



## Research article

# Assessment of rice genotypes through the lens of morpho-physiological and biochemical traits in response to arsenic stress

Sanaullah Jalil<sup>a</sup>, Muhammad Mudassir Nazir<sup>b</sup>, Mohamed A. Eweda<sup>a</sup>,  
Faisal Zulfiqar<sup>c,\*</sup>, Hayssam M. Ali<sup>d</sup>, Jean Wan Hong Yong<sup>e,\*\*</sup>, Xiaoli Jin<sup>a,\*\*\*</sup>

<sup>a</sup> The Advanced Seed Institute, Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou 310058, China

<sup>b</sup> School of Environment and Safety Engineering, Jiangsu University, Zhenjiang, 212013, China

<sup>c</sup> Department of Horticultural Sciences, Faculty of Agriculture and Environment, The Islamia University of Bahawalpur, Bahawalpur, 63100, Pakistan

<sup>d</sup> Department of Botany and Microbiology, College of Science, King Saud University, Riyadh, 11451, Saudi Arabia

<sup>e</sup> Department of Biosystems and Technology, Swedish University of Agricultural Sciences, 23456, Alnarp, Sweden



## ARTICLE INFO

## Keywords:

Antioxidants  
Arsenic  
Catalase  
Malondialdehyde  
Rice  
ROS

## ABSTRACT

Rice is a globally important food crop which is sensitive to the presence of a metalloid, arsenic (As). There is limited research pertaining to identifying relevant As-tolerant rice germplasm in adaptive breeding research initiatives, despite the fact that As contamination in rice has long been known. This study served to identify the growth performance of different rice genotypes under high levels of As. Rice seed germination analysis (germination percentage, GP) was performed to categorize the eight different rice genotypes and growing under varying As levels including As<sub>25</sub>, 25 μM and As<sub>50</sub>, 50 μM. The Zhenong 41 was identified as the highly tolerant genotypes with lowest decrease in GP by 87 %, plant height (PH) by 26 %, and dry weight (DW) by 16 %; while 9311 was observed to be the most sensitive genotype with highest reduction in GP by 44 %, PH by 48 % and DW by 54 % under As<sub>25</sub> stress conditions, compared to control treatment. The higher As<sub>50</sub> stress treatment delivered more adverse growth inhibitory effects than the rice plants cultivated under As<sub>25</sub>. Specifically, the As-sensitive rice genotype 9311 showed significantly higher decrease in foliar chlorophyll contents relative to the other genotypes, especially Zhenong 41 (As-tolerant). During exposure to high As levels, the rice genotype 9311 significantly modulated and augmented the production of MDA and H<sub>2</sub>O<sub>2</sub> by stimulating the activities of POD, SOD, and CAT. This study revealed interesting insights into the responses of rice genotypes to variable As stresses throughout the various growth stages. Overall, the findings of this study could be harnessed to support any ongoing As-tolerant rice breeding agendas for cultivation in As-polluted environments.

\* Corresponding author.

\*\* Corresponding author.

\*\*\* Corresponding author.

E-mail addresses: [ch.faisal.zulfiqar@gmail.com](mailto:ch.faisal.zulfiqar@gmail.com) (F. Zulfiqar), [jean.yong@slu.se](mailto:jean.yong@slu.se) (J.W.H. Yong), [jinxl@zju.edu.cn](mailto:jinxl@zju.edu.cn) (X. Jin).

<https://doi.org/10.1016/j.heliyon.2024.e36093>

Received 16 December 2023; Received in revised form 4 August 2024; Accepted 9 August 2024

Available online 10 August 2024

2405-8440/© 2024 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Rice, is the most extensively consumed grains worldwide, functions as a dietary staple, particularly in Asian countries where it is a fundamental component of daily meals [1]. It serves as a pivotal source of carbohydrates, energy, and vital nutrients, nourishing billions of people every day [2]. Therefore, in anticipation of Asia's projected population growth by 2050, a substantial surge of 60%–70% in rice production is imperative to meet the escalating demands, grain supply reliability, grain safety, and with minimal impact to the environment [3,69–71]. Heavy metals and metalloids are known environmental pollutants, and their toxicity is of increasing significance for agricultural, ecological, evolutionary, nutritional and environmental reasons [56–65]. Despite the nutritional significance and extensive utilization of rice, arsenic (As) contamination has emerged as recent alarming issue [4,70,71]. The element Arsenic (As) is a poisonous metal/metalloid that is widely recognized as a carcinogen affecting humans [5,55]. Anthropogenic activities such as sewage sludge, industrial waste, mining and the use of As-containing agrochemicals contribute to As contamination in rice-growing regions. The anthropogenic sources exacerbate the problem, leading to enhanced accumulation levels of As in rice grains [6,70,71]. Moreover, exposure to As-contaminated rice poses various health issues to individuals, including: cardiovascular diseases, leukaemia, skin lesions, diabetes, lung cancer, kidney cancer, and reproductive abnormalities like premature birth [7]. In addition, many rice growing areas have very high levels of As in the groundwater; the WHO's recommended level of As was established at 10 ppb (parts per billion) for drinking water [8,55]. However, due to the lack of policy guidelines and environmental chemistry technical limitations in many developing countries, there is no formal guidelines for As levels in irrigation water. To lower the levels of As in grains and straws below the safe levels for ingestion by humans and animals, one of the plausible option was to develop As tolerant rice varieties/genotypes. The ill effects of As have been extensively documented, revealing its ability to significantly impede metabolic functions, hinder development, and inhibit growth in plants [6,7]. Rice, barley, maize and wheat grains serves as primary source of As ingestion for humans. As major part of the population relies on rice food, its higher tendency to accumulate As, especially in the grains, makes it a major contributor to the incorporation of inorganic As into the human food system compared to other grain crops [8]. By 2050, the population is anticipated to steadily increase to 9.7 billion people, creating a pressing need for a 50% increase in crop yield production compared to current levels [5–7]. Rice, as the essential staple food worldwide, is a major contributor in the livelihoods of millions of people, particularly in Asian nations, where it constitutes 90% of both production and consumption. Specifically, the As accumulation in paddy soils and rice cultivation in polluted waters will ultimately jeopardize the health of people and animals.

