



Article Can Larix sp. Mill. Provide Suitable Habitats for Insects and Lichens Associated with Stems of *Picea abies* (L.) H. Karst. in Northern Europe?

Jūratė Lynikienė^{1,*}, Artūras Gedminas¹, Adas Marčiulynas¹, Diana Marčiulynienė¹ and Audrius Menkis²

- ¹ Institute of Forestry, Lithuanian Research Centre for Agriculture and Forestry, Liepu ştr. 1, Kaunas District, LT-53101 Girionys, Lithuania
- ² Department of Forest Mycology and Plant Pathology, Uppsala BioCenter, Swedish University of Agricultural Sciences, P.O. Box 7026, SE-75007 Uppsala, Sweden
- Correspondence: jurate.lynikiene@lammc.lt

Abstract: Recent observations suggest that climate change affects the growth conditions and range of tree species distribution in Europe. This may also have a major effect on communities of different organisms associated with these tree species. We aimed to determine whether *Larix* sp. could provide suitable habitats to insects and lichens associated with *P. abies* to conserve their biodiversity under climate change. The study sites were 10 *Larix* sp. and 10 *P. abies* forest stands in Lithuania. Both living and dead trees were included. Sticky traps, bark sheets, and exit hole methods were used for the assessment of insects. Independent plots on tree stems were established for the assessment of lichens. There were 76 and 67 different insect species on dead and living *P. abies*, respectively, using sticky traps. Similarly, there were 64 and 68 on dead and living *Larix* sp., respectively. The overall community of xylophagous insects consisted of nine and eight species, which were detected using the bark sheet and exit hole methods, respectively. The bark area colonized by lichens was 34.3% on dead *P. abies* and 63.2% on dead *Larix* sp., and 40.4% on living *P. abies* and 78.0% on living *Larix* sp. Taken together, the results demonstrate that native *P. abies* and introduced *Larix* sp. support similar diversity of stem-associated insect and lichen species.

Keywords: biodiversity; climate change; Norway spruce; larch; insects; lichens; forest management

1. Introduction

The ongoing process of climate change can be expected to have profound consequences for European forests, especially if species-specific climatic thresholds are surpassed. Prominent climatic changes, which are primarily affecting tree productivity, are mainly associated with increased droughts [1]. Droughts, especially in combination with different biotic factors, such as attacks by pests and pathogens, are known to make trees weaker or even cause mortality [1]. Consequently, the distributional range of different tree species and the composition of European forests can be expected to change in the future [2–4]. In north temperate and boreal European forests, the most economically, ecologically, and spatially important and abundant tree species are Scots pine (*Pinus sylvestris*, L.) and Norway spruce (*Picea abies*, (L.) H. Karst.), which are regionally experiencing increased mortality rates [2,5–12].

Picea abies is one of the most canonical tree species in the forest ecosystems of Eurasia. The area of its natural distribution is vast and ranges from western Siberia to Fennoscandia and the mountain ranges of central Europe [13]. It grows under a wide range of climatic conditions and tolerates a cool and wet climate. It predominantly grows on fertile soils and is a relatively shade-tolerant tree species, forming pure or mixed forest stands with different tree species [14,15]. As it produces valuable timber and its stands are relatively easy to manage, *P. abies* has been extensively planted both within and outside the natural range of



Citation: Lynikienė, J.; Gedminas, A.; Marčiulynas, A.; Marčiulynienė, D.; Menkis, A. Can *Larix* sp. Mill. Provide Suitable Habitats for Insects and Lichens Associated with Stems of *Picea abies* (L.) H. Karst. in Northern Europe? *Diversity* 2022, *14*, 729. https://doi.org/10.3390/d14090729

Academic Editors: Paolo Solari, Giorgia Sollai, Roberto Massimo Crnjar, Anita Giglio and Piero G. Giulianini

Received: 12 July 2022 Accepted: 2 September 2022 Published: 4 September 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). distribution, resulting in a considerable increase in its stands during the last century [14]. However, observations suggest that climate change is one of the most important factors leading to growth disturbances of *P. abies* throughout its distribution range [15–17]. A relatively shallow root system makes the tree species prone to both drought stress [8,18] and wind damage [19–21]. In addition, in the past decades, *P. abies* has been increasingly damaged over vast areas by the European spruce bark beetle (*Ips typographus* L., Coleoptera: Curculionidae). Outbreaks of *I. typographus* are frequently triggered by major storms and/or severe droughts [13,22–24]. Such disturbances can be expected to increase in the future, particularly at the edge of the current distribution range of *P. abies*, as the effects of climate change are likely to be most pronounced in these areas [25]. Consequently, the observed and predicted vulnerability of *P. abies* to abiotic and biotic damage requires special attention [13,26,27]. Indeed, different alternatives and solutions on how to mitigate the negative effects of climate change should be carefully considered [28].

Several studies provide valuable insights into the cultivation of some introduced coniferous tree species as an alternative to *P. abies*. In Western and Northern Europe, several exotic tree species within Pinaceae, namely, Douglas fir (*Pseudotsuga menziesii* (Mirb.) Franco), Lodgepole pine (*Pinus contorta* Doug. ex. Loud. var. latifolia Engelm.), Monterey pine (*Pinus radiata* D. Don.), and Sitka spruce (*Picea sitchensis* (Bong.) Carrière), presently constitute important portions of forested ecosystems [29,30]. Larch (*Larix* sp.) trees have also been considered as an alternative to *P. abies*, as it grows successfully in similar habitats to *P. abies* [31]. In addition, *Larix* sp. grows naturally in many areas of the northern hemisphere, being one of the components in boreal and mountain subalpine forests.

In Lithuania, *P. abies* constitutes ca. 21% of the forest area and is of great economic and ecological importance. However, as this area is close to the south-eastern edge of the natural distribution of *P. abies* in Europe, it is increasingly subjected to abiotic and biotic damage. In Lithuania, Larix sp. was introduced early in the 19th century as an exotic tree species. Nowadays, it is planted in monocultures or in mixed forest stands [32] but occupies only ca. 900 ha of forest area [33]. In comparison to *P. abies, Larix* generally possesses faster growth, more durable wood, and higher adaptability to different environmental conditions, which is partly due to the deep root system, making trees more resistant to windthrows and droughts [34]. Larix is an early successional tree species, and after disturbances such as large-scale windfall, it is able to establish on *Picea* sites [35]. *Larix* is a much more lightdemanding tree species as compared to *P. abies*. Additionally, it requires large openings for regeneration and juvenile growth, while *P. abie*s can regenerate in much smaller gaps or under the canopy [36]. Forest managers have attempted to cultivate several different Larix species in forest stands (L. leptolepis, L. decidua, and L. polonica), but L. decidua ssp. Polonica Ostenf et. Syrach shows the best growth rate (apart from its hybrids). Moreover, its productivity is significantly higher as compared to other coniferous tree species [37]. However, the productivity of *Larix* sp. can be reduced by insect pests, some of which are also able to damage *P. abies* or *P. sylvestris* [38].

The large-scale planting of introduced tree species instead of native species requires an evidence-based evaluation [30]. In addition to the productivity and adaptability or resilience to climate change, other factors, such as impacts on native flora and fauna communities, should be considered. Indigenous tree species are commonly associated with and/or provide habitats for a variety of different organisms, such as fungi, lichens, or insects. These organisms can be associated with their hosts [39]. Changes in the native forest structure and composition could lead to disturbances in the diversity and composition of these organisms and thus may affect the functioning of forest ecosystems.

Insects represent a key component in forest ecosystems [39], as they are involved in food web interactions (as herbivores, saprophages, predators, and parasites), ecosystem processes (such as pollination, energy flow, biogeochemical cycling, or ecological succession), and eco-evolutionary processes [40,41]. Epiphytic lichens are also an important component of forest biodiversity associated with coniferous forests in Europe [42]. Several studies have highlighted the importance of lichen diversity as an indicator of environmental change,

which is based on their response to air pollution [43,44], climatic conditions [45–47], and forest structure and dynamics [48–50]. The specific association between certain epiphytic lichens and host trees was demonstrated by Roper [51], and this is probably due to differences in the structure and acidity of the bark, thereby leading to sharp differences in lichen cover and diversity between different tree species. Although *Larix* sp. is often considered as an alternative tree species to *P. abies* for the future, the comparative analysis of insect and epiphytic lichen diversity on the stems of these tree species is generally lacking.

The aim of the present study was to determine whether *Larix* sp. could provide suitable habitats to insects and lichens associated with *P. abies* to conserve their biodiversity under climate change.

2. Materials and Methods

2.1. Study Site and Observation

The study sites were in *P. abies* and *Larix* sp. forest stands at 10 different locations in Lithuania (Figure 1 and Table 1).

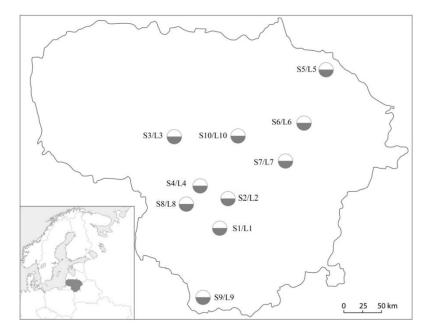


Figure 1. Map of Lithuania showing the distribution of study sites. Gray color in the circle indicates *Larix* sp. stands (L1–L10), and white color represents *P. abies* stands (S1–S10).

The identification of larch species is problematic due to their frequent hybridization [52,53], so in this study, they are referred to as *Larix* sp. At each site, there was one *P. abies* and one *Larix* sp. stand, which were within a radius of 200 m, so they were within the same geographical area and exposed to similar climatic conditions. The topography was similar in these areas. Information on the stand and site characteristics is in Table 1.

Study sites were selected based on forest inventory data from the State Forest Cadastre database. The criteria used for the selection of each study site were: (i) *P. abies* or *Larix* sp. trees were the prevailing species at the site; (ii) similar soil type [54]; and (iii) similar vegetation type [55]. Most of the study sites were characterized by soils of moderate fertility and normal humidity and by an *oxalidosa* vegetation type (Table 1).

At each *P. abies* or *Larix* sp. study site, up to five healthy-looking and up to five dead trees (dead trees were not always available) were randomly selected and used for the assessment of insects and lichens, which was carried out in 2018 and 2019.

Site *	Geographical Position	Age (y)	Mean Height (m)	Mean Diameter (cm)	Stocking Level	Forest Site Type **	Forest Vegetation Type ***	Tree Species Composition (%) ****
S1	54°33′18.88″ N, 23°53′14.53″ E	47	21.7	23.5	1.3	Ncs	ox	1005
L1	54°33′19.82″ N, 23°53′17.18″ E	47	28.1	34.3	0.9	Ncs	ox	100L
S2	54°51′36.84″ N, 24°4′25.17″ E	57	25.2	29.0	0.9	Ncp	ox	40S 20L 20Q 10T 10B
L2	54°51′37.18″ N, 24°4′29.02″ E	37	28.5	33.7	0.8	Ncp	ox	90L10T
S3	55°17′10.6″ N, 23°26′11.7″ E	55	23.6	26.0	0.9	Lds	hox	1005
L3	55°17′10.56″ N, 23°26′23.63″ E	50	29.1	43.5	0.7	Ldp	aeg	100L
S4	55°3′19.44″ N, 23°31′8.07″ E	67	24.6	26.2	0.8	Ncp	ox	80S 10P 10S
L4	55°3′18.74″ N, 23°31′4.2 ″ E	72	35.9	42.8	0.8	Ncl	ox	90L 10P
S5	55°55′53.91″ N, 25°36′33.16″ E	35	19.0	24.0	0.6	Ldp	oxn	805 20Q
L5	55°57′51.93″ N, 25°37′7.89″ E	80	28.0	34.0	0.6	Ldp	aeg	70L 20Pt 10B
S6	55°30′46.23″ N, 25°5′33.21″ E	50	19.0	18.0	0.9	Ncl	ox	50S 30P 20T
L6	55°30′46.9″ N, 25°5′35.92″ E	55	25.0	24.0	0.9	Lcl	ox	50P 30L 20S
S7	55°15′4.99″N, 24°48′58.27″ E	38	17.5	19.4	0.6	Ncl	ox	90S 10P
L7	55°15′53.53″ N, 24°48′50.76″ E	38	24.7	29.4	0.9	Ncl	ox	100 L
S8	54°48′57.86″ N, 23°25′24.43″ E	66	25.5	27.9	1.0	Nbl	m	80S 20P
L8	54°49′24.5″ N, 23°25′29.83″ E	66	32.7	32.2	0.8	Ncl	ox	80L 20P
S9	54°0′24.82″ N, 23°44′31.7″ E	84	25.4	28.0	0.7	Nbl	v	60S 10P 10S 20S
L9	54°0′20.68″ N, 23°38′7.07″ E	59	32.2	38.6	0.6	Ncl	ox	100L
S10	55°23'12.67" N, 24°7'10.42" E	58	24.4	20.5	1.2	Nds	hox	90S 10B
L10	55°23'14.38" N, 24°7'13.74" E	58	26.4	29.2	0.7	Nds	hox	90L 10B

Table 1. Characteristics of investigated *Picea abies* and *Larix* sp. stands. Information is based on forest inventory data obtained from the State Forest Cadastre as of 1 December 2020.

