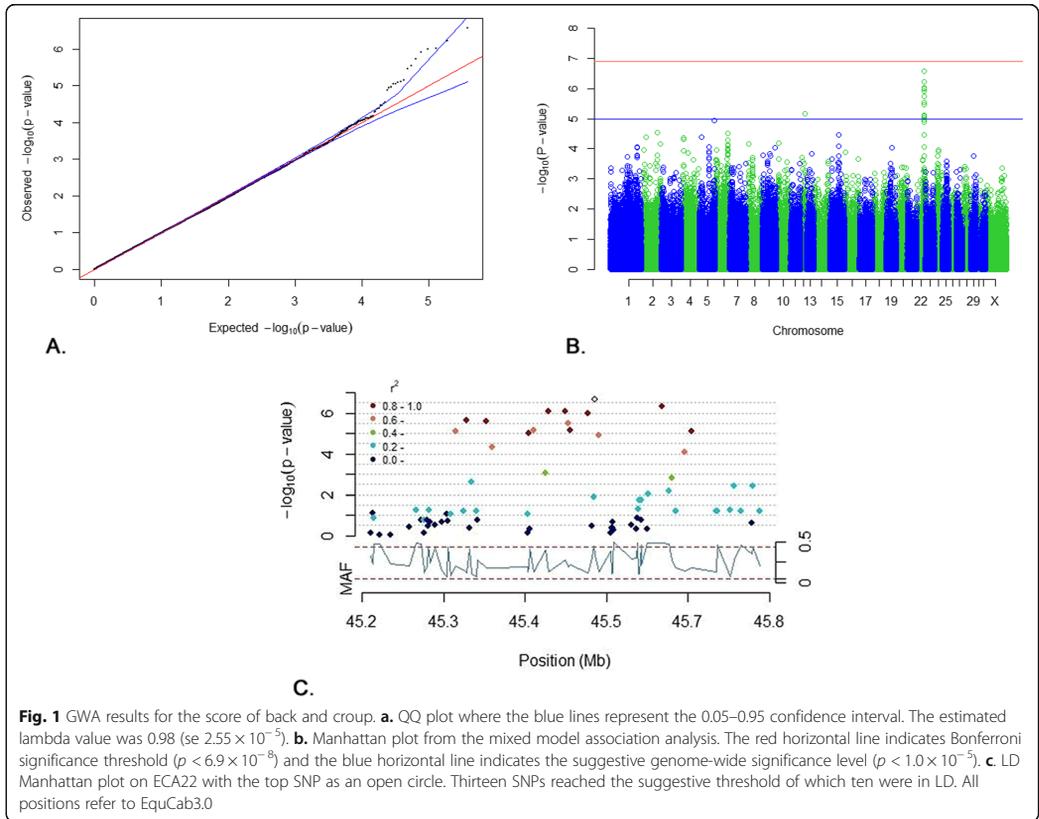


Table 1 Results from haplotype analysis for the score of back and croup

Haplotypes (SNPs numbers ^a)										Coef	Freq	<i>p</i> -value	Sim. <i>p</i> -value
1	2	3	4	5	6	7	8 ^a	9	10				
G	T	C	A	T	A	T	A	A	T	-0.300	0.383	< 0.001	< 0.001
G	T	C	A	T	A	T	A	G	C	0.090	0.021	0.657	0.718
G	T	C	A	G	G	G	A	A	T	0.119	0.027	0.518	0.889
G	C	T	C	T	A	G	A	A	T	0.090	0.025	0.626	0.963
A	C	T	C	G	G	G	G	G	C	0.300	0.474	< 0.001	< 0.001

Sim. *p*-value = *p*-value adjusted by using 100,000 permutations
 Significant results in bold
 Coef. coefficient, estimated effect of the haplotype on the score of back and croup from the glm model in the haplotype analysis
 Freq. frequencies
^aSNP numbers in bp position order with top SNP as number 8 with reference allele A and alternate allele G

Phenotype association of the haplotypes with a significant effect on the score of back and croup

The t-test analyses revealed that several traits in addition to back and croup significantly differed in mean scores between horses with the favourable and unfavorable haplotype. The two haplotype groups differed significantly in mean scores (*p*-value ≤ 0.05) for the gait traits tölt and pace (Table 2). The two haplotype groups also differed significantly in means for the zoometric measurements of depth at breast, width of hips and thigh bones, and length of the forelimbs. In addition to this, there were significant differences between the two haplotype groups for the sub-traits backline and the croup type.

Allele frequency of top SNP and DMRT3 in different breeds

Comparing allele frequencies of the top SNP identified from GWA analysis between different breeds revealed a higher frequency of the alternate allele (the favorable

Table 2 Significant results from *t*-test comparing phenotypes in horses with different haplotypes

Trait	Favorable haplotype		Unfavorable haplotype		t-value	df	p-value
	N	Mean	N	Mean			
Back and croup	34	8.29	28	7.71	4.05	58.08	< 0.001
Tölt ^a	33	8.41	27	7.96	2.52	45.79	0.015
Pace ^a	33	7.18	27	6.09	2.99	50.24	0.004
Slow tölt ^a	33	8.14	26	7.73	2.14	45.19	0.038
Depth at breast (M4) ^b	33	63.2	28	64.6	-3.52	56.22	0.001
Width of the hips (M7) ^b	23	47.0	20	48.1	-2.21	37.54	0.033
Width between thigh bones (M8) ^b	23	43.0	20	44.2	-2.23	38.86	0.031
Length of forelimbs (M1-2xM4) ^b	33	15.2	28	12.1	3.22	40.81	0.003
Backline ^c	34	1.79	28	2.25	-2.69	58.91	0.009
Croup type ^c	34	1.85	28	2.18	-2.31	53.23	0.025

N Number of horses

^aSubjectively assessed traits (scale 5–10)

^bZoometric measurements (cm)

^cSubjectively assessed sub-traits (scale 1–3)

allele) in the Icelandic breed compared with all other investigated breeds (Table 3).

Functional annotation of genes in the region associated with the score of back and croup

The detected QTL ECA22: 45347522–45,662,708 harbors the genes *Chromosome 22 C20orf85 homolog (C22H20orf85)*, *Ankyrin repeat domain 60 (ANKRD60)* and *LOC100056167* described as *serine/threonine-protein phosphatase 4 regulatory subunit 1*. The SNP on ECA12 (position 26,756,656–26,756,656) was located close to the gene *solute carrier family 22 member 8 (SLC22A8)*. None of the significant SNPs (on ECA12 and 22) overlapped any known QTL for conformation in horses [39].