The presence manifestation of As in the irrigation water and topsoil cause diverse hazardous impacts on rice plant, often compounded by additional soil-related challenges inherent to rice cultivation and such problems frequently congregated with other soil-related issues in rice farming [9]. The occurrence of As in rice grown fields triggers several detrimental responses, such as deprived germination, poor roots establishment, reduced photosynthesis activity, decreased biomass, leaf discoloration and stunted plant growth [10]. The impact of As toxicity on rice plants depends on its chemical structure [11]. While, the incorporation and translocation of inorganic As species are highly relatively effective into rice plant system [12]. Among the inorganic As sources, the more hazardous trivalent arsenite AsIII and the less poisonous pentavalent arsenate AsV are the main species found in water and soils [13]. Arsenite AsIII dominates in anaerobic lowland submerged paddy soils, while arsenate AsV prevails in aerobic rainfed upland paddy soils [14, 15]. The strong affinity of AsIII(III) for sulfhydryl-containing enzymes negatively affects the functionality of important enzymes in plants [16]. Despite the apparent scrutiny on the interaction between rice and As for the past two decades, there remains limited knowledge about the most As-resistant cultivars, genotypic variants, and screening techniques. At present, there is no known As-excluding donor germplasm; or useful As-related germplasm utilized in breeding programmes to develop As-resistant rice cultivars. In regions heavily affected by As contamination, many farmers face extreme poverty and lack the resources to afford expensive mitigation techniques or alternative food sources [16]. Furthermore, exposure to As stress leads to phytotoxic effects on rice plants through escalating electrolyte leakage, and membrane damage levels [17].

During an exposure to high As level by plants, there will be a biochemical burst of reactive oxygen species (ROS); consequently suppressing the antioxidant defence system. To counteract the effects of ROS during high As exposure, rice plants will employ the antioxidant defence mechanisms to cope with the perturbation induced by the ROS [18]. In addition, various antioxidative components also contribute to the neutralization of ROS generated under As toxicity [19]. Under stress scenarios, the antioxidant enzymes activities repressed due to the imbalances in enzyme functioning, the glyoxalase system and biomolecules. This biochemical imbalance within the tissues ultimately results in excessive oxidative stress [20]. Despite significant attention on the interaction between rice and As in the past two decades, there remains limited knowledge about the most resistant cultivars, genotypic variants, and screening techniques. At present, there is no known As-excluding donor germplasm has been identified or utilized in breeding efforts to develop resistant rice cultivars. In regions heavily affected by As contamination, many farmers face environmental challenges, extreme poverty and limited resources to cope with As-related pollution [21].

We selected and assessed the eight rice genotypes growing under different levels of As stress in order to comprehend the variation in genetic makeup concerning germination ability, alteration in photosynthetic parameters, growth related traits, and the modulation of their antioxidant and ROS system. Based on our hypothesis, the selected rice genotypes were expected to exhibit a wide range of morphological trait diversity under As-induced toxicity. The objectives were as follows: (a) to determine the germination potential of selected genotypes under various levels of As stress; (b) to investigate the genetic susceptibility/tolerance behaviour of selected rice genotypes against As toxicity; and (c) to identify the most significant As-sensitive genotype. The identified As-sensitive genotype could be used for future research pertaining to developing mitigation strategies against high levels of As.

## 2. Plant materials and methods

### 2.1. Plant materials and study layout

Eight different rice genotypes: *Oryza sativa* subsp. japonica (Nipponbare); 7 *Oryza sativa* subsp. indica rice (9311, Guiyin 206, Minghui 63, Zhenong 34, Zhenong 41, Zhenong 37 and Shenghai 1), were selected to assess their responses to As-induced toxicity. During the year of 2023, the genotypes were carefully selected and the whole experiment was conducted using a Randomized Complete Block Design (RCBD) at the Jin's laboratory, College of Agriculture and Biotechnology, Zhejiang University, Hangzhou, China; three replication of each treatment for the entire study.

### 2.2. Seed germination analysis

Seed germination analysis was done at 25 °C in a growth chamber to observe the germination ability of all selected genotypes under varied levels of As stress. Surface sterilize the rice seeds by soaking them in a 2 % H<sub>2</sub>O<sub>2</sub> (v/v) solution for 30 min. Seed rinsing was again done with distilled water. The seeds of each genotype were germinated in hydroponic growth medium within the boxes of 12 cm × 18 cm size; consisted 30 seeds of each genotype. Three treatments of inorganic As, 0 μM (control, CK), 25 μM (As<sub>25</sub>) and 50 μM (As<sub>50</sub>) were applied in the form of sodium arsenate dibasic heptahydrate (AsH<sub>15</sub>Na<sub>2</sub>O<sub>11</sub>) to determine the germination ability. Three replicates were prepared for each genotype and the respective As concentrations, in order to ensure statistical validity. After 10 days of seed sowing, the number of germinated seeds was noted. The germination percentage was evaluated by dividing the germinated seeds numbers by the total sowed seeds and multiplying by 100, in all treatments. After examining the germination percentages, the genotypes were organised into four groups including: highly tolerant >80 %, moderately tolerant >50 %, moderately susceptible <50 %; and the highly susceptible <20 %; following the formula of Murugaiyan et al. [22] for the As tolerance index (germination %).

### 2.3. Plant growth conditions for seedling stage screening

The germinated seeds of all the selected genotypes were subsequently transferred and grown into plastic pots under hydroponic conditions to evaluate the response of genotypes under different As concentrations (0, 25 and 50 μM) at seedling stage of rice plants. The pots were then adjusted in a growth chamber having 8 h dark and 16 h light photoperiod along with 22 °C for dark photoperiod and 28 °C for light photoperiod, light intensity (250 μmol, m<sup>-2</sup> s<sup>-1</sup>) and relative humidity of 70 %. During the preliminary 7 d after transplantation, the pots were irrigated with half-strength hydroponic nutrient solution (HNS). Following which, they were switched to full-strength HNS for the next 14 d during the experiment [11].