* S1–S10: *Picea abies* stands; L1–L10: *Larix* sp. stands as in Figure 1. ** N: Normal humidity; L: temporarily waterlogged mineral soils; b: low fertility; c: moderate fertility; d: high fertility; l: light soil texture; p: two-layered soil structure with a light fraction on a heavy fraction or vice versa; s: heavy soil texture [54]. *** v: *vacciniosa;* m: *myrtilliosa;* ox: *oxalidosa;* hox: *hepatico-oxalidosa;* oxn: *oxalido-nemorosa;* aeg: *aegopodiosa* [55]. **** S: *Picea abies;* L: *Larix* sp.; P: *Pinus sylvestris;* Q: *Quercus robur;* B: *Betula pendula;* T: *Tilia cordata;* Pt: *Populus tremula.* In each stand, tree species composition is based on the volume.

2.2. Assessment of Insects Associated with Tree Stems

Three different methods were used for the assessment of insects: (i) using sticky traps, which were used to capture insects occurring on the surfaces of living and dead *P. abies* and *Larix* sp. trees, (ii) recording signs of xylophagous insects under the bark of dead trees, and (iii) recording exit holes of xylophagous insects on the bark of dead trees. For the capture of insects on the surface of tree stems [56], two sticky traps, which were made of 20×20 cm polyethylene sheets treated with non-drying glue (Pestifix, "Flora", Talinn, Estonia), were attached to each of the five living and five dead *P. abies* standing tree stems. Both traps were placed at the same height of ca. 1.5 m above the ground to prevent interference from grasses and shrubs. Sticky traps on living and dead *Larix* sp. tree stems were established in the same way. The assessment of insects using sticky traps with trapped insects were collected once a month and replaced with new ones, which resulted

in three time points (June, July, and August). Collected sticky traps were transported to the laboratory the same day and stored at 5 $^{\circ}$ C until the identification of insect species using a binocular Zeiss Stemi 2000-C microscope (Oberkochen, Germany) and morphological insect identification keys [57–62]. Many insects were identified to the species level, while others were identified to the order, family, or genus level. Several insects remained unidentified, which was largely because they were missing body parts or were heavily covered by glue from sticky traps, thereby making reliable identification impossible. After the identification of insect species, accidentally trapped insects, i.e., species specifically associated with the tree crowns or non-target species, the development and feeding of which are not dependent on tree stems, were excluded from further analyses. However, predators and parasites of insects associated with tree stems were included in analyses.

Xylophagous insects, which are wood- and bark-boring insects, were assessed on dead *P. abies* and *Larix* sp. trees, and the signs and areas of their activity under the bark, i.e., larval tunnels, pupal chambers, and adult holes in the wood, were recorded. This was carried out once in August 2019 by removing a 20×20 cm bark sheet at a height of ca. 1.5 m above the ground [56]. In each study site, one bark sheet was removed from each of the five dead trees of each tree species, resulting in a total of 0.4 m² bark area in each site. The area of removed bark was photographed, signs of insect activity were analyzed in the laboratory, and insect species were identified.

For the assessment of exit holes of adult insects and the identification of their species, five dead trees of *P. abies* and *Larix* sp. per study site were visually inspected in August 2018. On each tree stem (1–1.5 m above the ground), insect exit holes were recorded on five plots, each 0.01 m² in size, and were situated along the stem and from four different geographical directions (N, S, E, and W), resulting in a 0.2 m² area per tree in total. This method was adopted from Asta et al. [63]. The number of exit holes was recorded for each insect species separately. Insect species for which exit holes were clearly species-specific, e.g., *Ips typograhus* and *Pityogenes chalcographus*, were identified to the species level, while others were identified to the family or genus level.

2.3. Assessment of Epiphytic Lichens

Epiphytic lichens were assessed at the same study sites in August 2018 (Figure 1 and Table 1). In each *P. abies* or *Larix* sp. study site, five healthy and five dead trees were selected. The selected trees were ca. 18–20 cm in diameter at a height of 1.3 m above the ground, stem inclination was not more than 20° , trees were without wounds on the stem, and bark structure and bark thickness were similar for all trees of each tree species. For the assessment of lichens, on each tree, four independent plots, each 10×10 cm in size and each facing a different geographical direction (N, S, E, or W), were established at ca. 1.5 m above the ground [63]. All lichen species present within each plot and the area covered by each of them were recorded. Most of the lichen specimens were identified to the species, genus, or family level, but several species remained unidentified.

2.4. Statistical Analysis

Differences in the richness of insect or lichen taxa between dead or living trees of *P. abies* and *Larix* sp. were compared by nonparametric chi-square test [64], taking into account the Bonferroni correction. The Shannon diversity index, qualitative Sorensen similarity index, and nonmetric multidimensional scaling (NMDS) in Canoco 5 [65–67] were used to characterize the diversity and composition of insect and lichen communities. The nonparametric Mann–Whitney test in Minitab v.19.2 (Minitab[®] Inc., Pennsylvania State University, State College, PA, USA) was used to test if the Shannon diversity index among different samples differed significantly or not. ANOVA in Minitab was used to evaluate whether the bark area colonized by lichens differed among different tree species.

3. Results

3.1. Insects

In total, there were 20,226 insects trapped using sticky traps (Table 2). When all sites were taken together, on dead *P. abies*, there were 76 different insect species identified among 10,858 (53.3%) insects trapped, while on dead *Larix* sp., there were 64 different species identified among 1017 (5%) insects trapped.

Table 2. Diversity of insects detected in sticky traps at different *Picea abies* and *Larix* sp. study sites.

Site	Tree State	Tree Species	Relative Abundance, % (No. of Insects)	Richness, % (No. of Insect Species)	Shannon H	Sørensen Cs *
	Live	Picea Larix	0.4 (74) 0.3 (69)	22.1 (21) 15.8 (15)	2.34 2.16	0.44
S1/L1	Dead	Picea Larix	0.3 (63)	24.2 (23)	2.53	-
	Т	otal	1.0 (206)	40.0 (38)	2.35	0.48
	Live	Picea Larix	0.5 (109) 0.2 (32)	25.3 (24) 15.8 (15)	2.46 2.33	0.36
S2/L2	Dead	Picea Larix	29.1 (5885) 0.1 (16)	25.3 (24) 9.5 (9)	0.07 1.85	0.42
	Т	otal	29.9 (6042)	43.2 (41)	0.22	0.50
	Live	Picea Larix	0.4 (82) 1.1 (221)	18.9 (18) 18.9 (18)	2.14 1.86	0.39
S3/L3	Dead	Picea Larix	0.71 (144) 1.6 (325)	27.4 (26) 25.3 (24)	2.32 1.63	0.40
	Т	otal	3.8 (772)	48.4 (46)	2.24	0.54
	Live	Picea Larix	0.5 (102) 0.4 (91)	29.5 (28) 21.1 (20)	2.81 2.16	0.46
S4/L4	Dead	Picea Larix	4.5 (918) 0.3 (64)	34.7 (33) 24.2 (23)	0.86 2.52	0.57
	Т	otal	5.8 (1175)	56.8 (54)	1.68	0.60
	Live	Picea Larix	0.6 (127) 0.5 (99)	18.9 (18) 22.1 (21)	2.06 2.58	0.46
S5/L5	Dead	Picea Larix	1.4 (286)	28.4 (27)	2.21	-
	т	Total	2.5 (512)	44.2 (42)	2.66	0.41
	Live	Picea Larix	0.6 (120) 0.4 (78)	23.2 (22) 17.9 (17)	1.76 1.94	0.41
S6/L6	Dead	Picea Larix	0.9 (194) 0.7 (146)	29.5 (28) 25.3 (24)	2.48 2.01	0.46
	T	otal	2.7 (538)	53.7 (51)	2.49	0.56
	Live	Picea Larix	31.5 (6369) 0.6 (122)	30.5 (29) 25.3 (24)	0.25 2.22	0.49
S7/L7	Dead	Picea Larix	1.2 (241) 0.8 (165)	30.5 (29) 29.5 (28)	2.32 2.24	0.46
	T	otal	34.1 (6897)	62.1 (59)	0.64	0.54

Site	Tree State	Tree Species	Relative Abundance, % (No. of Insects)	Richness, % (No. of Insect Species)	Shannon H	Sørensen Cs *
	Live	Picea Larix	0.5 (110) 0.3 (54)	28.4 (27) 17.9 (17)	2.57 2.25	0.59
S8/L8	Dead	Picea Larix	8.4 (1705) 0.4 (82)	36.8 (35) 20.0 (19)	1.05 2.49	0.56
-	Т	otal	9.6 (1951)	51.6 (49)	1.53	0.60
	Live	Picea Larix	1.0 (207) 0.5 (95)	25.3 (24) 25.3 (24)	2.10 2.38	0.58
S9/L9	Dead	Picea Larix	6.0 (1222) 0.6 (117)	35.8 (34) 23.2 (22)	0.99 2.39	0.39
-	Т	otal	8.1 (1641)	55.8 (53)	1.74	0.52
	Live	Picea Larix	0.6 (120) 0.3 (70)	28.4 (27) 17.9 (17)	2.71 2.00	0.55
S10/L10	Dead	Picea Larix	1.0 (200) 0.5 (102)	33.7 (32) 25.3 (24)	2.68 2.68	0.57
-	Т	otal	2.4 (492)	53.7 (51)	2.84	0.57
All sites	Live Dea	e Picea e Larix d Picea d Larix	36.7 (7420) 4.6 (931) 53.7 (10,858) 5.0 (1017)	70.5 (67) 71.6 (68) 80 (76) 67.4 (64)		
	All total		100 (20,226)	100 (95)		

Table 2. Cont.

* Sørensen similarity index in rows Total shows the comparison between all *Picea abies* and all *Larix* sp. trees within adjacent study sites, e.g., S2 and L2.

Consequently, the chi-square test showed that the richness of insect species was significantly higher on dead *Larix* sp. than on dead *P. abies* (p < 0.0001). Similarly, on living *P. abies*, there were 67 different insect species among 7420 (36.7%) insects trapped, while on living *Larix* sp., there were 68 different species among 931 (4.6%) insects trapped. The richness of insect species was significantly higher on living *Larix* sp. than on living *P. abies* (p < 0.0001).

Many insect species were shared between dead trees and between living trees of both tree species. Among the 95 insect species identified, 5 were unique to living *P. abies*, and 4 were unique to living *Larix* sp. trees. Similarly, two insect species were unique to dead *P. abies*, and five were unique to dead *Larix* sp. (Figure 2).