Discussion

Conformation of the back and croup plays an important role for riding ability, gait ability, welfare, and longevity of the horse [1, 3, 13, 40]. The present study was performed to identify genomic regions associated with conformation of the back and croup in Icelandic horses and investigate their effects on riding ability. A novel QTL was detected on ECA22 with candidate genes associated with scoliosis and anthropometric traits in humans [41, 42]. Our results show that this QTL is of importance not only for conformation of back and croup, but also for riding ability traits, especially lateral gait quality, in Icelandic horses.

Possible links between scoliosis, motor laterality and lateral gaits

The detected QTL for the trait back and croup harbors the genes *C22H20orf85* and *ANKRD60*, both of which are potentially linked to adolescent idiopathic scoliosis (AIS) in humans [41]. Scoliosis is defined as a lateral curvature of the spine and it is the most common

vertebral disorder in children and adolescents [43]. In humans, scoliosis can be caused by muscular dystrophy or cerebral palsy, but the cause is usually unknown and therefore referred to as idiopathic [43]. AIS in humans has been shown to result in a generalized skeletal muscle weakness, respiratory impairment and exercise limitation [44]. Studies on scoliosis in humans have also shown correlation between handedness and truncal asymmetry [45–49] and that molecular basis of handedness are more likely formed by spinal gene expression asymmetries rather than in the motor cortex [50]. Symptoms of scoliosis in horses has been described as an S-shaped bend of the caudal thoracic vertebral column, resulting in restricted movements of the hind limbs and inflexibility of the back [51]. Another report described symptoms as a lateral deviation of the head and cervical and cranial thoracic vertebral column to one side, and associated rotation of the thoracic vertebrae. These deviations result in difficulties for a horse to walk in a straight line [52]. However, severe thoracic vertebral malformations in horses are infrequent, and mild to moderate forms of scoliosis may go undetected as the strong dorsal spinal musculature can mask subtle deviations of the vertebral column [52]. Scores for conformation of back and croup in horses involve both muscular and skeletal assessments, which may indicate that the back and croup phenotype shares some features with mild forms of scoliosis. It is well known that horses commonly demonstrate motor laterality (handedness) [53–55] and some even have difficulties walking in a straight line at the beginning of training. The latter often need more time in training to improve their balance and straightness.

In general, disorders of the back appear to be relatively common in horses and lead to pain and decreased performance [51]. However, to our knowledge, there are no

Table 3 Allele frequency of top SNP for back and croup and *DMRT3*

Breed	Top SNP			<i>DMRT3</i>		
	N	AF alt	Source	N	AF alt	Source
Icelandic horses included in present study ^a	177	0.50	Array genotyping	177	0.94	Array genotyping
Icelandic horses unassessed ^b	49	0.51	SNP genotyping	49	0.90	SNP genotyping
Other gaited breeds						
Rocky-Mountain	36	0.33	SNP genotyping	27	1	SNP genotyping
Colombian paso horses	37	0.24	Array genotyping	37	0.0	SNP genotyping
Colombian trocha						
Colombian trot and gallop	11	0.23	Array genotyping	11	0.0	SNP genotyping
Colombian paso fino	38	0.29	SNP genotyping	28	1	[28]
Partly gaited breeds						
American Curly	27	0.32	SNP genotyping	101	0.70	[29]
American Saddlebred	42	0.29	SNP genotyping	89	0.28	[30]
Morgan	30	0.44	SNP genotyping	59	0.14	[29]
Non- gaited breeds						
Exmoor	279	0.01	[31]	27	0.0	[31]
Connemara Pony	40	0.05	[32]	35	0.0	[30]
Swedish Warmblood	379	0.26	[33]	64	0.0	[30, 34]
Thoroughbred racehorses	370	0.14	[35]	55	0.0	[30, 34]
Persian-Arabian horses	101	0.32	[36]	69	0.0	[30]
North-Swedish draught	25	0.38	[37]	34	0.0	[30, 34]
Harness racing breeds						
Coldblooded trotters	565	0.13	[38]	306	0.45	[30]
Standardbred	40	0.29	SNP genotyping	270	0.97	[30, 34]

N number of horses included in dataset

Top SNP the top SNP identified from the GWA analysis for back and croup

AF alt frequency of alternate allele

DMRT3 AF alt allele frequency of the alternate allele A in the *DMRT3* gene known as the “Gait Keeper” mutation

^aThe 177 Icelandic horses included in the present study

^bIcelandic horses used for riding but that had not attended breeding field test

studies reporting the prevalence of back problems or scoliosis in Icelandic horses, and it is generally hard to diagnose back pain in horses. The effect of the QTL is more likely related to functional advantage or disadvantage for movements and strength of the back and croup in horses rather than the result of more severe dysfunctions and pain. This is supported by the relatively high frequency of the unfavorable haplotype among the Icelandic horses in the present study.

Top SNP allele frequency in other breeds

Icelandic horses had a higher frequency of the alternate allele (the favorable allele) of the top SNP for back and croup compared with all other investigated breeds, including the other gaited and partly gaited breeds. In addition, the Icelandic horses with the favorable haplotype had on average higher scores for the lateral gaits tölt and pace. Therefore, it is likely that the quality of the lateral gaits rather than the ability to perform the gaits is affected by the QTL. Almost all Icelandic horses

carry at least one copy of the mutant allele A in the *DMRT3* gene known as the “Gait Keeper” mutation [30, 34]. This mutation is known to affect the pattern of locomotion in horses and the ability to perform lateral gaits [34]. The Icelandic horses in the present study had a high frequency of the *DMRT3* “Gait Keeper” mutation (0.94), 157 of the 177 horses were homozygous AA. The *DMRT3* genotype was taken into account in the phenotype association analysis. Pace scores in horses with the CA genotype were considered as a missing value. Despite this, the Icelandic horses with the favorable haplotype had higher scores for pace. This further supports our hypothesis that the detected QTL affects the quality and not the ability of lateral gaits. The genotyped gaited breed Rocky-Mountain Horse is known to be fixed for the *DMRT3* “Gait Keeper” mutation [30]. The other genotyped gaited breeds American Curly, American Saddlebred and Morgan horses have a moderate high frequency of the *DMRT3* “Gait Keeper” mutation [30, 34, 56]. These breeds are considered as partly gaited as not all horses within the breed perform ambling gaits. Trotters are also known to

perform lateral gaits, and the reported frequency of the *DMRT3* mutation is high in Standardbreds (0.97–1.00) [30, 34] and relatively high in Coldblooded trotters (0.45) [30]. All of these gaited and partly gaited breeds had a higher frequency of the reference allele than the alternate allele for the top SNP of back and croup. The genotyped Colombian paso horses (CPH) included a group of horses that perform trocha and one group that only perform trot and gallop. The trocha gait is defined as a four-beat gait that includes a lateral step but it is diagonally coupled and therefore not considered a lateral gait [28, 57]. The allele frequency of the top SNP did not differ between these two groups. A group of CPH that perform the lateral gait *paso fino* was also genotyped. However, like all the other genotyped breeds, this group had a lower frequency of the alternate allele of the top SNP for back and croup compared to the Icelandic horses. None of the other genotyped breeds in this study segregates for the *DMRT3* mutation [30, 34], nor do they perform lateral gaits.