### 2.4. SPAD value and plant growth analysis

The relative foliar chlorophyll contents (SPAD values) were noted with a portable instrument (SPAD-502<sup>+</sup>, Tokyo, Japan) 7 d after As treatment [67]. Plant sample collection for the other analysis was also done on the same day. The roots of seedlings were washed with double distilled water (ddH<sub>2</sub>O). Plant height (Ph) was measured with a ruler while an analytical balance was employed to determine the dry weight of rice seedlings following Jalil et al. [11]. Furthermore, a subgroup of these samples was frozen using liquid nitrogen and then stored at -80 °C [23,24].

### 2.5. Determination of arsenic contents

The roots of rice seedling were cleaned with ddH<sub>2</sub>O. Then they were soaked in EDTA (20 mM) at 25 °C for 30 min in order to remove adhered metal ions on the root surface. Later, the root samples were oven-dried at 70 °C for 48 h. The digestion of the dried samples was done following the Nazir et al. [24]. After digestion, the samples were diluted in ddH<sub>2</sub>O and final volume (10 mL) was made. For As contents quantification, the digested samples were in an atomic absorption spectrophotometer (AA6300, Shimadzu, Japan) [25].

### 2.6. Measurement of malondialdehyde and hydrogen peroxide contents

The malondialdehyde (MDA) contents from the shoot samples were determined following the methods of Morales and Munné-Bosch [26] and while H<sub>2</sub>O<sub>2</sub> contents were determined by described methods of Srivastava et al. [27]. The details of protocols have been presented in Jalil et al. [11] and with some modifications [68].

### 2.7. Measurement of antioxidant enzyme activities

The sample (0.5 g) from shoots was taken and the extracts were made following the protocols detailed described in our previous study by Jalil et al. [11]. SOD enzyme activity was measured at 560 nm following the protocol of Giannopolitis and Ries [28]. POD activity was measured following the methodology of Maehly [29] at 470 nm. Moreover, CAT activity was recorded at 240 nm following Aebi [30].

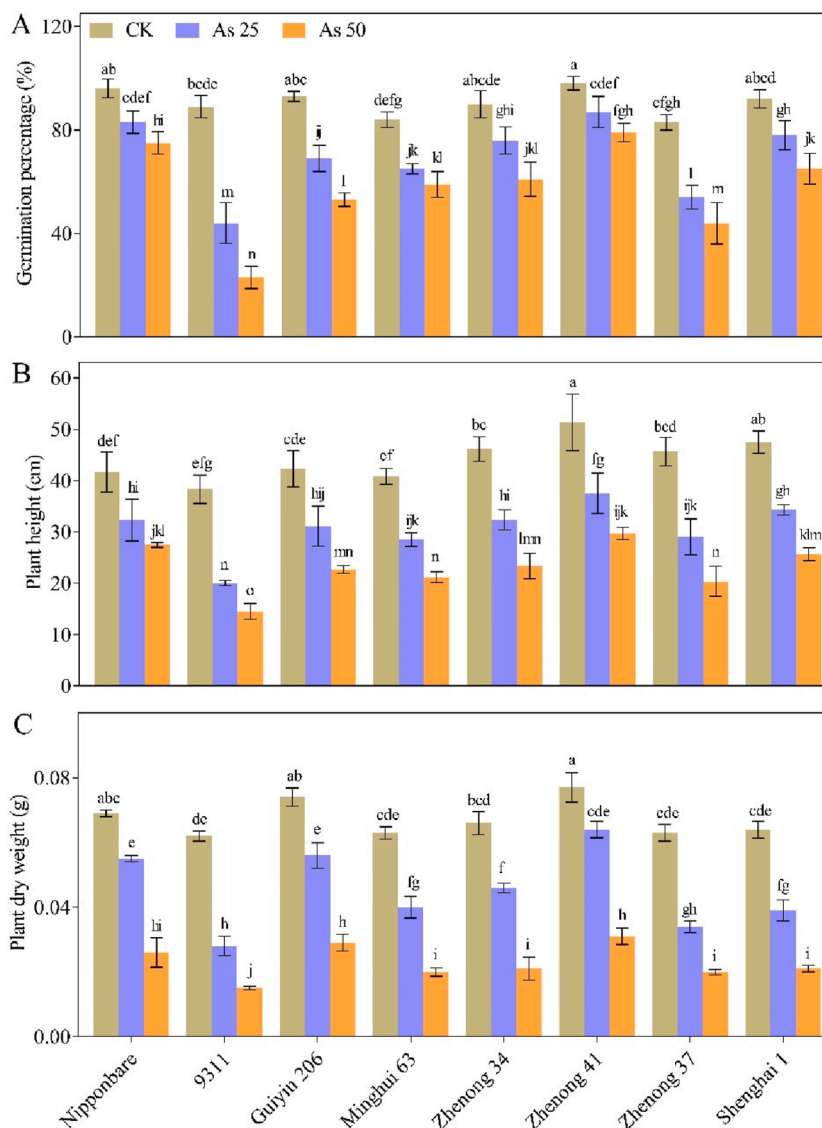
## 2.8. Statistical analysis

Statistical analysis was undertaken using Statistix 10. The data on the graphs were shown as the mean and standard error (SE) of three replicates; these were plotted later using Graph Pad Prism (8.0.2). Data were exposed to two way analysis of variance and significant means were divided using LSD at 5 % level of probability.

## 3. Results

### 3.1. Effects of arsenic on seed germination

The different rice genotypes response varied under altered As stress conditions, significantly affecting their germination capacity. When the concentrations of As were increased, the germination percentages (GP) of the genotypes showed a marked reduction. Under control conditions (without As), all eight genotypes demonstrated a high GP, with the Zhenong 41 genotype exhibiting the highest germination rate of 98 %. However, the impact of varying As concentrations on GP revealed more pronounced effects across the genotypes (Fig. 1A). However, the better response of genotypes to As stress was showed under the As<sub>25</sub> treatment compared with the



**Fig. 1.** Effects of arsenic (As) treatments on the (A) seed germination and (BC) plant growth traits of various rice genotypes. Vertical bars depict the mean  $\pm$  SD of three replicates. Variation in the above bar letters highlights significant difference at  $p \leq 0.05$ . Treatment details include: As<sub>50</sub> as 50  $\mu\text{M L}^{-1}$ ; As<sub>25</sub> as As 25  $\mu\text{M L}^{-1}$ ; and CK as control.