Consequently, the Sørensen similarity index of insect communities was moderate when compared between dead trees of both tree species and living trees of both tree species (Table 2). The Mann–Whitney test showed that the Shannon diversity index of insect communities was similar between dead trees (p > 0.05) and between living trees (p > 0.05) when compared between *P. abies* and *Larix* sp., respectively. NMDS showed that insect communities on living *P. abies* and living *Larix* sp. were partially overlapping (Figure 3a). By contrast, insect communities on dead *P. abies* and dead *Larix* sp. were separated along the diagonal (Figure 3a). However, NMDS showed that there was a partial overlap between insect communities on living *P. abies* and dead *Larix* sp. (Figure 3a). Assessments that were conducted in June, July, and August showed that there were only minor variations in the abundance of dominant insect species on both living and dead *P. abies* and *Larix* sp. (Table 3). The most common insect species on *P. abies* were *Crypturgus pusillus, Ichneomonidae* sp., and *Eucnemidae* sp. 2 (Table 3). All insect species detected using sticky traps are in Table S1.

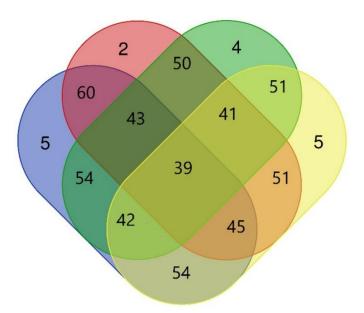


Figure 2. Venn diagram showing the species richness and overlap of insect species collected using sticky traps. The data from different study sites are combined. Different colors show: blue—living *Picea abies*; pink—dead *P. abies*; green—living *Larix* sp.; and yellow—dead *Larix* sp.

The overall community of xylophagous insects consisted of nine species detected using bark sheets and eight species detected using the exit hole method (Table 4). When all sites were taken together, the colonized bark area was 39.0% on dead *P. abies* and 47.3% on *Larix* sp. trees. Consequently, the chi-square test showed that the bark area colonized was significantly higher on dead *Larix* sp. trees than on *P. abies* (p < 0.0001). The number of exit holes of xylophagous insects was 2509 (75.4%) on dead *P. abies* and 819 (24.6%) on *Larix* sp. The number of exit holes was significantly higher on *P. abies* than on *Larix* sp. trees (p < 0.0001) (Table 4). More importantly, communities of xylophagous insects detected using bark sheet and exit hole methods were similar when compared between *P. abies* and *Larix* sp. (Figure 3b,c). In support, the Sørensen similarity index was 0.80, showing high species similarity between all dead *P. abies* and *Larix* sp. trees using both methods (Table 4). The Mann–Whitney test showed that the Shannon diversity index of xylophagous insect communities was similar using bark sheet (p > 0.05) and exit hole methods (p > 0.05) when a comparison was made between dead *P. abies* and *Larix* sp., respectively.

However, the species composition of xylophagous insects was quite different when compared between bark sheet and exit hole methods (Table 5). The most common xylophagous insects detected using bark sheets on *P. abies* were *Polygraphus poligraphus* (31.0%), *Molorchus* sp. (27.7%), and *Callidium* sp. (18.0%), while those on *Larix* sp. were *Callidium* sp. (35.7%), *Cerambycidae* sp. (27.2%), and *Rhagium* sp. (17.5%) (Table 5). The most common xylophagous insects detected using the exit hole method on *P. abies* were *Pityogenes chalcographus* (44.2%), *Hylurgops palliatus* (25.5%), and *Trypodendron lineatum* (16.6%), while on *Larix* sp., they were *Buprestidae* sp. (28.6%), *Scolytinae* sp. (22.6%), and *Cerambycidae* sp. (22.0%) (Table 5).

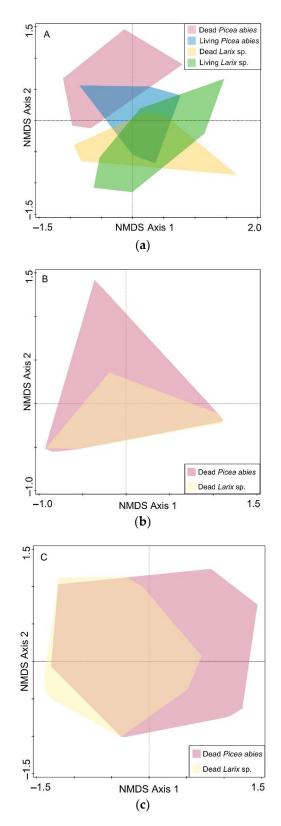


Figure 3. Ordination diagram based on nonmetric multidimensional scaling of insect communities detected in association with *Picea abies* and *Larix* sp. trees. Insects were assessed using: (**a**) sticky traps attached to the surface of dead and living trees (47.1% variation explained on axis 1 and 31.9% explained on axis2, (**b**) bark sheets removed from dead trees (52.7% on axis 1 and 31.8% on axis 2), and (**c**) insect exit holes recorded on dead trees (47.2% on axis 1 and 28.5% on axis 2).

Order/Family

sect spe	ecies trap	ped us	sing sticl	ky traps.	All stu	dy sites	are com	bined.				
cea abi	es Trees							Larix s	p. Trees			
у	Aug	ust	То	tal	Ju	ne	Ju	ly	Aug	gust	То	tal
L	D	L	D	L	D	L	D	L	D	L	D	L
87.0	59.2	_	83.8	82.7	_	_	03	_	_	_	0.2	_

Table 3. Relative abundance (%) of the 20 most common insect specie	ies trapped using sticky traps. All study sites are combined.

Picea

July

D

June D* L**

Coleoptera/Curculionidae	Crypturgus pusillus Erich.	91.6	80.0	69.8	87.0	59.2	-	83.8	82.7	-	-	0.3	-	-	-	0.2	-
Hymenoptera/Ichneomonidae	Ichneomonidae sp.	1.9	3.1	6.9	5.1	8.0	22.8	3.6	5.0	23.2	26.2	24.8	29.1	30.0	28.1	24.6	27.8
Coleoptera/Eucnemidae	<i>Eucnemidae</i> sp. 1	1.2	3.9	2.1	1.3	1.2	3.9	1.4	2.1	25.9	23.2	31.9	18.8	5.4	5.6	27.0	19.0
Coleoptera/Eucnemidae	Eucnemidae sp. 2	0.7	3.2	2.6	1.5	1.8	6.3	1.3	2.1	4.9	13.9	7.5	14.8	4.1	8.2	6.1	13.7
Coleoptera/Elateridae	Dalopius marginatus L.	0.7	2.9	-	-	-	-	0.4	0.8	19.0	11.5	-	-	-	-	7.8	4.5
Coleoptera/Anobiidae	Hadrobregmus pertinax L.	0.2	-	3.8	0.1	-	-	1.2	0.1	1.7	-	5.0	1.8	-	-	3.2	0.9
Coleoptera/Elateridae	Conoderus sp.	-	-	2.7	0.5	1.7	2.8	0.9	0.4	0.5	0.6	3.4	3.1	7.3	1.2	2.6	1.9
Coleoptera/Cleridae	Tillus elongatus L.	0.1	0.6	1.2	0.7	0.3	-	0.4	0.7	2.9	1.9	5.9	5.5	-	0.5	4.1	3.5
Coleoptera/Cleridae	Thanasimus formicarius L.	0.4	0.6	1.7	0.5	1.8	0.7	0.9	0.5	1.5	1.7	1.0	0.5	-	-	1.1	0.9
Coleoptera/Anthribidae	Anthribus nebulosus Forst.	0.1	0.6	0.5	0.6	2.0	15.3	0.3	1.0	1.1	0.3	0.6	1.8	7.4	10.3	1.4	2.2
Coleoptera/Ptinidae	Anobium rufipes Fabr.	0.1	0.3	1.5	0.3	0.3	2.5	0.5	0.4	-	2.3	3.3	-	-	-	1.6	0.9
Coleoptera/Scolytidae	Polygraphus poligraphus L.	0.2	-	0.0	0.1	12.1	1.0	0.7	0.1	0.8	-	0.2	-	0.6	1.1	0.5	0.1
Hymenoptera/Formicidae	Formica rufa L.	0.1	0.7	0.1	0.3	1.8	16.0	0.2	0.8	-	0.8	-	0.3	0.6	4.1	0.1	0.9
Coleoptera/Scolytidae	Trypodendron lineatum Ol.	0.8	0.4	0.2	-	0.3	0.7	0.6	0.1	1.6	0.3	-	0.2	3.2	4.5	1.0	0.7
Coleoptera/Cantharidae	<i>Malthodes</i> sp.	-	0.3	0.2	0.3	0.6	0.8	0.1	0.3	0.3	1.4	1.2	3.1	1.8	-	0.9	2.1
Coleoptera/Scolytidae	Pityogenes chalcographus L.	0.1	0.1	1.4	0.1	-	-	0.5	0.1	-	0.3	-	-	-	-	-	0.1
Coleoptera/Nitidulidae	Glischrochilus hortensis Geoffr.	0.2	0.2	0.3	-	-	0.3	0.2	0.1	1.3	0.9	0.8	0.3	-	-	0.9	0.5
Coleoptera/Trogossitidae	Nemozoma elongatum L.	2.2	0.1	0.6	-	0.3	-	0.3	0.1	-	-	-	-	-	-	-	-
Coleoptera/Dermestidae	Megatoma undata L.	0.1	0.1	0.5	0.1	-	-	0.2	0.1	0.5	1.2	0.2	-	-	-	0.3	0.5
Hymenoptera/Myrmicidae	<i>Myrmica</i> sp.	-	-	0.2	0.1	0.5	5.5	0.1	0.2	-	-	-	0.9	1.3	3.9	0.1	0.9
Total of 20	0 species	98.7	97.2	96.4	98.9	91.9	78.6	97.7	97.9	85.1	86.5	86.3	80.0	61.8	67.5	83.5	81.1

* D: dead trees; ** L: living trees.

Species

			Bark Shee	ets			Exit Hole	S	
Site	Tree Species	Richness, % (No. of Insect Species)	Bark Area Colonized, %	Shannon H	Sørensen Cs	Richness, % (No. of Insect Species)	Amount, % (No. of Exit Holes)	Shannon H	Sørensen Cs
S1/L1	Picea Larix	44.4 (4)	54.2	1.03	-	62.5 (5)	3.3 (109)	0.79	-
То	otal	44.4 (4)	54.2	-		62.5 (5)	3.3 (109)	-	
S2/L2	Picea Larix	22.2 (2) 44.4 (4)	18.3 49.2	0.56 0.23	0.67	50.0 (4) 50.0 (4)	0.9 (3)1 3.2 (108)	0.84 0.91	1.00
То	otal	44.4 (4)	40.0	0.58	0.07	50.0 (4)	4.2 (139)	1.08	1.00
S3/L3	Picea Larix	22.2 (2) 11.1 (1)	35.7 58.0	0.69 0.00	0.67	50.0 (4) 25.0 (2)	2.9 (96) 0.3 (10)	0.95 0.33	0.33
To	otal	22.2 (2)	45.0	0.57		62.5 (5)	3.2 (106)	1.18	
S4/L4	Picea Larix	44.4 (4) 22.2 (2)	42.5 75.0	0.97 0.64	0.00	75.0 (6) 37.5 (3)	8.2 (273) 1.6 (53)	1.46 1.08	0.44
To	otal	66.7 (6)	49.0	1.48		87.5 (7)	9.8 (326)	1.64	
S5/L5	Picea Larix	44.4 (4)	55.0	1.27 -	-	50.0 (4)	3.0 (100)	0.60	-
То	otal	44.4 (4)	55.0	-		50.0 (4)	3.0 (100)	-	
S6/L6	Picea Larix	55.6 (5) 22.2 (2)	13.8 38.0	1.10 0.60	0.57	62.5 (5) 37.5 (3)	9.0 (300) 3.2 (108)	1.13 0.77	0.50
Te	otal	55.6 (5)	23.1	1.01		75.0 (6)	12.3 (408)	1.34	
S7/L7	Picea Larix	22.2 (2) 33.3 (3)	71.0 69.0	0.59 1.08	0.40	62.5 (5) 62.5 (5)	18.3 (609) 1.8 (60)	0.83 1.33	0.60
Te	otal	44.4 (4)	70.0	1.24		87.5 (7)	20.1 (669)	1.09	
S8/L8	Picea Larix	22.2 (2) 11.1 (1)	23.5 20.0	0.52 0.00	0.00	87.5 (7) 50.0 (4)	10.0 (333) 3.2 (106)	0.81 0.79	0.73
Te	otal	33.3 (3)	22.0	0.98		87.5 (7)	13.2 (439)	0.93	
S9/L9	Picea Larix	44.4 (4) 22.2 (2)	53.3 59.0	1.00 0.63	0.33	87.5 (7) 50.0 (4)	16.7 (556) 7.3 (242)	1.35 0.72	0.55
To	otal	55.6 (5)	55.9	1.43	0.000	100 (8)	24.0 (798)	1.72	0.000
S10/L1	0 Picea Larix	22.2 (2) 33.3 (3)	31.4 29.2	0.66 0.43	0.80	37.5 (3) 25.0 (2)	3.1 (102) 3.3 (111)	0.85 0.48	0.40
Te	otal	33.3 (3)	30.4	1.07		50.0 (4)	3.4 (113)	1.21	
All sites	Picea Larix	88.9 (8) 77.8 (7)	39.0 47.3	1.65 1.55	0.80	100 (8) 87.5 (7)	75.4 (2509) 24.6 (819)	1.44 1.61	0.80
All	total	100 (9)	42.2	1.94		100 (8)	100 (3328)	1.77	