The 49 unassessed Icelandic horses had a similar allele frequency of the top SNP for back and croup as well as for the *DMRT3* mutation as the 177 assessed Icelandic horses included in the present study. The unassessed group included riding school horses and horses used for hobby riding. It could be argued that balance and straightness is even more essential for the training of Icelandic horses as they carry relatively heavy (adult) riders, relative to their size, in lateral gaits such as tölt and pace with strong focus on the gait quality. In addition, the Icelandic horses with the favorable haplotype had higher average scores for the lateral gaits tölt and pace, which are highly valued traits in the breed. It is likely that there has been selection for the alternate allele of the top SNP in Icelandic horses.

Genes within the QTL associated with musculoskeletal traits

The gene *ANKRD60* is associated with body height in humans [42] and a recent study in American Miniature Horses reported a QTL for withers height close to another Ankyrin Repeat Domain gene *ANKRD1* [58]. The QTL region on ECA22 harbors the gene *LOC100056167* that is not well annotated in horses. The gene is described as serine/threonine-protein phosphatase 4 regulatory subunit 1 and appears to blast with the pseudogene *PPP4R1L* in humans with 84.17% identity [59]. The pseudogene *PPP4R1L* is transcribed in humans and *LOC100056167* has exons. *PPP4R1L* has a potential effect on bone mineral density as it has a protein phosphatase regulator activity [60]. *PPP4R1L* is regulated by an enhancer (Genehancer ID GH20J058887) with potential implications on body height and BMI-adjusted waist circumference in humans [61, 62]. Therefore, it is possible that the detected QTL effects both the muscular and skeletal system.

The horses with the favorable haplotype in the present study had longer forelimbs than those with the unfavorable haplotype. This may be explained, at least to some extent, by the effects of the genes *ANKRD60* and *LOC100056167*. According to a previous study, high-class Icelandic horses are distinguished from low-class horses by an uphill conformation [3]. High-class horses have higher withers and higher set neck and back, compared to height at croup and tuber coxae [3]. Uphill conformation is believed to facilitate ease of collection and lightness in the front part, features that are taken into account when gait quality is subjectively assessed at breeding field tests [12]. Stride length is associated with limb length in horses and other species [63–65] and stride length is also taken into account when assessing the gait quality at breeding field tests [12]. Consequently, stride length and uphill conformation are important factors for higher gait quality scores, both of which may be connected to longer forelimbs. This further supports the results from this study as the horses with the favorable haplotype had both longer forelimbs and higher scores for tölt and pace. In line with this, the horses with the unfavorable haplotype also had a deeper breast and more negative standardized marks for the sub-trait backline compared with the ones with the favorable haplotype. This indicates that a downhill conformation is more common in horses with the unfavorable haplotype. It is possible that a downhill inclination creates an imbalance between the front and back of the horse, causing difficulties for the horse to stretch the hind legs forward, thus losing the ability for self-carriage and collection. This may also result in a shorter stride length, causing lower scores for tölt and pace.

Length and form of the croup are also known to discriminate between high-class and low-class Icelandic horses [3]. In the present study, horses with the favorable haplotype had more positive standardized marks for the sub-trait croup type. This trait is defined as how evenly the croup is shaped and suggests that the haplotype does not influence the length or inclination of the croup, but only the shape of it. The difference between the two haplotype groups for the width of hips (M7) and width between the thighbones (M8) suggest that horses with the favorable haplotype may have a slimmer framed croup than horses with the unfavorable haplotype.

Complexity of the phenotype

Until around year 2010, a soft, lower backline was considered to be favorable for the assessment of back and croup of Icelandic horses, as a low position of the back was assumed desirable for tölt [12]. A study in American Saddlebred horses detected a region on ECA20 associated with extreme lordosis (swayback) [66]. However, in the present study no significant association with back

and croup was detected on ECA20. Horses with the haplotype associated with lower score of back and croup were more inclined to have a forward sloping and/or swayback backline.

The back and croup is a complex trait, with muscular as well as skeletal features of both the back and the croup subjectively assessed and scored together as a single trait. Our results show that the novel detected QTL associated with back and croup conformation influences various riding ability and conformation traits. It should be noted that the complex conformation and riding ability traits are likely to be influenced by many different genes as well as environmental factors such as feeding and training. Therefore, further studies are needed to determine the effects of this newly discovered QTL.

Conclusions

This study provides valuable information about the genetics of conformation of the back and croup in Icelandic horses. A novel QTL for the trait back and croup was detected on ECA22: 45347522–45,662,708. The QTL is associated with the back inclination, the form of the croup, and length of limbs as well as the quality of the lateral gaits pace and tölt. These findings could result in the offering of a genetic test to aid in the selection of breeding horses, thus they are of major interest for horse breeders. The genomic region harbors genes associated with scoliosis and anthropometric traits in humans. The findings could serve as a platform to study any potential link between scoliosis and motor laterality in horses and other species. Further analyses are needed to fully understand the biological function of this genomic region on the conformation of back and croup and its influence on gait quality.

Methods

Animals

In total, 177 Icelandic horses (77 males and 100 females) born between 1993 and 2014 were included in the study. Hair samples were collected at breeding field tests and by visiting trainers and breeders in Iceland and Sweden. A few samples were also sent in by horse owners after personal contact and posting on social media. Only privately owned horses participated in the study and the horses were not specifically selected based on conformation of back and croup. Pedigree data were obtained from the international Icelandic horse database Worldfengur [67]. Maximum relatedness between horses was limited to half-siblings.

Phenotyping

Phenotype data were obtained from the international Icelandic horse database Worldfengur [67]. The phenotype used for the genome-wide association (GWA)

analysis consisted of the subjectively assessed score for back and croup recorded at breeding field tests between 1999 and 2018. Additional conformation and riding ability traits assessed at breeding field tests were used to investigate the effects of genomic regions detected from GWA analysis. Of the 177 horses had 115 attended more than one breeding field test. For these horses, information from the latest assessment was used. The majority of horses were assessed in year 2018 ($n = 89$). The horses were assessed in Iceland ($n = 81$), Sweden ($n = 87$), Germany ($n = 3$), Denmark ($n = 2$) and Norway ($n = 4$). Icelandic horses can attend breeding field test from when they are 4 years old. The age of assessment was on average 6.7 years and ranged from 4 to 15 years. In our sample, 173 horses were assessed for both conformation and riding ability traits, and 4 horses were only assessed for conformation traits as the ridden test is optional. Pace scores for horses with the CA genotype for the DMRT3 gene ($n = 20$) were treated as missing values.