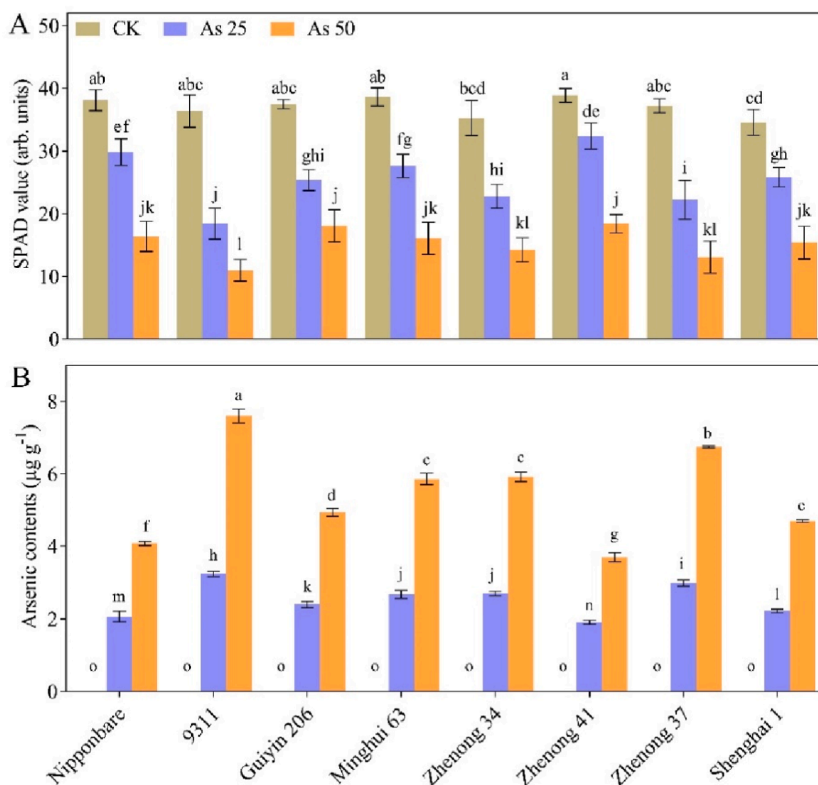
As<sub>50</sub> treatment, as observed in seed germination analysis. Specifically, the results of As<sub>25</sub> treatment demonstrated that Nipponbare and Zhenong 41 genotypes exhibited germination rates of 83 % and 87 %, respectively, which is more than 80 % and categorized as highly tolerant genotypes. Conversely, the genotype 9311 displayed a GP of 44 % and was classified as moderately susceptible genotype. Furthermore, the Guiyin 206, Minghui 63, Zhenong 34, Zhenong 37 and Shenghai 1 genotypes exhibited GP ranging between 50 and 80 %, placing them in moderately tolerant category. Interestingly, all genotypes represented a substantial decline in the GP under As<sub>50</sub> treatment, even the tolerant genotypes (Nipponbare and Zhenong 41), which showed moderate tolerance against the As stress. Specifically, Zhenong 37 was re-classified from the moderately tolerant category to the moderately susceptible category with a GP of 41 %. These findings indicated that the As<sub>50</sub> treatment was more hazardous for plants of all genotypes than the As<sub>25</sub> treatment.

### 3.2. Plant growth parameters

The exposure of all eight genotypes to different concentrations of As for a duration of 21 days resulted in the substantial reduction in plant height (PH) and dry weight (DW), compared with the plants grown under control conditions. The genotype 9311 exhibited the most substantial reduction, with a decrease of 48 % and 62 % in PH, and 54 % and 75 % in DW under the As<sub>25</sub> and As<sub>50</sub> treatments, respectively, than control (CK). In contrast, the least significant decrease in PH by 22 % and 34 % was observed in Nipponbare, while substantial reduction in DW by 16 % and 60 %, respectively was observed in Zhenong 41 under As<sub>25</sub> and As<sub>50</sub> treatments, compared with CK treatment (Fig. 1B and C). Other genotypes also displayed variable responses to growth parameters with suppression of PH (ranging from 26 % to 36 %) and DW (ranging from 20 % to 40 %) under As<sub>25</sub> stress conditions. Overall, our findings indicated that the As<sub>50</sub> treatment delivered more negative effects on the growth parameters compared to the As<sub>25</sub> treatment.

### 3.3. Chlorophyll content and As contents

We observed a reduction in the leaf SPAD value due to As stress, however, no significant differences were found among genotypes under control conditions. The genotype 9311 displayed the most significant reduction in SPAD values, experiencing a substantial decrease of 49 % and 70 % under the As<sub>25</sub> and As<sub>50</sub> treatments, respectively, compared to the CK treatment. Following closely, genotype Zhenong 37 exhibited reductions of 40 % and 65 % under the same and As<sub>50</sub> treatments, respectively. (Fig. 2A), indicating their high sensitivity to As stress. Other genotypes showed modest reductions in SPAD value compared to mentioned genotypes. For



**Fig. 2.** Effects of arsenic (As) treatments on the (A) SPAD value and (B) As contents of various rice genotypes. Vertical bars depict the mean + SD of three replicates. Variation in the above bar letters highlights significant difference at  $p \leq 0.05$ . Treatment details include: As<sub>50</sub> as As 50  $\mu\text{M L}^{-1}$ ; As<sub>25</sub> as As 25  $\mu\text{M L}^{-1}$ ; and CK as control.