Table 4. Diversity of xylophagous insects detected using bark sheet and exit hole methods on dead *Picea abies* and *Larix* sp.

Table 5. Relative abundance (%) of xylophagous insects colonizing dead wood of *Picea abies* and/or *Larix* sp. detected using bark sheet and exit holes methods.

Order/Family	Incast Spacing	Bark S	Sheets	Exit Holes		
Order/ramity	Insect Species	Picea abies	Larix sp.	Picea abies	Larix sp.	
Coleoptera/Cerambycidae	<i>Callidium</i> sp. Fabr.	18.0	35.7	-	-	
Coleoptera/Cerambycidae	<i>Cerambycidae</i> sp. Latr	0.2	27.2	4.3	22.0	
Coleoptera/Curculionidae	Ips typographus L.	11.0	-	7.1	-	
Coleoptera/Cerambycidae	Molorchus sp. Fabr.	27.7	5.6	-	-	
Coleoptera/Curculionidae	Polygraphus poligraphus L.	31.0	-	-	-	
Coleoptera/Curculionidae	Rhagium sp. Fabr.	7.0	17.5	-	-	

Order/Fermiler	Incost Succios	Bark S	heets	Exit Holes		
Order/Family	Insect Species	Picea abies	Larix sp.	Picea abies	Larix sp.	
Coleoptera/Curculionidae	<i>Scolytinae</i> sp. Latr.	3.8	2.9	1.2	22.6	
Hymenoptera/Siricidae	Siricidae sp. Fabr.	1.2	0.3	-	-	
Coleoptera/Cerambycidae	<i>Tetropium</i> sp. Kirby	-	10.8	-	-	
Coleoptera/Curculionidae	Pityogenes chalcographus L.	-	-	44.2	6.3	
Coleoptera/Curculionidae	Trypodendron lineatum Oliv.	-	-	16.6	17.8	
Coleoptera/Buprestidae	<i>Buprestidae</i> sp. Leach	-	-	0.9	28.6	
Coleoptera/Curculionidae	Hylurgops palliatus Gyll.	-	-	25.5	-	
Hymenoptera/Siricidae	Sirex juvencus L.	-	-	0.2	2.7	

Table 5. Cont.

3.2. Lichens

The overall lichen community detected in the present study consisted of twelve species, among which eight were on dead *P. abies*, ten were on dead *Larix* sp., ten were on living *P. abies*, and eleven were on living *Larix* sp. (Table 6). The bark area colonized by lichens was 34.3% on dead *P. abies* and 63.2% on dead *Larix* sp., and 40.4% on living *P. abies* and 78.0% on living *Larix* sp. (Table 6).

Table 6. Diversity and occurrence of epiphytic lichens on the bark of dead and living *Picea abies* and *Larix* sp. trees.

Site	Tree State	Tree Species	Richness, % (No. of Lichen Species)	Bark Area Colonized, %	Shannon H	Sørensen Cs *
	Live	Picea Larix	66.6 (8) 50.0 (6)	89.5 71.2	1.18 1.25	0.71
S1/L1	Dead	Picea	66.6 (8)	95.5	1.06	-
	Deud	Larix	-	-	-	
	Т	otal	75.0 (9)	85.8	1.31	0.80
	Live	Picea Larix	58.3 (7) 50.0 (6)	51.8 28.5	1.61 1.35	0.62
S2/L2	Dead	Picea Larix	58.3 (7) 33.3 (4)	40.4 26.8	1.44 1.09	0.73
	Т	otal	75.0 (9)	47.7	1.54	0.62
	Live	Picea Larix	8.3 (1) 75.0 (9)	20.0 77.2	0.00 1.52	0.20
S3/L3	Dead	Picea Larix	- 33.3 (4)	- 86.1	- 1.11	-
	Т	otal	75.0 (9)	50.2	1.44	0.20
	Live	Picea Larix	50.0 (6) 58.3 (7)	37.3 81.1	1.17 1.13	0.62
S4/L4	Dead	Picea Larix	16.7 (2) 41.7 (5)	17.8 49.5	0.53 0.85	0.29
	Т	otal	83.3 (10)	48.7	1.17	0.57
	Live	Picea Larix	16.7 (2) 75.0 (9)	19.0 83.2	0.65 1.13	0.36
S5/L5	Dead	Picea Larix	41.7 (5)	40.2	1.00	-
	Т	otal	75.0 (9)	49.6	1.32	0.71

Site	Tree State	Tree Species	Richness, % (No. of Lichen Species)	Bark Area Colonized, %	Shannon H	Sørensen Cs *
	Live	Picea Larix	25.0 (3) 41.7 (5)	26.0 84.9	1.03 0.67	0.75
S6/L6	Dead	Picea Larix	8.3 (1) 50.0 (6)	5.0 89.2	0.00 0.82	0.29
-	Т	otal	50.0 (6)	54.2	0.96	0.80
	Live	Picea Larix	66.7 (8) 58.3 (7)	58.3 71.7	1.13 0.70	0.80
S7/L7	Dead	Picea Larix	8.3 (1) 16.7 (2)	12.0 31.0	$\begin{array}{c} 0.00\\ 0.14\end{array}$	0.00
-	Т	otal	75.0 (9)	44.4	1.11	0.80
	Live	Picea Larix	58.3 (7) 58.3 (7)	46.8 57.4	0.96 1.42	0.71
S8/L8	Dead	Picea Larix	58.3 (7) 75.0 (9)	55.9 49.2	1.34 1.36	0.75
-	Т	otal	83.3 (10)	52.5	1.42	0.75
	Live	Picea Larix	8.3 (1) 33.3 (4)	17.0 80.9	0.00 1.21	0.40
S9/L9	Dead	Picea Larix	25.0 (3) 25.0 (3)	28.7 90.1	0.58 0.86	0.67
-	Т	otal	50.0 (6)	59.9	1.10	0.25
	Live	Picea Larix	66.7 (8) 58.3 (7)	24.8 4.8	1.39 1.17	0.80
S10/L10	Dead	Picea Larix	41.7 (5) 8.3 (1)	22.5 68.0	1.28 0.00	0.33
-	Т	otal	75.0 (9)	56.3	1.16	0.80
		e Picea e Larix	83.3 (10) 91.7 (11)	40.4 78.0	1.42 1.48	0.86
All sites		d Picea d Larix	66.7 (8) 83.3 (10)	34.3 63.2	1.38 1.11	0.89
-	All Total		100 (12)	54.8	1.45	0.86

Table 6. Cont.

* Sørensen similarity index in rows "Total" shows the comparison between all *Picea abies* and all *Larix* sp. trees within adjacent study sites, e.g., S2 and L2.

ANOVA showed that the bark area colonized by lichens was significantly larger on dead and living *Larix* sp. trees than on corresponding *P. abies* trees (p < 0.0001). The Mann–Whitney test showed that the Shannon diversity index of lichen communities was similar between living *P. abies* and *Larix* sp. trees (p > 0.05) and between dead *P. abies* and *Larix* sp. trees (p > 0.05) (Table 6).

The most abundant lichen species was *Lepraria* sp., which composed 69.9% of the total bark area colonized by lichens. The relative abundance of this species was 60.4% and 59.9% on dead and living *P. abies*, respectively, and 76.4% and 71.4% on dead and living *Larix* sp., respectively (Table 7). The other most common lichen species detected on dead and living *P. abies* were *Phlyctis argena* (13.6% and 19.9%, respectively) and *Lecidea elaeochroma* (6.7% and 7.7%, respectively), while those on dead and living *Larix* sp. were *Hypogimnia physodes* (14.3% and 6.5%, respectively) and Unidentified sp. 1 (4.3% and 6.7%, respectively). Unidentified sp. 2, with a relative abundance of 7.0%, was detected only on living *Larix* sp. trees (Table 7).

Family	Lichen Species	Picea	abies	Lar	ix sp.	Total
ганнту	Lichen Species	Dead	Living	Dead	Living	Iotai
Stereocaulaceae	<i>Lepraria sp.</i> Ach.	60.4	59.9	76.4	71.4	69.9
Parmeliaceae	Hypogimnia physodes (L.) Nyl.	3.3	4.8	14.3	6.5	8.1
Phlyctidaceae	Phlyctis argena Spreng.	13.6	19.9	1.9	1.9	5.8
Unknown	Unidentified sp. 1	4.1	5.1	4.3	6.5	5.4
Lecanoraceae	Lecidea elaeochroma Ach.	6.7	7.7	1.4	2.4	3.4
Unknown	Unidentified sp. 2	-	-	-	7.0	3.2
Parmeliaceae	Parmelia sulcate Taylor.	5.5	1.0	0.9	1.3	1.6
Unknown	Unidentified sp. 3	4.8	1.1	0.2	0.3	0.9
Physciaceae	Physcia stellaris (L.) Nyl.	1.6	0.2	0.3	1.0	0.8
Teloschistaceae	Xanthoria parietina (L.) Th.Fr.	-	0.3	0.1	1.2	0.6
Unknown	Unidentified sp. 4	-	-	0.1	0.6	0.3
Unknown	Unidentified sp. 5	-	0.1			0.0

Table 7. Relative abundance (%) of epiphytic lichen species on colonized dead and living *P. abies* and *Larix* sp. trees. Different sampling sites are combined.

NMDS showed that lichen communities associated with dead and living trees of *P. abies* and *Larix* sp. were largely the same and thus overlapping (Figure 4). In agreement, the Sørensen similarity index of lichen communities was 0.86 between *P. abies* and *Larix* sp. trees, showing a high species similarity (all study sites combined) (Table 6).

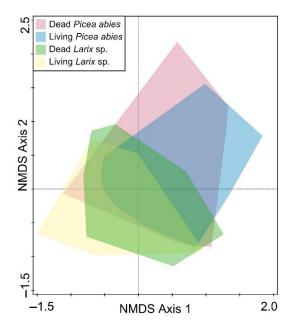


Figure 4. Ordination diagram based on nonmetric multidimensional scaling (NMDS) of lichen communities on dead and living trees of *Picea abies* and *Larix* sp. In NMDS, 44.2% variation was explained on axis 1, and 34.5% was explained on axis 2.