Back and croup

Back and croup, along with other conformation and riding ability traits assessed at breeding field tests, were subjectively scored on a scale from 5 to 10 with 0.5 intervals, where a score of 5 was only given if a trait was not presented. Assessment of the trait back and croup comprises several aspects of the conformation of the back, croup and loins. The slope and shape of the backline, which is defined as the line from the base of withers to the lumbosacral joint, were assessed. Length and slope of the croup were also assessed, as well as the width and muscularity of the back, the length and width of the loins and the form and muscularity of the croup [12]. A high score for back and croup represents a strong, well-balanced backline and a well-muscled wide back. The croup should be long, evenly formed, well-muscled and adequately sloping. A low score is associated with a swayback, stiff or forward sloping backline, a too short or too long and/or unevenly formed croup and poorly muscled back and croup [12]. When the judging panel has reached a consensus on a score for back and croup according to the judging scale, they have the possibility to use standardized marks to describe the most prominent positive and/or negative attributes of the trait.

Pictures with examples of horses representing high and low score for back and croup are presented in Fig. 2. The 177 horses in the study had a score of back and croup that ranged from 6.5 to 9.0 with a mean value of 8.1 (SD 0.56) (Fig. 3). The distribution of the scores for back and croup was slightly negatively skewed (coefficient of skewness -0.36). Transformation of the raw data to increase normality was tested but was found to not affect the results. Moreover, the residuals from the

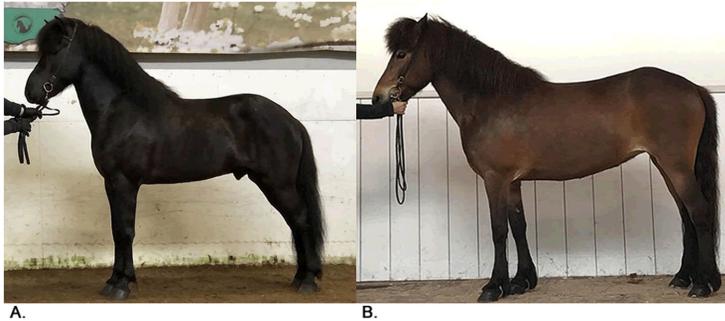


Fig. 2 Examples of Icelandic horses representing high and low score of back and croup. **a.** Icelandic horse that represents high score of back and croup. The backline is well-balanced and the back is wide and well-muscled. (Photo: Hrefna María Ómarsdóttir). **b.** Icelandic horse that represents a low score of back and croup with a forward sloping backline and a less muscled croup. (Photo: The Swedish Icelandic Horse Association, SIF). Images used in the figure are privately owned and thus were not taken from previously published sources requiring written permission for use

linear models were normally distributed (results not presented).

Sub-traits based on standardized marks for back and croup

For the purpose of more detailed analysis of the score for back and croup, the standardized marks used to describe prominent positive and negative attributes of the trait were defined as two different sub-traits; backline and croup type. These sub-traits were analysed on a linear scale ranging from 1 to 3, where 1 represented a positive mark, 3 represented a negative mark and 2 represented no mark and was interpreted as an intermediate description of the trait (not positive or negative). A positive mark for the sub-trait backline was given for good

backline (well-balanced backline) and the options for negative marks were forward sloping back, straight back, sway back and/or stiff loins. For the sub-trait croup type, a positive mark was given for evenly formed croup and the options for negative marks were rounded croup, narrowing croup, roof-shaped croup and/or coarse croup.

Additional trait assessment scores from breeding field tests

Besides the conformation trait back and croup, scores for the gait traits tölt, slow tölt, trot, pace, gallop, canter and walk and the trait form under rider were included in this study. Features of each gait such as beat, suppleness, stride length, leg-action, speed capacity, collection and lightness were taken into account when assessing the

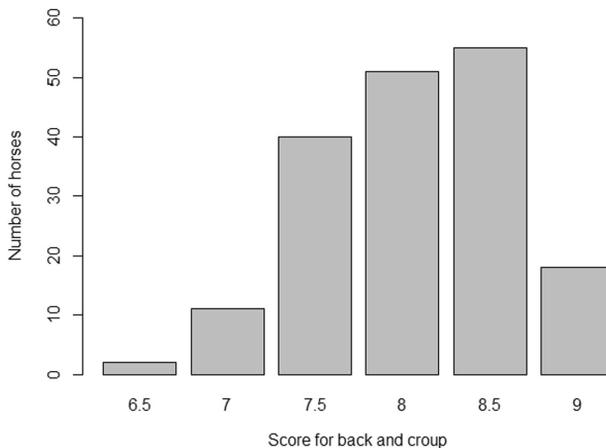


Fig. 3 Distribution of scores for back and croup in the 177 horses. (Picture generated from the data analysis in R)

gaits [12]. Scores of all these traits were included to investigate the effects of the detected regions from GWA analysis on the trait back and croup.

Zoometric traits measured at breeding field tests

Zoometric measurements are traditionally recorded at breeding field tests to corroborate the subjective conformation assessments [12]. All these measurements were included to investigate the effects of the detected genomic regions from GWA analysis for the trait back and croup. The measurements consisted of height at withers (M1), height at lowest point of back (M2), height at croup (M3), depth of breast (M4), length of body from the point of shoulder to tuber ischii (M5), width of chest between the points of the shoulders (M6), width of the hips between the tuber coxae (M7) and width of the hips between the hip joints (M8) (Fig. 4). Length of forelimbs is traditionally assessed from calculation of the difference between height at withers and depth at breast times two (M1-2xM4), as it gives better comparison of the leg length to consider the variation in breast depth between different horses. Other calculated measurements used for conformation assessments were difference between height at withers and height at back (M1-M2), difference between height at withers and height at croup (M1-M3), difference between height at croup and height at back (M3-M2), difference between length of the horse and height at withers (M5-M1), difference between length of the horse and height at croup (M5-M3) and difference between width of hips and width between thigh bones (M7-M8).

DNA isolation

DNA was extracted from hair roots using a standard procedure of hair preparation. One hundred eighty-six microlitre of 5% Chelex® 100 Resin (Bio-Rad Laboratories, Hercules, CA) and 14 µL of proteinase K (20 mg/mL;

Merck KgaA, Darmstadt, Germany) were added to each sample. This mix was incubated at 56 °C for 2 h at 600 rpm and proteinase K was inactivated for 10 min at 95 °C.

Genotyping and quality control

The 177 Icelandic horses were genotyped on the 670 K+ Axiom Equine Genotyping Array. Quality control (QC) was performed with the package GenABEL [69] in R [70] to remove poorly genotyped and noisy data based on the following thresholds: missing genotypes per single nucleotide polymorphism (SNP) (> 0.10), missing SNPs per sample (> 0.10), minor allele frequency (MAF) (< 0.05) and Hardy-Weinberg equilibrium (p -value $1e^{-10}$).