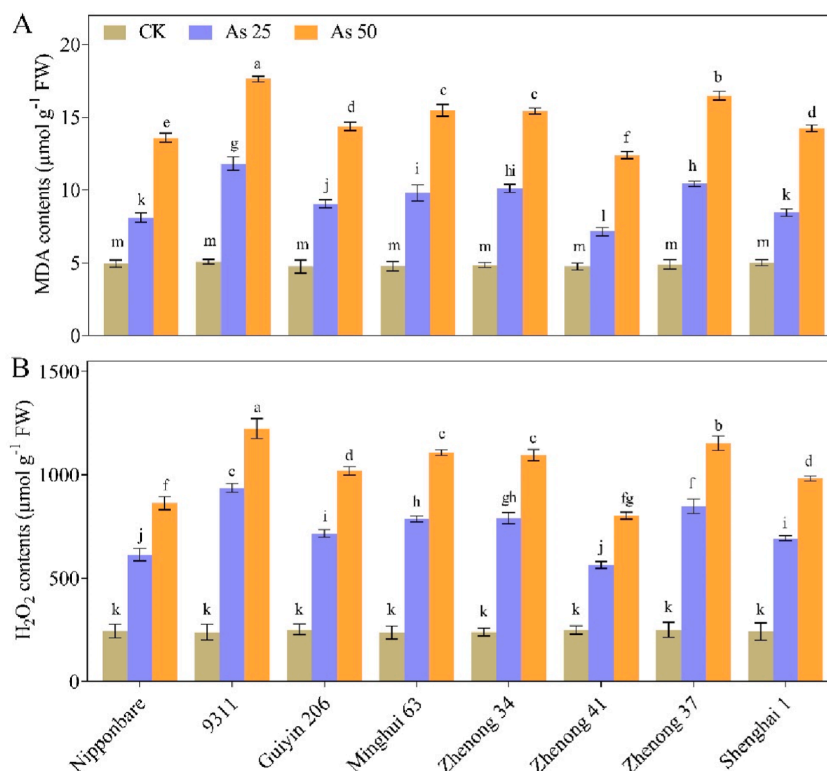
instance, Zhenong 41 showed a minimal decrease of 17 % in SPAD value under  $As_{25}$  treatment. However, under  $As_{50}$  stress conditions, there was a more pronounced toxic effect on SPAD values compared to the  $As_{25}$  stress conditions. Furthermore, we determined the accumulation of As in plants of all genotypes under  $As_{25}$  and  $As_{50}$  treatments. The results revealed that genotypes under  $As_{50}$  treatment contained higher concentration of As compared to those under  $As_{25}$  treatment (Fig. 2B). In addition, among all genotypes, 9311 accumulated a substantially higher amount of As, while Zhenong 41 accumulated a lower amount of As compared to other genotypes, indicating their sensitivity and tolerance, respectively to As stress conditions.

### 3.4. Quantification of MDA and $H_2O_2$ contents

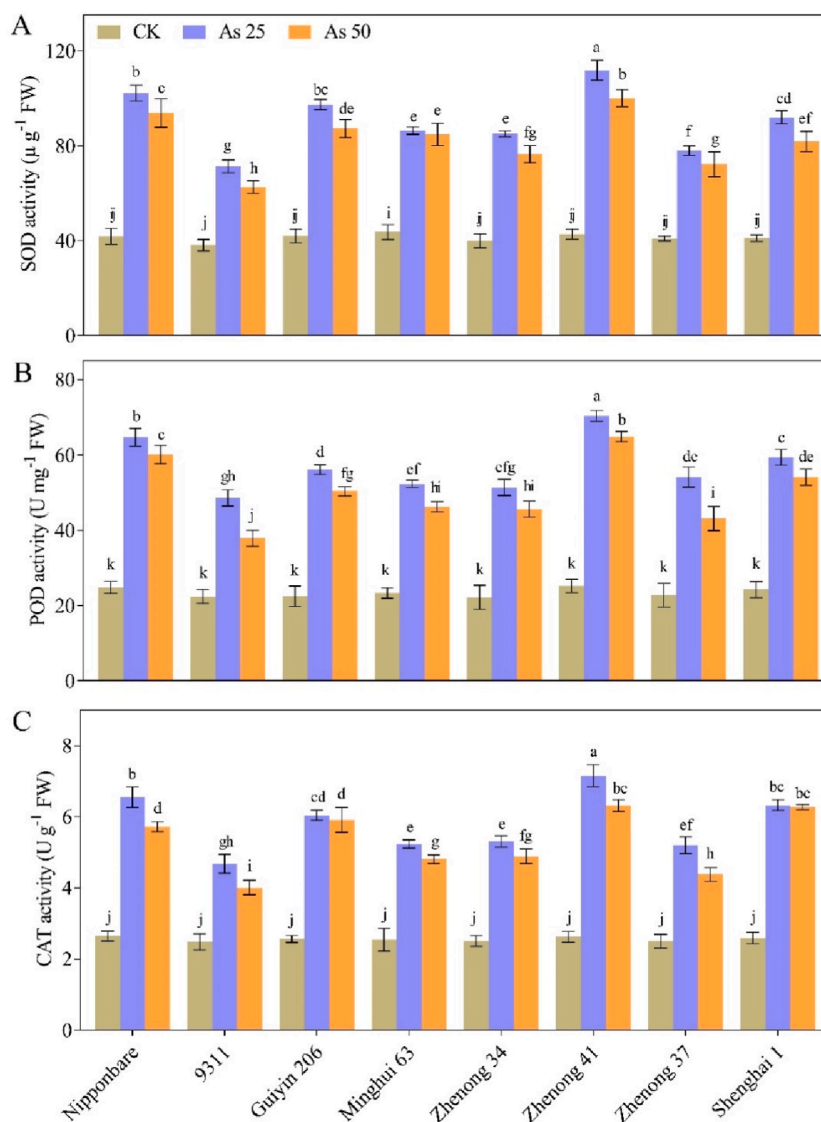
The values of MDA contents and  $H_2O_2$  production did not differ significantly from each other (Fig. 3A and B). However, when exposed to As stress, a considerable upsurge was observed in the values of MDA contents by 39 %, 57 %, 48 %, 51 %, 52 %, 34 %, 53 % and 41 % and  $H_2O_2$  contents by 60 %, 75 %, 65 %, 70 %, 69 %, 56 %, 71 % and 65 %, in Nipponbare, 9311, Guiyin 206, Minghui 63, Zhenong 34, Zhenong 41, Zhenong 37 and Shenghai 1, respectively under the  $As_{25}$  treatment. Similar trend was observed under  $As_{50}$  treatment, but with a more substantial increase in values compared with  $As_{25}$  treatment (Fig. 3A and B). Our results demonstrated that the values of MDA and  $H_2O_2$  significantly enhanced in the As-sensitive genotypes, while only slight increases were observed in the values of As tolerant genotypes, particularly in more sensitive genotype 9311 and more tolerant genotype Zhenong 41.