4. Discussion

The results demonstrate that the two coniferous tree species, namely, native *P. abies* and introduced *Larix* sp., support a similar diversity of stem-associated insect and lichen communities, but the species composition was only partially overlapping (Figures 2–4 and Tables 3–7). Consistently, for both insects and lichens, the Sørensen similarity index ranged from moderate to high, while the Shannon diversity index was similar between the two tree species. Therefore, *Larix* sp. has the potential to provide suitable habitats for some insect and lichen species associated with stems of *P. abies*. However, other organisms associated with these tree species should also be considered, as the replacement of native tree species

by introduced ones is a drastic event and may affect biodiversity at both local and regional scales [68]. In addition, different groups of organisms may respond to introduced tree species, i.e., to a new habitat, in different ways [69]. Several studies have evaluated the effect of introduced trees on a particular group of organisms, e.g., plants [70], insects [29,30,71,72], or birds [73,74]. Nevertheless, similar studies that simultaneously assessed different groups of organisms are scarce but can be particularly valuable [75], especially if a number of factors, such as the age of forest stands, microclimate conditions, the type of forest management, or the volume of deadwood, are taken into consideration [76–79], as these may also have a strong impact on associated biodiversity [80].

4.1. Insects

In the present study, the use of different assessment methods (sticky traps, bark sheets, and exit holes) provided a comprehensive comparison of the diversity and composition of stem-associated insects (Tables 3–5 and Figure 3), thereby allowing the overall insect diversity to be estimated [39,81,82]. Among these methods, sticky traps represent one of the most commonly used types of passive traps [83], but additional methods are often needed, as these may provide valuable complementary information [84,85]. However, to increase the accuracy of species identification, additional methods such as DNA sequencing may be needed, as for several insects trapped, the species identity could not be established using morphological methods (Table 3). Nevertheless, sticky traps allowed the collection of important and host-tree-specific insect species but, in some cases, also resulted in unspecific individuals, e.g., *Dalopius marginatus* L., *Conoderus* sp. (Coleoptera: Elateridae), Formica rufa L. (Hymenoptera: Formicidae), Myrmica sp. (Hymenoptera: Myrmicidae), or Malthodes sp. (Coleoptera: Cantharidae) (Table 3). Although the latter insects are abundant in the Palaearctic and Nearctic regions and play important roles in forest ecosystems [86], they are not specifically associated with *P. abies* or *Larix* sp. Beetles dominated insect communities in sticky traps, among which two species of aggressive bark beetles, i.e., *Polygraphus poligraphus* and *Pityogenes chalcographus*, which regularly attack and can kill living trees, were detected (Table 3). Interestingly, P. poligraphus and P. chalcographus are among the phloeophagous insect species, which are known to be specifically associated with the genus Picea [87], but in the present study, these were detected on both P. abies and *Larix* sp. (Table 3). Similarly, on both tree species, there were also several secondary bark beetle species, which are deadwood-dependent and colonize trees following attacks by aggressive bark beetles. These included Crypturgus pusillus, Hadrobregmus pertinax, Anobium rufipes, and Trypodendron lineatum (Table 3). Among these, C. pusillus is known to be a P. abies-dependent species that colonizes trees following attacks by I. typographus [87,88].

Interactions between xylophagous insects and their predators are common in nature and may have a direct effect on the health and sustainability of forest stands [89]. Several predators were detected, among which probably the most interesting was *Nemozoma* elongatum (Coleoptera: Trogossitidae), as it is one of the most important predators of *P. chalcographus* [90]. However, *N. elongatum* was captured in low abundance and only on dead and living *P. abies* (Table 3), even though its trapping coincided with the flying period (June–July) of P. chalcographus [91]. Zahradník and Zahradníková [92], using pheromone baited traps, showed a strong positive correlation between the abundance of *P. chalcographus* (1–4%) and N. elongatum (up to 60%). Among other predatory insects captured on P. abies and Larix sp. tree stems were Thanasimus formicarius, Tillus elongatus, Glischrochilus hortensis, and Anthribus nebulosus (Table 3). Thanasimus formicarius and Glischrochilus hortensis are predators of many different bark beetle species from the subfamily Scolytinae, including *I. typographus* [89,93–96]. *Tillus elongatus* is also a predator of bark beetles, attacking them in larval tunnels [97]. Anthribus nebulosus is a predator of soft-scale insects from the family Coccidae [98]. Despite the detection, the relative abundance of predator insects was low, and in many cases, the host insects were absent, suggesting that their trapping could be accidental. The use of sticky traps also revealed the presence of Tetropium gabrieli on Larix sp. trees (Table S1), which is an important secondary pest of *Larix* sp. in Europe and was

detected for the first time in Lithuania [99]. Among the deadwood-dependent insects, there were two species from the family *Eucnemidae*, which were trapped on both *P. abies* and *Larix* sp. (Table 3). Larvae of these insects develop in the wood of dead or dying deciduous or coniferous trees [85,100]. *Eucnemidae* may play an important role in the interactions between trees, fungi, and forest regeneration and can be used as an indicator species of forest biodiversity [101].

Many previous studies have shown that deadwood is one of the most important substrates in forests and supports a high diversity of xylophagous insects [88,102–104]. Indeed, deadwood as a substrate is required for many species of beetles, bees, wasps, ants, flies, mosquitoes, and other invertebrates [100,105–110]. It may serve as a resource for feeding, breeding, overwintering, or refuge [39,88,111,112]. In the present study, xylophagous insects detected in dead trees of *P. abies* and *Larix sp.* using bark sheet and exit hole methods were rather different as compared to those detected using sticky traps (Tables 3–5), thereby repeatedly showing that all of these methods complemented each other. However, the diversity of xylophagous insects detected using bark sheet and exit hole methods was generally limited (Table 4). In comparison, other studies have shown a much higher diversity of xylophagous insect species associated with deadwood of P. abies. For example, there were 47 insect species reported by Jonsell and Weslien [113] and 66 species reported by Seedre [87]. The lower diversity of xylophagous insects could be due to specific stand characteristics, i.e., middle-aged monocultures with routine and intensive forest management and a relatively low occurrence of dead trees (Table 1). Intensive forest management was also shown to have a negative impact on the diversity of xylophagous insects [114]. In addition, the diversity and composition of xylophagous insects may also depend on other factors, such as tree species, degree of decay, and the cause of tree death [115].

Although on *P. abies* and *Larix* sp., the diversity and composition of xylophagous insects were similar (Figure 3b,c), the use of bark sheet and exit hole methods showed certain specificity, which can probably be attributed to the biology and ecology of specific insect species. For example, adults of P. chalcographus and H. palliatus make numerous exit holes on tree stems, but their larvae are relatively small and colonize a relatively small area as compared to large larvae of Callidium sp., which was detected using the bark sheet method (Table 5). Furthermore, Trypodendron lineatum was abundantly detected on both tree species, but only using the exit hole method (Table 5). Galleries of T. lineatum are found ca. 7 cm deep in the wood and are undetectable using the bark sheet method [116]. Similarly, Sirex juvencus was detected on both P. abies and Larix sp. using the exit hole method, as its larvae occur ca. 15–30 cm deep in the wood and leave no signs of activity under the bark [117]. Siricidae woodwasps make circular and smooth-edged exit holes of ca. 4–10 mm in diameter, making identification of the species relatively easy [118,119]. Despite the importance of the conservation of many xylophagous insect species and the promotion of deadwood habitats, the risk of bark beetle outbreaks should also be considered [120]. Bark beetle species such as *I. typographus*, *P. polygraphus*, or *P. chalcographus*, which usually colonize weakened and/or dying trees, can cause extensive damage [121]. In the present study, these were mainly associated with *P. abies*, suggesting that *Larix* sp. under the given conditions was less susceptible to their attack (Table 5). However, it was shown that Larix sp. can be vulnerable to attacks by bark beetles of the genus Ips, including *I. typographus* and *I. cembrae* [34,122]. Therefore, slight differences in the composition of xylophagous insects between *Larix sp.* and *P. abies* trees can probably be explained by certain host specificity. Xylophagous beetles colonizing fresh wood or dying trees need to overcome the tree resistance in the form of chemical barriers [123] and, therefore, are much more host-adapted than those of later decomposition stages [102]. Interestingly, Muller et al. [124] showed a low ranking of *Larix decidua* as the host, which was due to a generally lower number of herbivorous species, including saproxylic beetles, colonizing this tree species as compared to other coniferous tree species, e.g., *P. abies*.

4.2. Lichens

In the present study, the diversity of epiphytic lichens was generally low on both *P. abies* and *Larix* sp. trees (Tables 6 and 7). By contrast, Giordani et al. [50] reported a relatively high diversity of lichen species in mixed *P. abies* forests, but this diversity was similar between *P. abies* and *Larix decidua* trees. It is known that epiphytic lichens can be sensitive to several abiotic factors, such as light [125], temperature and annual precipitation [45–47,126], pH value and nutrient availability on the tree bark [42], and air pollution [43,44]. Forest structure and dynamics are among other determinants of the diversity of epiphytic lichens [48,49,127,128]. In addition, the diversity and biomass of epiphytic lichens appear to be higher in unmanaged old-growth forests than in managed ones [128]. Indeed, Marmor et al. [125] showed that on *P. abies*, the diversity of lichen species (see above), specific characteristics of *P. abies* and *Larix* sp. stands (Table 1) were likely among the main determinants of the low diversity of epiphytic lichens.

In agreement with results of the present study, Hauck [42] and Marmor et al. [125] showed that *Lepraria* sp. and *H. physodes* were among the most dominant lichen species on *P. abies* in boreal forests of Europe. Interestingly, both lichen species showed a higher preference for dead or living *Larix* sp. than for corresponding *P. abies* (Table 7). By contrast, P. argena showed a higher preference for dead or living P. abies than for corresponding *Larix* sp. (Table 7). Several studies have emphasized the effect of the tree species on the diversity and composition of lichen communities, e.g., [129,130]. This effect appears to be mainly due to species-specific differences in chemical and physical traits of the bark, e.g., [129,131]. Bark pH, which is usually between 3.0 and 4.0 for different conifer tree species [42], is among the principal factors that determine the occurrence and abundance of epiphytic lichens [132]. As the pH of the bark for both *P. abies* and *Larix sp.* was shown to be similar [133,134], this has likely led to the overlap of lichen communities associated with dead or living *P. abies* and *Larix* sp. (Figure 4). Consequently, the detected lichen species appear to be generalists, i.e., adapted to different tree species, as they only showed a preference for a particular tree species to a small extent. On the other hand, the larger bark area colonized by lichens on dead and living Larix sp. than on corresponding P. abies (Table 6) shows that the growth of lichens is faster on the former tree species.

In summary, the results revealed that *P. abies* and *Larix* sp. share a large number of stem-associated insect and lichen species. As climate change can be expected to have a strong negative effect on *P. abies* in the area, its gradual replacement by *Larix* sp. is likely to provide appropriate habitats for investigated insects and lichens, thereby supporting forest biodiversity. However, the possibility should not be excluded that some wood-boring insect species will not be able to jump between host tree species and may be lost if the mortality of *P. abies* drastically increases in the future.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/d14090729/s1. Table S1: Relative abundance (%) of insect species detected using sticky traps on dead and living trees of *Picea abies* and *Larix* sp. S1–S10 and L1–L10 denote different study sites.

Author Contributions: Conceptualization, A.M. (Audrius Menkis); methodology, A.G. and J.L.; software, A.M. (Audrius Menkis) and A.M. (Adas Marčiulynas); validation, A.M. (Audrius Menkis) and J.L.; formal analysis, J.L.; investigation, A.G., J.L., A.M. (Adas Marčiulynas), and D.M.; resources, A.G.; data curation, A.G. and J.L; writing—original draft preparation, J.L. and A.M. (Audrius Menkis); writing—review and editing, J.L. and A.M. (Audrius Menkis); visualization, A.M. (Adas Marčiulynas); supervision, A.M. (Audrius Menkis); project administration, D.M.; funding acquisition, A.M. (Audrius Menkis). All authors have read and agreed to the published version of the manuscript.