Genome-Wide Association Study (GWAS)

GWA analyses were performed using the package GenABEL [69] in R [70]. Possible fixed effects were tested in a linear model using anova as a post hoc test. The tested fixed effects were sex (male or female), age at assessment in age classes (4, 5, 6 or ≥ 7 years old), age at assessment in years as a linear regression, country of assessment in two classes (Iceland or Sweden/other countries) and year of assessment in five classes (< 2010, 2010–2015, 2016, 2017 or 2018). The division of year of assessment classes was based on change in how the back and croup phenotype was to be assessed, and number of horses in the data from different years. The DMRT3 genotype was also tested as an effect. None of these fixed effects were found to be significant ($p \leq 0.05$) for the trait back and croup and were thus not included in the GWA models. To investigate potential stratification, a multidimensional scaling (MDS) plot was constructed based on a genomic relationship matrix using the GenABEL package and `ibs()` function [69]. No outliers were apparent on the MDS plot and no stratification of horses with low and high score of back and croup was detected. A

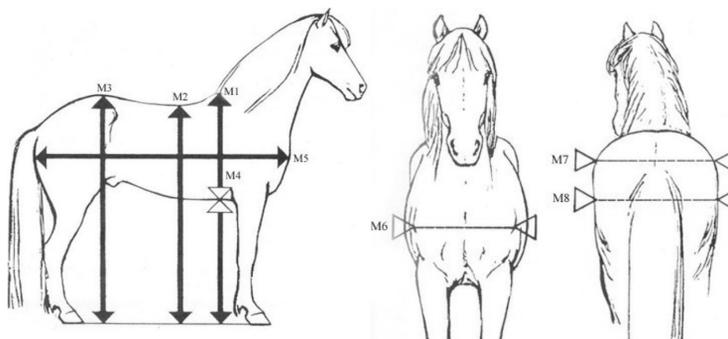


Fig. 4 Zoometric measurements recorded at standardized breeding field tests for Icelandic horses [68]. Original images created by Pétur Behrens. Images used in the figure are privately owned and thus were not taken from previously published sources requiring written permission for use

visualization of the genomic-kinship matrix using MDS is shown in Fig. 5.

The genomic-kinship matrix together with the phenotype of back and croup were passed to the polygenic_hglm function using family gaussian in GenABEL [69, 71]. To account for any population stratification, the GWA analysis was performed using a mixed model-structured association approach with the mmscore function in GenABEL [69]. Genome-wide significance was determined by Bonferroni correction and a suggestive genome-wide significance threshold was set at 1.0×10^{-5} [72, 73]. QQ and linkage disequilibrium (LD) manhattan plots were performed using the package cgmisc 2.0 [74].

Haplotype analysis

Haplotype analysis was performed with the haplo.stats package in R [70]. A linkage disequilibrium plot was constructed and the ten significant SNPs in LD ($r^2 \geq 0.8$) were used in the function haplo.em to estimate haplotypes. The haplotype effect on the score of back and croup was estimated by a generalized linear model (glm) with the function haplo.glm. The most frequent haplotype was used as a reference and only haplotypes with frequencies greater than 0.02 were included. A simulated p -value was estimated by using 100,000 permutations considering an additive effect.

Phenotype association of significant haplotypes

Phenotype association of the horses homozygous for the haplotypes that had a significant effect on the conformation of back and croup was performed using a two-tailed Student's t -test in R [70]. Significance level was set at p -value ≤ 0.05 . Traits tested were all the zoometric

traits, the subjectively scored riding ability traits and the subjectively assessed sub-traits.

Genotyping of the top SNP and *DMRT3* in other gaited and partly gaited breeds

Horses of other gaited breeds (Rocky-Mountain: 36 horses, Colombian paso fino Curly: 38 horses) and partly gaited breeds (American Curly: 27 horses, American Saddlebred: 42 horses, Morgan: 30 horses and Standardbred: 40 horses) were genotyped for the top SNP using StepOnePlus Real-Time PCR System (Life Technologies) with a custom TaqMan SNP genotyping assay (Applied Biosystems). A group of 49 Icelandic horses used for riding but that had not attended breeding field test was also genotyped. The sequence of the primers and probes was designed as follows: forward primer: 5'-GGAAGTTTCTAAACATTTTGAAGGC TTTT-3'; reverse primer: GGAGGGGAAGTCAATTGAC AAACG; mutant probe (FAM): 5'-CCTCCACGGC ATCA-3'; reference probe (VIC): 5'-TCCCTCCACA GCATCA-3'. The reaction volume of 15 μ l contained: 1.5 μ l DNA, 0.38 μ l Genotyping Assay 40X, 7.50 μ l Genotyping Master Mix 2X, and 5.62 μ l deionized water. The thermal cycle included 95 °C for 10 min, 40 cycles of 95 °C for 15 s, and 60 °C for 1 min.

SNP genotyping of the *DMRT3_Ser301STOP* marker known as the "Gait Keeper" mutation was performed using custom designed TaqMan SNP Genotyping Assays (Applied Biosystem) as described previously [30, 34].

Functional annotation

The bioinformatics database NCBI was used to screen for candidate genes based on the EquCab3.0 reference

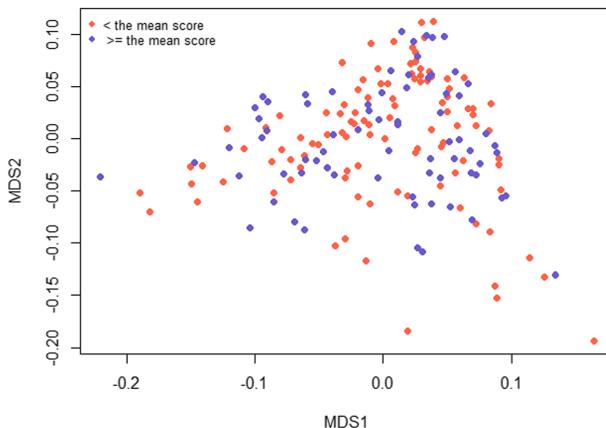


Fig. 5 MDS plot for the score back and croup. Visualization of population stratification across the 177 Icelandic horses that passed the QC for the score back and croup. Red represents horses that had a score lower than the mean 8.1 and blue represents horses that had a score higher or equal to 8.1

genome and annotation release 103 [75] and HorseQTLdb release 41 to search for known quantitative trait loci (QTLs) for conformation in horses [39]. Functional annotation of possible candidate genes was performed using the GeneCards database [76]. All positions refer to the EquCab3.0 reference genome.