### 3.5. Antioxidant enzymes assays

All genotypes increased their antioxidant levels under As stress. Interestingly, these increases were more obvious in the values of SOD, POD and CAT activities within the high As tolerant rice genotype, Zhenong 41 with increase of 60 %, 66 % and 63 %, respectively under  $As_{25}$  stress conditions as compared with CK plants (Fig. 4A,B,C). In contrast, the SOD, POD and CAT activities in the As-sensitive rice genotype, 9311, were significantly lower, with increase of 47 %, 54 % and 45 %, respectively, compared with CK treatment. The other genotypes showed modest increase in the levels of antioxidant enzyme activities; and represented non-significant differences among each other. Additionally, all the genotypes showed similar trend under  $As_{50}$  stress conditions but with slightly reduction in SOD, POD and CAT values, comparison with  $As_{25}$  stress conditions. Moreover, no noteworthy change was found in enzyme activities in plants from all genotypes grown under the controlled conditions without As stress.



**Fig. 3.** Effects of arsenic (As) treatments on the (A) MDA (malondialdehyde), and (B)  $H_2O_2$  (hydrogen peroxide) content of various rice genotypes. Vertical bars depict the mean  $\pm$  SD of three replicates. Variation in the above bar letters highlights significant difference at  $p \leq 0.05$ . Treatment details include:  $As_{50}$  as As 50  $\mu\text{M L}^{-1}$ ;  $As_{25}$  as As 25  $\mu\text{M L}^{-1}$ ; and CK as control.



**Fig. 4.** Effects of arsenic (As) treatments on the (A) SOD (superoxide dismutase), (B) POD (peroxidase) and (C) CAT (catalase) activity of various rice genotypes. Vertical bars depict the mean  $\pm$  SD of three replicates. Variation in the above bar letters highlights significant difference at  $p \leq 0.05$ . Treatment details include:  $\text{As}_{50}$  as As 50  $\mu\text{M L}^{-1}$ ;  $\text{As}_{25}$  as As 25  $\mu\text{M L}^{-1}$ ; and CK as control.

#### 4. Discussion

Among all abiotic stress factors, the potential hazards posed by heavy metals and metalloids (HMs) are one of the main concerns regarding human and environmental health issues [31,32,70,71]. HMs have high reactivity due to their oxidative state and caused consequent toxicity to the majority of species [33]. The pollution caused by HMs has significantly impeded crop production in recent years. In addition, HMs severely jeopardizes the human food and nutrition, especially when it comes to rice [34]. The As is considered to be one of the most harmful HM and recognized as non-essential and hazardous HM due to its no essential physiological roles in plants [35,56,71]. The enhanced accumulation of As disturbs the plant development and growth, leading to substantial changes in the morphological and physiological functions of plants; specifically, the high As exposure in rice will cause the following: lower seed germination, poor seedling health, reduced photosynthesis, reduced biomass, and negative alteration of metabolic activities and ROS production. The toxicity of As also caused strait head disease in rice grown in soils with high As levels [10,17]. It is of utmost importance to take the appropriate action to address the issue of As toxicity in rice. Several research reports have explored the hazardous impacts of As toxicity in rice [27,36,37]. In line with these concerns, the findings revealed a reduction in seed germination and overall plant growth under As stress. The application of two different concentration of  $\text{As}_{25}$  and  $\text{As}_{50}$  led to a decrease in GP, DW and PH of rice seedlings (Fig. 1), corroborating the results of Murugaiyan et al. [22] and Kumar et al. [38]. Based on the GP ratios, we classified the studied genotypes into different As tolerance groups. Genotype 9311 was identified as a moderately susceptible genotype

(GP <50 %), while Nipponbare and Zhenong 41 with GP exceeding 80 %, were classified as highly tolerant genotypes. The remaining five genotypes exhibited moderate tolerance under the As<sub>25</sub> stress conditions (Fig. 1). Furthermore, it was observed that As<sub>50</sub> treatment resulted in more pronounced deleterious effects on GP of genotypes, indicating that higher concentrations of As severely impaired the growth of rice plants. This study aligned with previous research representing the detrimental effects of As stress on plant phenotypic parameters in crops such as soybean, canola, and barley [24,39]. Additionally, the reports on Cr toxicity in rice crops have also reported negative impacts on seed germination rates [40,65], which parallel the results obtained in our study under As toxicity.

Chlorophyll, an essential component of plants involved in the photosynthesis and an integral component in plant development and serves as a valuable indicator to find stress levels caused by HMs in crop plants [41,65,66]. In our study, we lower SPAD values of rice seedlings under As stress, and these reductions in SPAD value increased with higher As concentrations (Fig. 2). The decline in chlorophyll content was correlated with a decrease in plant biomass [42,67]. Consistent with previous experiments, we observed a positive link between chlorophyll contents and shoot development in rice plants. The high levels of metalloid As prevented the production of chlorophyll contents by interfering with the action of chlorophyll synthase. All the rice genotypes in our study were adversely affected by As stress, except for Zhenong 41, which exhibited high tolerance against As toxicity. In contrast, genotype 9311 showed increased susceptibility to As toxicity, resulting in more pronounced detrimental effects on SPAD value. Additionally, our findings revealed that the rice genotypes generally accumulated higher amount of As concentration under As<sub>50</sub> treatment than the As<sub>25</sub> treatment; and these higher As concentrations were associated with more negative effects on plant growth parameters compared to plants grown. These results highlighted the negative correlation between growth parameters and As accumulation, indicating that the rice growth parameters were suppressed in a dose-dependent manner which was consistent with the Mousavi et al. [43] and Wu et al. [44].