Funding: This project has received funding from the European Social Fund (project no. 09.3.3-LMT-K-712-01-0039) under a grant agreement with the Research Council of Lithuania (LMTLT).

Institutional Review Board Statement: Not applicable.

Data Availability Statement: Data can be supplied by the corresponding author upon reasonable request.

Acknowledgments: We thank Vytautas Tamutis for his help with the identification of insect species.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

References

- Buras, A.; Menzel, A. Projecting Tree Species Composition Changes of European Forests for 2061–2090 Under RCP 4.5 and RCP 8.5 Scenarios. *Front. Plant Sci.* 2019, 9, 1986. [CrossRef] [PubMed]
- Rigling, A.; Bigler, C.; Eilmann, B.; Feldmeyer-Christe, E.; Gimmi, U.; Ginzler, C.; Graf, U.; Mayer, P.; Vacchiano, G.; Weber, P.; et al. Driving factors of a vegetation shift from Scots pine to pubescent oak in dry Alpine forests. *Glob. Chang. Biol.* 2013, 19, 229–240. [CrossRef] [PubMed]
- Fekete, I.; Lajtha, K.; Kotroczó, Z.; Várbíró, G.; Varga, C.; Tóth, J.A.; Demeter, I.; Veperdi, G.; Berki, I. Long-term effects of climate change on carbon storage and tree species composition in a dry deciduous forest. *Glob. Chang. Biol.* 2017, 23, 3154–3168. [CrossRef] [PubMed]
- 4. Scherrer, D.; Massy, S.; Meier, S.; Vittoz, P.; Guisan, A. Assessing and predicting shifts in mountain forest composition across 25 years of climate change. *Divers Distrib.* 2017, 23, 517–528. [CrossRef]
- 5. Bigler, C.; Bräker, O.U.; Bugmann, H.; Dobbertin, M.; Rigling, A. Drought as an inciting mortality factor in Scots pine stands of the Valais, Switzerland. *Ecosystems* **2006**, *9*, 330–343. [CrossRef]
- Kohler, M.; Sohn, J.; Nägele, G.; Bauhus, J. Can drought tolerance of Norway spruce (*Picea abies*) be increased through thinning? *Eur. J. For. Res.* 2010, 129, 1109–1118. [CrossRef]
- Lévesque, M.; Saurer, M.; Siegwolf, R.; Eilmann, B.; Brang, P.; Bugmann, H.; Rigling, A. Drought response of five conifer species under contrasting water availability suggests high vulnerability of Norway spruce and European larch. *Glob. Chang. Biol.* 2013, 19, 3184–3199. [CrossRef]
- 8. Zang, C.; Hartl-Meier, C.; Dittmar, C.; Rothe, A.; Menzel, A. Patterns of drought tolerance in major European temperate forest trees: Climatic drivers and levels of variability. *Glob. Chang. Biol.* **2014**, *20*, 3767–3779. [CrossRef]
- 9. Huang, W.; Fonti, P.; Larsen, J.B.; Ræbild, A.; Callesen, I.; Pedersen, N.B.; Hansen, J.K. Projecting tree-growth responses into future climate: A study case from a Danish-wide common garden. *Agric. For. Meteorol.* **2017**, 247, 240–251. [CrossRef]
- 10. Martínez-Sancho, E.; Dorado-Liñán, I.; Hacke, U.G.; Seidel, H.; Menzel, A. Contrasting hydraulic architectures of Scots pine and sessile oak at their southernmost distribution limits. *Front. Plant Sci.* **2017**, *8*, 598. [CrossRef]
- Rehschuh, R.; Mette, T.; Menzel, A.; Buras, A. Soil properties affect the drought susceptibility of Norway spruce. *Dendrochronologia* 2017, 45, 81–89. [CrossRef]
- 12. Buras, A.; Schunk, C.; Zeiträg, C.; Herrmann, C.; Kaiser, L.; Lemme, H.; Straub, C.; Taeger, S.; Gößwein, S.; Klemmt, H.J. Are Scots pine forest edges particularly prone to drought-induced mortality? *Environ. Res. Lett.* **2018**, *13*, 25001. [CrossRef]
- 13. Honkaniemi, J.; Rammer, W.; Seidl, R. Norway spruce at the trailing edge: The effect of landscape configuration and composition on climate resilience. *Landscape Ecol.* **2020**, *35*, 591–606. [CrossRef] [PubMed]
- Johann, E.; Agnoletti, M.; Axelsson, A.L.; Bürgi, M.; Östlund, L.; Rochel, X.; Schmidt, U.E.; Schuler, A.; Skovsgaard, J.P.; Winiwater, V. History of secondary Norway Spruce forests in Europe. In *Norway Spruce Conversion Options and Consequences*; Hansen, J., Klimo, E., Spiecker, H., Eds.; Brill: Leiden, The Netherlands; Boston, MA, USA, 2004; pp. 25–62.
- 15. Jandl, R. Climate-induced challenges of Norway spruce in Northern Austria. Trees For. People 2020, 1, 100008. [CrossRef]
- 16. Schlyter, P.; Stjernquist, I.; Bärring, L.; Jönsson, A.M.; Nilsson, C. Assessment of the impacts of climate change and weather extremes on boreal forests in northern Europe, focusing on Norway spruce. *Clim. Res.* **2006**, *31*, 75–84. [CrossRef]
- 17. Hanewinkel, M.; Cullmann, D.; Schelhaas, M.J.; Nabuurs, G.J.; Zimmermann, N.E. Climate change may cause severe loss in the economic value of European forest land. *Nat. Clim. Chang.* 2013, *3*, 203–207. [CrossRef]
- 18. Pretzsch, H.; Schütze, G.; Uhl, E. Resistance of European tree species to drought stress in mixed versus pure forests: Evidence of stress release by inter-specific facilitation. *Plant Biol.* **2013**, *15*, 483–495. [CrossRef]
- 19. Zeng, H.; Garcia-Gonzalo, J.; Peltola, H.; Kellomaki, S. The effects of forest structure on the risk of wind damage at a landscape level in a boreal forest ecosystem. *Ann. For. Sci.* **2010**, *67*, 111. [CrossRef]
- Seidl, R.; Rammer, W.; Blennow, K. Simulating wind disturbance impacts on forest landscapes: Tree-level heterogeneity matters. Environ. Model Softw. 2014, 51, 1–11. [CrossRef]
- 21. Peltola, H.; Kellomäki, S.; Väisänen, H.; Ikonen, V.P. A mechanistic model for assessing the risk of wind and snow damage to singletrees and stands of Scots pine, Norway spruce, and birch. *Can. J. For. Res.* **1999**, *29*, 647–661. [CrossRef]
- 22. Seidl, R.; Spies, T.A.; Peterson, D.L.; Stephens, S.L.; Hicke, J.A. Searching for resilience: Addressing the impacts of changing disturbance regimes on forest ecosystem services. *J. Appl. Ecol.* **2016**, *53*, 120–129. [CrossRef] [PubMed]
- 23. Stadelmann, G.; Bugmann, H.; Wermelinger, B.; Bigler, C. Spatial interactions between storm damage and subsequent infestations by the European spruce bark beetle. *For. Ecol. Manag.* **2014**, *318*, 167–174. [CrossRef]
- 24. Lausch, A.; Heurich, M.; Fahse, L. Spatio-temporal infestation patterns of *Ips typographus* (L.) in the Bavarian Forest National Park, Germany. *Ecol. Indic.* 2013, *31*, 73–81. [CrossRef]

- Jacob, D.; Petersen, J.; Eggert, B.; Alias, A.; Christensen, O.B.; Bouwer, L.M.; Braun, A.; Colette, A.; De-que, M.; Georgievski, G.; et al. EURO-CORDEX: New high-resolution climate change projections for European impact research. *Reg. Environ. Chang.* 2014, 14, 563–578. [CrossRef]
- Hlásny, T.; Barka, I.; Roessiger, J.; Kulla, L.; Trombik, J.; Sarvašová, Z.; Bucha, T.; Kovalčík, M.; Čihák, T. Conversion of Norway spruce forests in the face of climate change: A case study in Central Europe. *Eur. J. Forest Res.* 2017, 136, 1013–1028. [CrossRef]
- 27. Seidl, R.; Vigl, F.; Rössler, G.; Neumann, M.; Rammer, W. Assessing the resilience of Norway spruce forests through a model-based reanalysis of thinning trials. *For. Ecol. Manag.* **2017**, *388*, 3–12. [CrossRef]
- Walentowski, H.; Falk, W.; Mette, T.; Kunz, J.; Bräuning, A.; Meinardus, C.; Zang, C.; Sutcliffe, L.; Luschner, C. Assessing future suitability of tree species under climate change by multiple methods: A case study in southern Germany. *Ann. For. Res.* 2017, 60, 101–126. [CrossRef]
- 29. Goßner, M.; Ammer, U. The effects of Douglas-fir on tree-specific arthropod communities in mixed species stands with European beech and Norway spruce. *Eur. J. Forest Res.* **2006**, *125*, 221–235. [CrossRef]
- Gossner, M.M. Introduced tree species in central Europe—Consequences for arthropod communities and species interactions. In Introduced Tree Species in European Forests: Opportunities and Challenges; Krumm, F., Vitková, L., Eds.; European Forest Institute: Freiburg, Germany, 2016; pp. 264–282.
- Lepage, B.; Basinger, J. The evolutionary history of the genus Larix (Pinaceae). In General Technical Report—Intermountain Research Station, USDA Forest Service, Proceedings of the Ecology and Management of Larix Forests: A Look Ahead, Whitefish, MT, USA, 5–9 October 1992; Schmidt, W.C., McDonald, K.J., Eds.; No.INT-GTR-319; US Department of Agriculture: Washington, DC, USA, 1995; Volume 5, pp. 19–29.
- 32. Danusevičius, J. Maumedžių veisimo patirtis ir perspektyvos Lietuvoje. Mūsų Girios 2006, 8, 8–10. (In Lithuanian)
- Lithuanian Statistical Yearbook of Forestry. 2017. Available online: http://www.amvmt.lt/images/veikla/stat/miskustatistika/ 2017/01%20Misku%20ukio%20statistika%202017_m.pdf (accessed on 21 February 2022).
- 34. Da Ronch, F.; Caudullo, G.; Tinner, W.; de Rigo, D. Larix decidua and other larches in Europe: Distribution, habitat, usage and threats. In *European Atlas of Forest Tree Species*; San-Miguel-Ayanz, J., de Rigo, D., Caudullo, G., Houston Durrant, T., Mauri, A., Eds.; Publications Office of the EU: Luxembourg, 2016; pp. 108–110. Available online: https://boris.unibe.ch/80793/1/Larix_ decidua.pdf (accessed on 10 March 2022).
- 35. Liu, Q.J. Structure and dynamics of the subalpine coniferous forest on Changbai mountain, China. *Plant Ecol.* **1997**, *132*, 97–105. [CrossRef]
- Zielonka, T.; Holeska, J.; Malcher, P. Disturbance events in a mixed spruce-larch forest in the Tatra Mts., Western Carpathians—A Tentative reconstruction. *Baltic Forestry* 2009, 15, 161–167.
- 37. Gradeckas, A.; Malinauskas, A. Miško Želdynu Veisimo Biologiniai Ir Ekologiniai Veiksniai Bei Patirtis Lietuvoje; Lututė: Kaunas, Lithuania, 2005; p. 404. (In Lithuanian)
- Žiogas, A.; Juronis, V.; Snieškienė, V. Pathological condition of Larix in Lithuania. In *Insects and Fungi in Storm Areas*; Proceedings of the IUFRO Working Party 7.03.10; Forest Research Institute: Zvolen, Slovakia, 2009; pp. 115–117.
- Salman, I.N.A.; Ferrante, M.; Möller, D.M.; Gavish-Regev, E.; Lubin, Y. Trunk Refugia: A Simple, Inexpensive Method for Sampling Tree Trunk Arthropods. J. Insect Sci. 2020, 20, 5. [CrossRef] [PubMed]
- 40. Price, P.W.; Denno, R.F.; Eubanks, M.D.; Finke, D.L.; Kaplan, I. *Insect Ecology: Behaviour, Populations and Communities*; Cambridge University Press: New York, NY, USA, 2011.
- 41. Brockerhoff, E.G.; Liebhold, A.M. Ecology of forest insect invasions. Biol. Invasions 2017, 19, 3141–3159. [CrossRef]
- 42. Hauck, M. Site factors controlling epiphytic lichen abundance in northern coniferous forests. Flora 2011, 206, 81–90. [CrossRef]
- 43. Cislaghi, C.; Nimis, P.L. Lichens, air pollution and lung cancer. *Nature* 1997, 387, 463–464. [CrossRef] [PubMed]
- Giordani, P. Is the diversity of epiphytic lichens a reliable indicator of air pollution? A case study from Italy. *Environ. Pollut.* 2007, 146, 317–323. [CrossRef]
- 45. Jovan, S.; McCune, B. Regional variation in epiphytic macrolichen communities in northern and central California forests. *Bryologist* **2004**, *107*, 328–339. [CrossRef]
- 46. Geiser, L.H.; Neitlich, P.N. Air pollution and climate gradients in western Oregon and Washington indicated by epiphytic macrolichens. *Environ. Pollut.* 2007, 145, 203–218. [CrossRef]
- 47. Giordani, P.; Incerti, G. The influence of climate on the distribution of lichens: A case study in a borderline area (Liguria, NW Italy). *Plant Ecol.* **2008**, *195*, 257–272. [CrossRef]
- 48. Hedenås, H.; Ericson, L. Epiphytic macrolichens as conservation indicators: Successional sequence in *Populus tremula* stands. *Biol. Conserv.* **2000**, *93*, 43–53. [CrossRef]
- 49. Johansson, P. Consequences of disturbance on epiphytic lichens in boreal and near boreal forests. *Biol. Conserv.* 2008, 141, 1933–1944. [CrossRef]
- 50. Giordani, P.; Brunialti, G.; Bacaro, G.; Nascimbene, J. Functional traits of epiphytic lichens as potential indicators of environmental conditions in forest ecosystems. *Ecol. Indic.* 2012, *18*, 413–420. [CrossRef]
- Roper, T. Lichen Abundance and Diversity in Relation to Host Tree Species and Lakeshore Proximity. *Conspec. Boreal.* 2018, *3*, 8.
 Navasaitis, M. *Dendrologija*, 2nd ed.; Margi Raštai: Vilnius, Lithuania, 2008; pp. 169–183. (In Lithuanian)
- 53. Scheepers, D.; Eloy, M.C.; Briquet, M. Identification of larch species (*Larix decidua*, *Larix kaempferi* and *Larix X eurolepis*) and estimation of hybrid fraction in seed lots by RAPD fingerprints. *Theor. Appl. Genet.* **2000**, 100, 71–74. [CrossRef]