Abbreviations

ADAMTS17: ADAM Metalloproteinase With Thrombospondin Type 1 Motif 17; AF: Allele frequency; AIS: Adolescent idiopathic scoliosis; ANKRD1: Ankyrin Repeat Domain 1; ANKRD60: Ankyrin repeat domain 60; BLAST: Basic Local Alignment Search Tool; C22H20orf85: Chromosome 22 C20orf85 homolog; CPH: Colombian paso horses; DMRT3: Doublesex And Mab-3 Related Transcription Factor 3; ECA: *Equus caballus* chromosome; GH1: Growth Hormone 1; glm: Generalized linear model; GWA: Genome-wide association; HMG2A: High Mobility Group AT-Hook 2; LASP1: LIM And SH3 Protein 1; LCORL: Ligand Dependent Nuclear Receptor Corepressor Like; LD: Linkage disequilibrium; MAF: Minor allele frequency; MDS: Multidimensional scaling; NCAPG: Non-SMC Condensin I Complex Subunit G; OSTN: Osteonin; PPP4R1L: Protein Phosphatase 4 Regulatory Subunit 1 Like; QC: Quality control; QTL: Quantitative trait loci; SD: Standard deviation; SIF: The Swedish Icelandic Horse Association; SLC22A8: Solute carrier family 22 member 8; SNP: Single nucleotide polymorphism; VR: The Swedish Research Council; ZFAT: Zinc Finger And AT-Hook Domain Containing

Supplementary Information

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Additional file 1.

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Authors' contributions

GL and SE initiated and designed the study. MKR, MS and JJJ collected the samples. MKR extracted the DNA. EA provided the phenotypic data from WorldFengur. MKR and HS performed the experiments and data analysis. MS, MKR and HS drafted the manuscript. AJ performed genotyping in other gaited breeds. RN and AJ performed the allele frequency analysis. MNB contributed with samples from CPH. SE and MS advised with the statistical analysis. MKR, HS, SE, MNB, EA, TK, MR, AV, BDV, JJJ, MS and GL contributed to the interpretation of the results. All authors read, suggested modifications and approved the final manuscript.

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Availability of data and materials

The datasets generated and analyzed during the current study are not publicly available since the study was performed in collaboration with the Icelandic horse breeding industry and has a commercial value for them. However, data is available from the corresponding author on reasonable request and with permission of the Icelandic horse association.

Declaration

Ethics approval and consent to participate

Hair samples were collected following owner's informed written consent and according to ethical approval by the Ethics Committee for Animal Experiments in Uppsala, Sweden (number: 5.8.18–15453/2017).

Consent for publication

Not applicable.

Competing interests

The authors declare competing interest concerning commercial applications of the current study. GL is a co-inventor on a patent application concerning commercial testing of the DMRT3 mutation. The stated patents does not restrict research applications of the method.

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References

1. Albertsdóttir E, Eriksson S, Näsholm A, Strandberg E, Árnason T. Genetic correlations between competition traits and traits scored at breeding field-tests in Icelandic horses. *Livest Sci.* 2008;114(2–3):181–7.
2. Solé M, Gómez MD, Galisteo AM, Santos R, Valera M. Kinematic characterization of the Menorca horse at the walk and the trot: influence of hind limb pastern angle. *J Equine Vet Sci.* 2013;33(9):726–32.
3. Kristjánsson T, Björnisdóttir S, Albertsdóttir E, Sigurdsson A, Pourcelot P, Crevier-Denoix N, et al. Association of conformation and riding ability in Icelandic horses. *Livest Sci.* 2016;189:91–101.
4. Holmström M, Back W. The effects of conformation. In: *Equine Locomotion*. 2nd ed. London: WB Saunders; 2013. p. 229–43.
5. Janczarek I, Wilk I, Strzelec K. Correlations between body dimensions of young trotters and motion parameters and racing performance. *Pferdeheilkunde.* 2017;33(2):139–45.
6. Sánchez-Guerrero MJ, Molina A, Gómez MD, Peña F, Valera M. Relationship between morphology and performance: signature of mass-selection in Pura Raza Español horse. *Livest Sci.* 2016;185:148–55.
7. Sánchez-Guerrero MJ, Cervantes I, Molina A, Gutiérrez JP, Valera M. Designing an early selection morphological linear traits index for dressage in the Pura Raza Español horse. *Animal.* 2017;11(6):948–57.
8. Jönsson L, Egenvall A, Roepstorff L, Näsholm A, Dalin G, Philipsson J. Associations of health status and conformation with longevity and lifetime competition performance in young Swedish Warmblood riding horses: 8,238 cases (1983–2005). *J Am Vet Med Assoc.* 2014 Jun 15;244(12):1449–61.
9. Jönsson L, Näsholm A, Roepstorff L, Egenvall A, Dalin G, Philipsson J. Conformation traits and their genetic and phenotypic associations with health status in young Swedish warmblood riding horses. *Livest Sci.* 2014 May 1;163(1):12–25.
10. Koenen EPC, Van Veldhuizen AE, Brascamp EW. Genetic parameters of linear scored conformation traits and their relation to dressage and show-jumping performance in the Dutch Warmblood Riding Horse population. *Livest Sci.* 1995;43:85–94.
11. Rustin M, Janssens S, Buys N, Gengler N. Multi-trait animal model estimation of genetic parameters for linear type and gait traits in the Belgian warmblood horse. *J Anim Breed Genet.* 2009;126(5):378–86.
12. FEIF. FEIF General Rules and Regulations. Available from: https://www.feiffengur.com/documents/FEIF_Rules_Regulations2020_complete.pdf [cited 2020 Aug 6].