Antioxidants, both non-enzymatic and enzymatic, have a helpful role in the defense by working to balance ROS generated within different cellular compartments [45]. In this experiment, we observed that the exposure of rice genotypes to varying levels of As toxicity led to a burst of H<sub>2</sub>O<sub>2</sub> and MDA contents in rice plants, particularly in the 9311 genotype, which displayed a sensitive response to As compared to other genotypes (Fig. 3). This increase in ROS production and MDA contents indicates the damage induced by As toxicity, leading to membrane degradation, lipid peroxidation, and enhanced oxidative damage to cells. These results were in agreement with the previous reports that have demonstrated similar As-induced oxidative indicators in barley, maize and wheat etc. [24,46,47]. In contrast, Zhenong 41 showed a low amount of MDA contents and H<sub>2</sub>O<sub>2</sub> production, indicating its high tolerance towards the As phytotoxic effects. It was plausible that the Zhenong 41 genotype possessed an efficient As-detoxification system, which reduced oxidative stress and enhanced the capacity for photosynthetic activity, leading to improved growth and biomass of rice seedlings. Gupta and Ahmad [48] had previously described in their study about differential responses of various rice genotypes to As toxicity, thereby supporting our findings. Our results also aligned with the observations of Khan et al. [49], whom reported excessive lipid peroxidation and ROS formation in As-sensitive rice cultivars growth under high As levels. Thus, the inhibitory role of higher As concentrations on the biology of rice plants health could be attributed to oxidative injury to the tissues [50]. Earlier studies have also supported the notion that plants develop mechanisms to promote cellular metabolism and to eliminate the production of free radicals; subsequently lowering the ROS levels and MDA contents [50,51].

To mitigate the damaging effects of oxidative injury due to higher As levels, plants have developed a specific mechanism known as the antioxidant dependant defence process [52,63,65,68]. Our study investigated how different rice genotypes remarkably boosted the activities of antioxidative enzymes, to combat As based oxidative injury (Fig. 4). It was observed that rice plants activated the antioxidant defence system to eliminate excessive ROS production within their cells, consequently reducing MDA levels. In our study, we observed a substantial increase in H<sub>2</sub>O<sub>2</sub> production and MDA contents with higher concentrations of As toxicity, indicating a positive correlation between these variables and As accumulation. Several studies have also reported similar findings in rice, wheat, and maize plants, demonstrating the activation of these antioxidative indicators [47,53,54]. Higher antioxidant enzyme activities were found in the As-tolerant rice genotypes, particularly Zhenong 41, compared to the As-sensitive genotype 9311; indicating a compromised response to As stress in the latter. It is possible that the As-sensitive rice genotype fails to maintain the redox balance, leading to excessive ROS accumulation within the cellular components under As stress. Therefore, the proposed method described a plausible approach for mitigating As stress in rice plants, particularly in As-sensitive cultivars such as 9311, which could be further selected in future research.

## 5. Conclusion

The study examined the effects of As, a non-essential and toxic HM, on rice genotypes. The study revealed that high As levels reduced seed germination, SPAD value, growth, and biomass of rice seedlings. Specifically, the higher As concentrations exacerbated the negative growth effects. The study also highlighted the complex interplay between oxidative stress and As toxicity. Rice genotypes responded to high As levels by activating antioxidant defense mechanisms, as evidenced by the heightened antioxidative enzyme activities. Interestingly, the rice genotype-specific responses were evident, with Zhenong 41 demonstrating notable As tolerance through effective maintenance of redox balance and adaptations; while 9311 was considered a susceptible genotype to As stress. These insights provided by this study contributed to the broader understanding of rice responses to As toxicity and offered plausible solutions in addressing this critical environmental issue affecting food safety.

## Funding

This work was supported by the Science and Technology Office of Zhejiang Province, China (project no. 2021C02063-6). This work was further funded by the Researchers Supporting Project number (RSP2024R123), King Saud University, Riyadh, Saudi Arabia.



## Data availability statement

Data will be made available on request to corresponding authors.

## CRediT authorship contribution statement

**Sanaullah Jalil:** Writing – original draft, Validation, Methodology, Conceptualization. **Muhammad Mudassir Nazir:** Writing – review & editing. **Mohamed A. Eweda:** Writing – review & editing. **Faisal Zulfiqar:** Writing – review & editing, Validation, Formal analysis. **Hayssam M. Ali:** Writing – review & editing. **Jean Wan Hong Yong:** Writing – review & editing. **Xiaoli Jin:** Writing – review & editing, Supervision.

## Declaration of competing interest

The authors declared that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## Acknowledgment

Authors would like to extend their sincere appreciation to the Researchers Supporting Project number (RSP2024R123), King Saud University, Riyadh, Saudi Arabia.