- 54. Vaičys, M. Miško dirvožemiu klasifikacija. In Lietuvos Dirvožemiai; Mokslas: Vilnius, Lithuania, 2001; pp. 1040–1043. (In Lithuanian)
- 55. Karazija, S. Miško Tipologija. In *Miško Ekologija*; Padaiga, V., Stravinskienė, V., Eds.; Enciklopedija: Vilnius, Lithuania, 2008; pp. 220–254. (In Lithuanian)
- 56. Nageleisen, L.M.; Bouget, C. Forest insect studies: Methods and techniques, key considerations for standardization. In *An Overview of the Reflections of the Entomological Forest Inventories Working Group (Inv. Ent. For.)*; ONF: Paris, France, 2009; p. 144.
- 57. Tamutis, V.; Aleseev, V. A survey of Lepturinae Latreille, 1802 (Coleoptera: Cerambycidae) of the south- eastern Baltic region (Lithuania and the Kaliningrad Region). *Biologia* 2020, *66*, 169–235. [CrossRef]
- 58. Tamutis, V.; Tamutė, B.; Ferenca, R. A catalogue of Lithuanian beetles (Insecta, Coleoptera). ZooKeys 2011, 121, 1–494. [CrossRef]
- Alonso-Zarazaga, M.A.; Barrios, H.; Borovec, R.; Bouchard, P.; Caldara, R.; Colonnelli, E.; Gültekin, L.; Hlaváč, P.; Korotyaev, B.; Lyal, C.H.; et al. Cooperative Catalogue of Palaearctic Coleoptera Curculionoidea. *Monogr. Electrónicas SEA* 2017, *8*, 5–547. Available online: https://www.biotaxa.org (accessed on 21 February 2022).
- 60. Ivinskis, P. Lietuvos drugiai; Annotated Catalogue; Petro ofsetas: Vilsnius, Lithuania, 2004; p. 379. (In Lithuanian)
- 61. Zubrik, M.; Kunca, A.; Novotny, J. Atlas Poškodni Lesnych Drevin Hmyz a Huby; Národné lesnícke centrum–Lesnícky výskumný ústav: Zvolen Slovakia, 2008; p. 178.
- 62. Pileckis, S. Lietuvos vabalai; Mokslo Ir Enciklopedijų Leidykla: Vilnius, Lithuania, 1996; p. 303. (In Lithuanian)
- Asta, J.; Erhardt, W.; Ferretti, M.; Fornasier, F.; Kirschbaum, U.; Nimis, P.I.; Purvis, O.W.; Pirintsos, S.; Scheidegger, C.; van Haluwyn, C.; et al. Mapping lichen diversity as an indicator of environmental quality. In *Monitoring with Lichens—Monitoring Lichens*; Nimis, P.L., Scheidegger, C., Wolseley, P.A., Eds.; Kluwer Academic: Dordrecht, The Netherlands, 2002; pp. 273–279.
- 64. Sokal, R.R.; Rohlf, F.J. *Biometry: The Principles and Practice of Statistics in Biological Research*, 3rd ed.; W.H. Freeman and Company: New York, NY, USA, 1995.
- 65. Magurran, A.E. Ecological Diversity and Its Measurement; Princeton University Press: Princeton, NJ, USA, 1988; p. 192.
- 66. Shannon, C.E. A mathematical theory of communication. Bell Syst. Tech. J. 1948, 27, 379–423. [CrossRef]
- 67. ter Braak, C.J.F.; Smilauer, P. Canoco Reference Manual and User's Guide to Canoco for Windows: Software for Canonical Community Ordination, Version 4; Microcomputer Power: Ithaca, NY, USA, 1998; p. 351.
- 68. Brockerhoff, E.G.; Ecroyd, C.E.; Langer, E.R. Biodiversity in New Zealand plantation forests: Policy trends, incentives, and the state of our knowledge. *N. Z. J. For.* **2001**, *46*, 31–37.
- 69. Rymer, L. Pine plantations in Australia as habitat for native animals. Environ. Conserv. 1981, 8, 95–96. [CrossRef]
- 70. Michelsen, A.; Lisanework, N.; Friis, I.; Holst, N. Comparisons of understory vegetation and soil fertility in plantations and adjacent natural forests in the Ethiopian highlands. *J. Appl. Ecol.* **1996**, *33*, 627–642. [CrossRef]
- 71. Sinclair, J.E.; New, T.R. Pine plantations in southern eastern Australia support highly impoverished ant assemblages. *J. Insect Conserv.* 2004, *8*, 277–286. [CrossRef]
- 72. Corley, J.; Sackmann, P.; Rusch, V.; Bettinelli, J.; Paritsis, J. Effects of pine silviculture on the ant assemblages (Hymenoptera: Formicidae) of the Patagonian steppe. *For. Ecol. Manag.* **2006**, 222, 162–166. [CrossRef]
- 73. Clout, M.N.; Gaze, P.D. Effects of plantations forestry on birds in New Zealand. J. Appl. Ecol. 1984, 21, 795–815. [CrossRef]
- 74. Carlson, A. A comparison of birds inhabiting pine plantation and indigenous forest patches in a tropical mountain area. *Biol. Conserv.* **1986**, *35*, 195–204. [CrossRef]
- 75. Paritsis, J.; Aizen, M.A. Effects of exotic conifer plantations on the biodiversity of understory plants, epigeal beetles and birds in Nothofagus dombeyi forests. *For. Ecol. Manag.* **2008**, 255, 1575–1583. [CrossRef]
- Djupström, L.B.; Weslien, J.; Schroeder, L.M. Dead wood and saproxylic beetles in set-aside and non-set-aside forests in a boreal region. *For. Ecol. Manag.* 2008, 255, 3340–3350. [CrossRef]
- Jonsell, M.; Weslien, J.; Ehnström, B. Substrate requirements of red-listed saproxylic invertebrates in Sweden. *Biodivers Conserv.* 1998, 7, 749–764. [CrossRef]
- 78. Siitonen, J. Forest management, coarse woody debris and saproxylic organisms: Fennoscandian boreal forests as an example. *Ecol. Bull.* **2001**, *49*, 11–41.
- 79. Jonsell, M.; Nittérus, K.; Stighäll, K. Saproxylic beetles in natural and man-made deciduous high stumps retained for conservation. *Biol. Conserv.* 2004, 118, 163–173. [CrossRef]
- 80. Lassauce, A.; Lieutier, F.; Bouget, C. Wood-fuel harvesting and biodiversity conservation in temperate forests: Effects of logging residue characteristics on saproxylic beetle assemblages. *Biol. Conserv.* **2012**, *147*, 204–212. [CrossRef]
- 81. Erwin, T.L. Tropical forests: Their richness in Coleoptera and other arthropod species. Coleopt. Bull. 1982, 36, 74–75.
- Basset, Y.; Cizek, L.; Cuénoud, P.; Didham, R.K.; Guilhaumon, F.; Missa, O.; Novotny, V.; Ødegaard, F.; Roslin, T.; Schmidl, J.; et al. Arthropod diversity in a tropical forest. *Science* 2012, 338, 1481–1484. [CrossRef] [PubMed]
- 83. Bar-Ness, Y.D.; McQuillan, P.B.; Whitman, M.; Junker, R.R.; Cracknell, M.; Barrows, A. Sampling Forest canopy arthropod biodiversity with three novel minimal-cost trap designs. *Aust. J. Entomol.* **2012**, *51*, 12–21. [CrossRef]
- Hyvärinen, E.; Kouki, J.; Martikainen, P. A comparison of three trapping methods used to survey forest dwelling Coleoptera. *Eur. J. Entomol.* 2006, 103, 397–407. [CrossRef]
- 85. Hilszczański, J.; Plewa, R.; Jaworski, T.; Sierpiński, A. Microrhagus pyrenaeus Bonvouloir, 1872—A false click beetle new for the fauna of Poland with faunistic and ecological data on *Eucnemidae* (Coleoptera, Elateroidea). *Spixiana* **2015**, *38*, 77–84.
- Fanti, F.; Michalski, A.R. An unusual fossil *Malthodes* with long elytra (Insecta Coleoptera Cantharidae). G. Ital. Di Entomol. 2018, 15, 127–132.