13. Albertsdóttir E, Eriksson S, Sigurdsson Á, Árnason T. Genetic analysis of "breeding field test status" in Icelandic horses. *J Anim Breed Genet.* 2011; 128(2):124–32.
14. François L, Fegraeus KJ, Eriksson S, Andersson LS, Tesfayonas YG, Viluma A, et al. Conformation traits and gaits in the Icelandic horse are associated with genetic variants in *Myostatin* (*MSTN*). *J Hered.* 2016;107(5):431–7.
15. Makvandi-Nejad S, Hoffman GE, Allen JJ, Chu E, Gu E, Chandler AM, et al. Four loci explain 83% of size variation in the horse. *PLoS One.* 2012;7(7):1–6.
16. Weedon MN, Lango H, Lindgren CM, Wallace C, David M, Mangino M, et al. Genome-wide association analysis identifies 20 loci that influence adult height. *Nat Genet.* 2008;40(5):575–83.
17. Metzger J, Schrimpf R, Philipp U, Distl O. Expression levels of *LCORL* are associated with body size in horses. *PLoS One.* 2013;8(2):56497.
18. Frischknecht M, Jagannathan V, Plattet P, Neuditschko M, Signer-Hasler H, Bachmann I, et al. A non-synonymous *HMG2A* variant decreases height in shetland ponies and other small horses. *PLoS One.* 2015;10(10):1–11.
19. Hayward JJ, Castelthano MG, Oliveira KC, Corey E, Balkman C, Baxter TL, et al. Complex disease and phenotype mapping in the domestic dog. *Nat Commun.* 2016;7.
20. Bouwman AC, Daetwyler HD, Chamberlain AJ, Ponce CH, Sargolzaei M, Schenkel FS, et al. Meta-analysis of genome-wide association studies for cattle stature identifies common genes that regulate body size in mammals. *Nat Genet.* 2018;50(3):362–7.
21. Mostafavi A, Asadi Fozzi M, Esmailzadeh Koshkooieh A, Mohammadabadi M, Ivanivna Babenko O, Ihorivna Klopenko N. Effect of *LCORL* gene polymorphism on body size traits in horse populations. *Acta Sci Anim Sci.* 2020;42. <https://doi.org/10.4025/actascianimsci.v42i1.47483>.
22. Abri MA Al Holl HM, Kalla SE, Sutter NB, Brooks SA. Whole genome detection of sequence and structural polymorphism in six diverse horses. *PLoS One.* 2020;15(4). <https://doi.org/10.1371/journal.pone.0238099>.
23. Posbergh CJ, Huson HJ. All sheeps and zorses: a genetic investigation of mature body size across sheep breeds reveals a polygenic nature. *Anim Genet.* 2020;52:99–107.
24. Signer-Hasler H, Flury C, Haase B, Burger D, Simianer H, Leeb T, et al. A genome-wide association study reveals loci influencing height and other conformation traits in horses. *PLoS One.* 2012;7(5):3–8.
25. Tetens J, Widmann P, Kühn C, Thaller G. A genome-wide association study indicates *LCORL/NCAPG* as a candidate locus for withers height in German Warmblood horses. *Anim Genet.* 2013;44(4):467–71.
26. Metzger J, Rau J, Naccache F, Bas Conn L, Lindgren G, Distl O. Genome data uncover four synergistic key regulators for extremely small body size in horses. *BMC Genomics.* 2018;19(1):1–15.
27. Gmel AI, Druml T, von Niederhäusern R, Leeb T, Neuditschko M. Genome-wide association studies based on equine joint angle measurements reveal new QTL affecting the conformation of horses. *Genes (Basel).* 2019;10(5):370.
28. Novoa-Bravo M, Jäderkvist Fegraeus K, Rhodin M, Strand E, García LF, Lindgren G. Selection on the Colombian paso horse's gaits has produced kinematic differences partly explained by the *DMRT3* gene. *PLoS One.* 2018; 13(8):e0202584.
29. Jäderkvist K, Kangas N, Andersson LS, Lindgren G. Gaitedness is associated with the *DMRT3* 'gait keeper' mutation in Morgan and American curly horses. *Anim Genet.* 2014;45(6):908–9.
30. Promerová M, Andersson LS, Juras R, Penedo MCT, Reissmann M, Tozaki T, et al. Worldwide frequency distribution of the 'Gait keeper' mutation in the *DMRT3* gene. *Anim Genet.* 2014;45(2):274–82.
31. Vellie BD, Shrestha M, François L, Schurink A, Tesfayonas YG, Stinckens A, et al. Using an Inbred Horse Breed in a High-Density Genome-Wide Scan for Genetic Risk Factors of Insect Bite Hypersensitivity (IBH). *PLoS One.* 2016; 11(4):e0152966 Barendse W, editor.
32. Finno CJ, Stevens C, Young A, Affolter V, Joshi NA, Ramsay S, et al. SERP INB11 Frameshift Variant Associated with Novel Hoof Specific Phenotype in Connemara Ponies. *PLoS Genet.* 2015;11(4):e1005122 Barsh GS, editor.
33. Ablondi M, Viklund Å, Lindgren G, Eriksson S, Mikko S. Signatures of selection in the genome of Swedish warmblood horses selected for sport performance. *BMC Genomics.* 2019;20(1):717.
34. Andersson LS, Larhammar M, Memic F, Wootz H, Schwachow D, Rubin CJ, et al. Mutations in *DMRT3* affect locomotion in horses and spinal circuit function in mice. *Nature.* 2012;488(7413):642–6.
35. Fawcett JA, Sato F, Sakamoto T, Iwasaki WM, Tozaki T, Innan H. Genome-wide SNP analysis of Japanese Thoroughbred racehorses. *PLoS One.* 2019; 14(7). <https://doi.org/10.1371/journal.pone.0218407>.
36. Sadeghi R, Moradi-Shahrbabak M, Reza Miraei Ashtiani S, Schlamp F, Cosgrove EJ, Antczak DF, et al. Genetic Diversity of Persian Arabian Horses and Their Relationship to Other Native Iranian Horse Breeds. *J Hered.* 2019; 110:173–82.
37. Vellie BD, Lillie M, Fegraeus KJ, Rosengren MK, Solé M, Wiklund M, et al. Exploring the genetics of trotting racing ability in horses using a unique Nordic horse model. *BMC Genomics.* 2019;20(1):104.
38. Vellie BD, Fegraeus KJ, Solé M, Rosengren MK, Roed KH, Ihler C-F, et al. A genome-wide association study for harness racing success in the Norwegian-Swedish coldblooded trotter reveals genes for learning and energy metabolism. *BMC Genet.* 2018;19(1). <https://doi.org/10.1186/s12863-018-0670-3>.
39. Horse QTL Database. Available from: <https://www.animalgenome.org/cgi-bin/QTLdb/EC/index>. [cited 2020 Aug 6]
40. Jönsson L. Orthopaedic health, conformation and longevity in riding horses. Doctoral thesis. SLU; 2013.
41. Liu J, Zhou Y, Liu S, Song X, Yang XZ, Fan Y, et al. The coexistence of copy number variations (CNVs) and single nucleotide polymorphisms (SNPs) at a locus can result in distorted calculations of the significance in associating SNPs to disease. *Hum Genet.* 2018;137(6–7):553–67.
42. Kim JJ, Lee HI, Park T, Kim K, Lee JE, Cho NH, et al. Identification of 15 loci influencing height in a Korean population. *J Hum Genet.* 2010 Jan 6;55(1): 27–31.
43. Konieczny MR, Senyurt H, Krauspe R. Epidemiology of adolescent idiopathic scoliosis. *J Child Orthop.* 2013;7:3–9 Springer.
44. Martínez-Llorens J, Ramírez M, Colomina M, Bagó J, Molina A, Cáceres E, et al. Muscle dysfunction and exercise limitation in adolescent idiopathic scoliosis. *Eur Respir J.* 2010;26:393–400.
45. Milenkovic SM, Kocjancic RI, Belojevic GA. Left handedness and spine deformities in early adolescence. *Eur J Epidemiol.* 2004;19:969–72.
46. Grivas T, Mihas K, Vasiladias E, Maziotou C, Karathanou S, Polyzois V. Handedness and laterality of trunk Rotation in children screened for scoliosis. Proceedings of the International Research Society of Spinal Deformities conference; 2004 June 10–12; Biennial Meeting. 2004:64–7.
47. de Yang Z, Li M. There may be a same mechanism of the left-right handedness and left-right convex curve pattern of adolescent idiopathic scoliosis. *Med Hypotheses.* 2011;76(2):274–6.
48. Catanzariti JF, Guyot MA, Agnani O, Demaille S, Kolanowski E, Donze C. Eye-hand laterality and right thoracic idiopathic scoliosis. *Eur Spine J.* 2014;23(6): 1232–6.
49. Grivas TB, Vasiladias ES, Polyzois VD, Mouzakis V. Trunk asymmetry and handedness in 8245 school children. *Pediatr Rehabil.* 2006;9(3):259–66.
50. Ocklenburg S, Schmitz J, Moinfar Z, Moser D, Klose R, Lor S, et al. Epigenetic regulation of lateralized fetal spinal gene expression underlies hemispheric asymmetries. *Elife.* 2017;6. <https://doi.org/10.7554/eLife.22784>.
51. Jeffcott LB. Disorders of the thoracolumbar spine of the horse — a survey of 443 cases. *Equine Vet J.* 1980;12(4):197–210.
52. Wong D, Miles K, Sponseller B. Congenital Scoliosis In A Quarter Horse Filly. *Vet Radiol Ultrasound.* 2006;47(3):279–82.
53. Lucidi P, Bacco G, Sticco M, Mazzoleni G, Benvenuti M, Bernabò N, et al. Assessment of motor laterality in foals and young horses (*Equus caballus*) through an analysis of derailment at trot. *Physiol Behav.* 2013;109(1):8–13.
54. McGreevy PD, Thomson PC. Differences in motor laterality between breeds of performance horse. *Appl Anim Behav Sci.* 2006;99(1–2):183–90.
55. Cully P, Nielsen B, Lancaster B, Martin J, McGreevy P. The laterality of the gallop gait in Thoroughbred racehorses. *PLoS One.* 2018;13(6):e0198545 Bartos L, editor.
56. Regatieri IC, Eberth JE, Sarver F, Lear TL, Bailey E. Comparison of *DMRT3* genotypes among American Saddlebred horses with reference to gait. *Anim Genet.* 2016 Oct 1;47(5):603–5.
57. Nicodemus MCC, Clayton HM. Temporal variables of four-beat, stepping gaits of gaited horses. *Appl Anim Behav Sci.* 2003;80(2):133–42.
58. Al Abri MA, Posbergh C, Palermo K, Sutter NB, Eberth J, Hoffman GE, et al. Genome-Wide Scans Reveal a Quantitative Trait Locus for Withers Height in Horses Near the *ANKRD1* Gene. *J Equine Vet Sci.* 2018;60:67–73 e1.
59. BLAST: Basic Local Alignment Search Tool. Available from: <https://blast.ncbi.nlm.nih.gov/Blast.cgi>. [cited 2020 Aug 7]
60. Gaudet P, Livstone MS, Lewis SE, Thomas PD. Phylogenetic-based propagation of functional annotations within the gene ontology consortium. *Brief Bioinform.* 2011;12(5):449–62.
61. Tachmazidou I, Süveges D, Min JL, Ritchie GRS, Steinberg J, Walter K, et al. Whole-genome sequencing coupled to imputation discovers genetic signals for anthropometric traits. *Am J Hum Genet.* 2017;100(6):865–84.