- Seedre, M. Saproxylic Beetles in Artificially Created High Stumps of Spruce and Birch Three Years after Cutting. Master's Thesis, Southern Swedish Forest Research Centre, Alnarp, Sweden, 2005. Available online: https://www.academia.edu/6066983/ (accessed on 18 February 2022).
- 88. Wermelinger, B.; Duelli, P.; Obrist, M.K. Dynamics of saproxylic beetles (Coleoptera) in windthrow areas in alpine spruce forests. *For. Snow Landsc. Res.* **2002**, *77*, 133–148.
- Marini, L.; Lindelöw, Å.; Jönsson, A.M.; Wulff, S.; Schroeder, L.M. Population dynamics of the spruce bark beetle: A long-term study. Oikos 2013, 122, 1768–1776. [CrossRef]
- 90. Baier, P. Untersuchungen zur abundanzdynamischen Relevanz der Beifange von Nemusuma eluzzgatum (L.) (Col., Ostomidae) in C H A L C O P R A X ~ bekoderten Flugbarrierefallen fur Pityogenes chalcographus (L.) (Col., Scolytidae). J. Appl. Ent. 1994, 117, 51–57. [CrossRef]
- 91. Belova, O.; Milišauskas, Z.; Padaiga, V.; Valenta, V.; Vasiliauskas, A.; Zolubas, P.; Žiogas, A. *Miško Apsaugos Vadovas*, 1st ed.; Lututė: Kaunas, Lithuania, 2000; p. 351. (In Lithuanian)
- 92. Zahradník, P.; Zahradníková, M. The relationships between *Pityogenes chalcographus* and *Nemozoma elongatum* in clear-cuts with different types of management. *Plant Protect. Sci.* 2020, *56*, 30–34. [CrossRef]
- Skrzecz, I.; Bulka, M. Insect assemblages in Norway spruce [*Picea abies* (L.) Karst.] stumps in the Eastern Sudetes. *Folia For. Pol.* 2010, 52, 98–107.
- 94. Akkuzu, E.; Sariyildiz, T.; Kucuk, M.; Duman, A. *Ips typographus* (L.) and *Thanasimus formicarius* (L.) populations influenced by aspect and slope position in Artvin-Hatila valley national park, Turkey. *Afr. J. Biotechnol.* **2009**, *8*, 877–882.
- 95. Warzee, N.; Gregoire, J.C. *Thanasimus formicarius* (Coleoptera: Cleridae): Why a Large Range of Prey for a Specialized Predator? In Proceedings of the Forest Insect Population Dynamics and Host Influences, International Symposium of IUFRO, Kanazawa, Japan, 14–19 September 2003; pp. 16–18.
- Kenis, M.; Wermelinger, B.; Gregoire, J.C. Research on parasitoids and predators of Scolytidae—A review. In *Bark and Wood Insects in Living Trees in Europe, a Synthesis*; Lieutuer, F., Day, K.R., Battisti, A., Gregoire., J.C., Evans, H.E., Eds.; Springer: Dordrecht, The Netherlands, 2007; pp. 237–290.
- 97. Ranius, T.; Jansson, N. The influence of forest regrowth, original canopy cover and tree size on saproxylic beetles associated with old oaks. *Biol. Conserv.* 2000, 95, 85–94. [CrossRef]
- 98. Dervišević, M.; Graora, D. The life cycle and efficacy of *Anthribus nebulosus* Forster. in reducing soft scale populations in Belgrade. *Fresenius Environ. Bull.* **2019**, *28*, 1981–1985.
- 99. Lynikienė, J.; Tamutis, V.; Gedminas, A.; Marčiulynas, A.; Menkis, A. First Report of the Larch Longhorn (*Tetropium gabrieli* Weise, Coleoptera: Cerambycidae: Spondylidinae) on *Larix* sp. in Lithuania. *Insects* **2021**, *12*, 9–11. [CrossRef]
- 100. Speight, M.C.D. Saproxylic Invertebrates and Their Conservation; Council of Europe: Strasbourg, France, 1989; p. 78.
- 101. Muona, J. A revision of the Nearctic Eucnemidae. Acta Zool. Fenn. 2000, 212, 1-106.
- 102. Stokland, J.N.; Siitonen, J.; Jansson, B.G. *Biodiversity in Dead Wood*, 1st ed.; Cambridge University Press: Cambridge, UK, 2012; p. 521.
- 103. Seibold, S.; Bassler, C.; Baldrian, P.; Reinhard, L.; Thorn, S.; Ulyshen, M.; Weiss, I.; Muller, J. Dead-wood addition promotes non-saproxylic epigeal arthropods, but effects are mediated by canopy openness. *Biol. Conserv.* **2016**, 204, 181–188. [CrossRef]
- 104. Ulyshen, M.D.; Šobotník, J. An Introduction to the Diversity, Ecology, and Conservation of Saproxylic Insects. In *Saproxylic Insects*; Ulyshen, M., Ed.; Zoological Monographs; Springer: Cham, Switzerland, 2018; Volume 1. [CrossRef]
- 105. Albrecht, L. Die Bedeutung des toten Holzes im Wald. Forstwiss. Cent. Bl. 1991, 110, 106-113. [CrossRef]
- 106. Glück, E.; Spelda, J. Sukzession der Destruentencoenosen von Sturmwurfflächen. Veröffentlichungen Proj. Angew. Okol. **1996**, 16, 367–377.
- 107. Irmler, U.; Heller, K.; Warning, J. Age and tree species as factors influencing the populations of insects living in dead wood (Coleoptera, Diptera: Sciaridae, Mycetophilidae). *Pedobiologia* **1996**, *40*, 134–148.
- 108. Økland, B.; Bakke, A.; Hågvar, S.; Kvamme, T. What factors influence the diversity of saproxylic beetles? A multi-scaled study from a spruce forest in southern Norway. *Biodivers Conserv.* **1996**, *5*, 75–100. [CrossRef]
- Köhler, F. Totholzkäfer in Naturwaldzellen des nördlichen Rheinlands. LÖBF-Schr.reihe Landesanst. Ökol. Bodenordn. Forsten Nord. Westfal. 2000, 18, 352. Available online: https://katalog.slub-dresd (accessed on 10 February 2022).
- 110. Schiegg, K. Effects of dead wood volume and connectivity on saproxylic insect species diversity. *Ecoscience* 2000, 7, 290–298. [CrossRef]
- Balfour, H.H.; Edelman, J.C.K.; Cook, F.E.; Barton, W.I.; Buzicky, A.W.; Siem, R.A.; Bauer, H. Isolates of *California encephalitis* (La Crosse) virus from field-collected eggs and larvae of *Aedes triseriatus*: Identification of the overwintering site of California encephalitis. *J. Infect. Dis.* 1975, 131, 712–716. [CrossRef]
- 112. Pekár, S. Some observations on overwintering of spiders (Araneae) in two contrasting orchards in the Czech Republic. *Agr. Ecosyst. Environ.* **1999**, *73*, 205–210. [CrossRef]
- 113. Jonsell, M.; Weslien, J. Felled or standing retained wood—It makes a difference for saproxylic beetles. *For. Ecol. Manag.* **2003**, 175, 425–435. [CrossRef]
- 114. Grove, S.L. Saproxylic insect ecology and the sustainable management of forests. Ann. Rev. Ecol. Syst. 2002, 33, 1–23. [CrossRef]
- 115. Langor, D.; Hammond, H.; Spence, J.; Jacobs, J.; Cobb, T. Saproxylic insect assemblages in Canadian forests: Diversity, ecology, and conservation. *Can. Entomol.* **2008**, 140, 453–474. [CrossRef]

- Lindgren, B.S. *Trypodendron lineatum* (Olivier) (Coleoptera: Scolytidae) breeding in bigleaf maple. *Acer. Macrophyllum. J. Entomol. Soc. B. C.* 1986, *83*, 44. Available online: https://agris.fao.org/agris-search/search.do?recordID=US201302689267 (accessed on 10 February 2022).
- 117. Hanson, H.S. Ecological Notes on the Sirex Wood Wasps and their Parasites. Bull. Entomol. Res. 1939, 30, 27-65. [CrossRef]
- 118. Schnaider, Z. Atlas Uszkodzen Drzew i Krzewow Powodowanych Przez Owady i Pajęczaki; Panstwowe Wydawnictwo Naukowe: Warszawa, Poland, 1976; p. 74.
- 119. Unger, A.; Schniewind, A.P.; Unger, W. Conservation of Wood Artifacts: A Handbook; Herrmann, B., Ed.; Springer: Berlin/Heidelberg, Germany; New York, NY, USA, 1998; pp. 80–81.
- 120. Simila, M.; Kouki, J.; Martikainen, P. Saproxylic beetles in managed and seminatural Scots pine forests: Quality of dead wood matters. *For. Ecol. Manag.* 2003, 174, 365–381. [CrossRef]
- 121. Weslien, J.; Schroeder, L.M. Population levels of bark beetles and associate insects in managed and unmanaged spruce stand. *For. Ecol. Manag.* **1999**, *115*, 267–275. [CrossRef]
- 122. Jankowiak, R.; Rossa, R.; Miśta, K. Survey of fungal species vectored by *Ips cembrae* to European larch trees in Raciborskie forests (Poland). *Czech Mycol.* 2007, 59, 227–239. [CrossRef]
- 123. Wagner, M.R.; Clancy, K.M.; Lieutier, F.D.P.T. *Mechanisms and Deployment of Resistance in Trees to Insects*; Kluwer Academic: Dordrecht, The Netherlands, 2002; p. 339.
- 124. Müller, J.; Wende, B.; Strobl, C.; Eugster, M.; Gallenberger, I.; Floren, A.; Gossner, M.M. Forest management and regional tree composition drive the host preference of saproxylic beetle communities. *J. Appl. Ecol.* **2015**, *52*, 753–762. [CrossRef]
- Marmor, L.; Tõrra, T.; Saag, L.; Randlane, T. Species Richness of Epiphytic Lichens in Coniferous Forests: The Effect of Canopy Openness. Ann. Bot. Fenn. 2012, 49, 352–358. [CrossRef]
- 126. Green, G.A.; Nash, T.H.; Lange, O.L. Physiological ecology of carbon dioxide exchange. In *Lichen Biology*; Nash, T.H., Ed.; Cambridge University Press: Cambridge, UK, 2008; pp. 152–181.
- 127. Nascimbene, J.; Marini, L.; Motta, R.; Nimis, P.L. Influence of tree age, tree size and crown structure on lichen communities in mature Alpine spruce forests. *Biodivers Conserv.* **2009**, *18*, 1509–1522. [CrossRef]
- 128. Nascimbene, J.; Marini, L.; Nimis, P.L. Epiphytic lichen diversity in old-growth and managed *Picea abies* stands in Alpine spruce forests. *For. Ecol. Manag.* 2010, 260, 603–609. [CrossRef]
- 129. Jüriado, I.; Liira, J.; Paal, J.; Suija, A. Tree and stand level variables influencing diversity of lichens on temperate broad-leaved trees in boreo-nemoral floodplain forests. *Biodivers Conserv.* **2009**, *18*, 105–125. [CrossRef]
- 130. Király, I.; Nascimbene, J.; Tinya, F.; Ódor, P. Factors influencing epiphytic bryophyte and lichen species richness at different spatial scales in managed temperate forests. *Biodivers Conserv.* **2013**, *22*, 209–223. [CrossRef]
- 131. Fritz, Ö.; Heilmann-Clausen, J. Rot holes create key microhabitats for epiphytic lichens and bryophytes on beech (*Fagus sylvatica*). *Biol. Conserv.* **2010**, *143*, 1008–1016. [CrossRef]
- 132. Hauck, M.; Javkhlan, S. Epiphytic lichen diversity and its dependence on bark chemistry in the northern Mongolian dark taiga. *Flora Morphol. Distrib. Funct. Ecol. Plants* 2009, 204, 278–288. [CrossRef]
- 133. Hauck, M.; Spribille, T. The significance of precipitation and substrate chemistry for epiphytic lichen diversity in spruce-fir forests of the Salish Mountains, northwestern Montana. *Flora Morphol. Distrib. Funct. Ecol. Plants* **2005**, *6*, 547–562. [CrossRef]
- 134. Schmull, M.; Hauck, M.; Vann, D.R.; Johnson, A.H.; Runge, M. Site factors determining epiphytic lichen distribution in a dieback-affected spruce-fir forest on Whiteface Mountain, New York: Stemflow chemistry. *Can. J. Bot.* 2002, *80*, 1131–1140. [CrossRef]