62. Hnisz D, Abraham BJ, Lee TI, Lau A, Saint-André V, Sigova AA, et al. Super-enhancers in the control of cell identity and disease. *Cell*. 2013;155(4):934.
63. Hoyt DF, Wickler SJ, Cogger EA. Time of contact and step length: the effect of limb length, running speed, load carrying and incline. *J Exp Biol*. 2000; 203(2):221–7.
64. Sparrow LM, Pellatt E, Yu SS, Raichlen DA, Pontzer H, Rolian C. Gait changes in a line of mice artificially selected for longer limbs. *PeerJ*. 2017;2017(2): e3008.
65. Pontzer H. Effective limb length and the scaling of locomotor cost in terrestrial animals. *J Exp Biol*. 2007;210(10):1752–61.
66. Cook D, Gallagher PC, Bailey E. Genetics of swayback in American Saddlebred horses. *Anim Genet*. 2010;41(SUPPL. 2):64–71.
67. WorldFengur The Studbook of Origin for the Icelandic horse. Available from: <https://www.worldfengur.com/>. [cited 2020 Aug 6]
68. The Icelandic Agricultural Advisory Centre. General Information on Breeding Field Tests – Measurements (body measures). Available from: https://www.mll.is/static/files/Hrossaraekt_RML/skrokkmal.pdf. [cited 2020 Aug 6]
69. Aulchenko YS, Ripke S, Isaacs A, van Duijn CM. GenABEL: an R library for genome-wide association analysis. *Bioinformatics*. 2007;23(10):1294–6.
70. R Core Team. R: a language and environment for statistical computing. Vienna: Foundation for Statistical Computing; 2015.
71. Rönnegård L, Shen X, Alam M. Hglm: a package for fitting hierarchical generalized linear models. *R J*. 2010;2(2):20–8.
72. Lander E, Kruglyak L. Genetic dissection of complex traits : reporting linkage results. *Nat Genet*. 1995;11:241–7.
73. Duggal P, Gillanders EM, Holmes TN, Bailey-Wilson JE. Establishing an adjusted p-value threshold to control the family-wide type 1 error in genome wide association studies. *BMC Genomics*. 2008;9:1–8.
74. Kierczak M, Jabłońska J, Forsberg SKG, Bianchi M, Tengvall K, et al. cgmisc: enhanced genome-wide association analyses and visualization. *Bioinformatics*. 2015;31:3830–1.
75. National Center for Biotechnology Information. Available from: <https://www.ncbi.nlm.nih.gov/>. [cited 2020 Aug 6]
76. Stelzer G, Rosen R, Plaschkes I, Zimmerman S, Twik M, Fishilevich S, et al. The GeneCards suite: from gene data mining to disease genome sequence analyses. *Current Protocols in Bioinformatics*. 2016;54:1.30.1–1.30.33. Available from: <https://www.genecards.org/>. [cited 2020 Aug 6]

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The overall aims of the thesis were to identify genetic variants associated with athletic performance and conformation traits in horses, as well as to investigate the level of inbreeding in Coldblooded trotters. The results contribute to the understanding of the genetics of complex traits such as athletic performance, blood pressure regulation, inflammation, and anthropometric traits in horses. This information may also be valuable for humans and other mammalian species.

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