## References

- [1] M. TatahMentan, S. Nyachoti, L. Scott, N. Phan, F.O. Okwori, N. Felemban, T.R. Godebo, Toxic and essential elements in rice and other grains from the United States and other countries, *Int. J. Environ. Res. Publ. Health* 17 (2020) 8128.
- [2] K. Kobayashi, X. Wang, W. Wang, Genetically modified rice is associated with hunger, health, and climate resilience, *Foods* 12 (2023) 2776.
- [3] S. Yuan, A.M. Stuart, A.G. Laborte, J.I. Rattalino Edreira, A. Dobermann, L.V.N. Kien, L.T. Thúy, K. Paothong, P. Traesang, K.M. Tint, Southeast Asia must narrow down the yield gap to continue to be a major rice bowl, *Nature Food* 3 (2022) 217–226.
- [4] M. Shri, S. Dubey, S. Dwivedi, P.K. Singh, R.D. Tripathi, Long-distance translocation mechanism of arsenic from soil to rice grain, in: *Arsenic in Rice*, Apple Academic Press, 2024, pp. 75–94.
- [5] S. Manawasinghe, R. Chandrajith, Growth responses of genetically diverse rice (*Oryza sativa* L.) cultivars to Arsenic stress, *Ceylon Journal of Science* 52 (2023) 381–389.
- [6] A. Majumdar, M.K. Upadhyay, B. Giri, P. Yadav, D. Moullick, S. Sarkar, T. Roychowdhury, Sustainable water management in rice cultivation reduces arsenic contamination, increases productivity, microbial molecular response, and profitability, *J. Hazard Mater.* 466 (2024) 133610.
- [7] K.M. Alwutayd, S.M.S. Alghanem, D. Alshehri, M.H. Saleem, S. Hussain, B. Ali, A.H. Abeer, Advancing arsenic toxicity mitigation in rice (*Oryza sativa* L.) with rice straw biochar and silicon: a study on morpho-physio-biochemical responses, *J. Soil Sci. Plant Nutr.* (2024) 1–15.
- [8] C.M. George, L. Sima, M.H.J. Arias, J. Mihalic, L.Z. Cabrera, D. Danz, W. Checkley, R.H. Gilman, Exposition à l'arsenic dans l'eau potable: Une menace méconnue pour la santé au Pérou, *Bull. World Health Organ.* 92 (2014) 565–572.
- [9] J. Li, Y. Chang, A.A. Al-Huqail, Z. Ding, M.S. Al-Harbi, E.F. Ali, S.A. Tammam, Effect of manure and compost on the phytostabilization potential of heavy metals by the halophytic plant wavy-leaved saltbush, *Plants* 10 (10) (2021) 2176.
- [10] X. Fang, I. Christl, A.E.C. Blanco, B. Planer-Friedrich, F.-J. Zhao, R. Kretzschmar, Decreasing arsenic in rice: interactions of soil sulfate amendment and water management, *Environ. Pollut.* 322 (2023) 121152.
- [11] S. Jalil, S.M. Alghanem, A.A. Al-Huqail, M.M. Nazir, F. Zulfiqar, T. Ahmed, S. Ali, A.H. Abeer, K.H. Siddique, X. Jin, Zinc oxide nanoparticles mitigated the arsenic induced oxidative stress through modulation of physio-biochemical aspects and nutritional ions homeostasis in rice (*Oryza sativa* L.), *Chemosphere* 338 (2023) 139566.
- [12] J. Kalita, A.K. Pradhan, S.Y. Jyoti, B. Kalita, S.J. Roy, I. Kalita, M. Das, H. Teronpi, P. Regon, B. Tanti, Translocation and accumulation of arsenic in rice grains: role of transporters in tolerance and its incorporation in the food chain, in: *Arsenic in Rice*, Apple Academic Press, 2024, pp. 119–142.
- [13] I. Khan, S.A. Awan, M. Rizwan, S. Ali, X. Zhang, L. Huang, Arsenic behavior in soil-plant system and its detoxification mechanisms in plants: a review, *Environ. Pollut.* 286 (2021) 117389.
- [14] M.K. Souiri, N. Alipanahi, M. Hatamian, M. Ahmadi, T. Tesfamariam, Elemental profile of heavy metals in garden cress, coriander, lettuce and spinach, commonly cultivated in Kahrizak, South of Tehran-Iran, *Open agriculture* 3 (2018) 32–37.
- [15] M.K. Souiri, M. Hatamian, T. Tesfamariam, Plant growth stage influences heavy metal accumulation in leafy vegetables of garden cress and sweet basil, *Chemical and Biological Technologies in Agriculture* 6 (2019) 1–7.
- [16] A.S. Maghsoudi, S. Hassani, K. Mirnia, M. Abdollahi, Recent advances in nanotechnology-based biosensors development for detection of arsenic, lead, mercury, and cadmium, *Int. J. Nanomed.* (2021) 803–832.
- [17] S. Samanta, A. Banerjee, A. Roychowdhury, Arsenic toxicity is counteracted by exogenous application of melatonin to different extents in arsenic-susceptible and arsenic-tolerant rice cultivars, *J. Plant Growth Regul.* 41 (2022) 2210–2231.
- [18] Y. Dong, M. Gao, Z. Song, W. Qiu, Microplastic particles increase arsenic toxicity to rice seedlings, *Environ. Pollut.* 259 (2020) 113892.
- [19] M.G. Mostofa, C.V. Ha, M. Rahman, K.H. Nguyen, S.S. Keya, Y. Watanabe, M. Itouga, A. Hashem, E.F. Abd Allah, M. Fujita, Strigolactones modulate cellular antioxidant defense mechanisms to mitigate arsenate toxicity in rice shoots, *Antioxidants* 10 (2021) 1815.
- [20] T. Ahmed, J. Guo, M. Noman, L. Lv, N. Manzoor, X. Qi, B. Li, Metagenomic and chemical analyses reveal the potential of silicon to alleviate arsenic toxicity in rice (*Oryza sativa* L.), *Environ. Pollut.* 345 (2024) 123537.
- [21] M.M. Metwaly, M.A. AbdelRahman, B. Abdellatif, Heavy metals and micronutrients assessment in soil and groundwater using geospatial analyses under agricultural exploitation in dry areas, *Acta Geophys.* (2022) 1–29.
- [22] V. Murugaiyan, J. Ali, M. Frei, F. Zeibig, A. Pandey, A. Wairich, L.-B. Wu, J. Murugaiyan, Z. Li, Identification of promising genotypes through systematic evaluation for arsenic tolerance and exclusion in rice (*Oryza sativa* L.), *Front. Plant Sci.* 12 (2021) 753063.
- [23] N. Masood, M.A. Irshad, R. Nawaz, T. Abbas, M.A. Abdel-Maksoud, W.H. AlQahtani, A.H. Abeer, Green synthesis, characterization and adsorption of chromium and cadmium from wastewater using cerium oxide nanoparticles; reaction kinetics study, *J. Mol. Struct.* 1294 (2023) 136563.
- [24] M.M. Nazir, Q. Li, M. Noman, S. Ullhassan, S. Ali, T. Ahmed, F. Zeng, G. Zhang, Calcium oxide nanoparticles have the role of alleviating arsenic toxicity of barley, *Front. Plant Sci.* 13 (2022).
- [25] S. Zhang, Q. Li, M.M. Nazir, S. Ali, Y. Ouyang, S. Ye, F. Zeng, Calcium plays a double-edged role in modulating cadmium uptake and translocation in rice, *Int. J. Mol. Sci.* 21 (2020) 8058.

- [26] M. Morales, S. Munné-Bosch, Malondialdehyde: facts and artifacts, *Plant Physiol.* 180 (2019) 1246–1250.
- [27] S. Srivastava, P. Suprasanna, R.D. Tripathi, Safeguarding rice from arsenic contamination through the adoption of chemo-agronomic measures, *Arsenic in Drinking Water and Food* (2020) 411–424. [https://doi.org/10.1007/978-981-13-8587-2\\_16](https://doi.org/10.1007/978-981-13-8587-2_16).
- [28] C.N. Giannopolitis, S.K. Ries, Superoxide dismutases: I. Occurrence in higher plants, *Plant Physiol.* 59 (1977) 309–314.
- [29] A. Maehly, The assay of catalases and peroxidases, *Methods Biochem. Anal.* (1954) 357–424.
- [30] H. Aebi, Catalase in vitro *Methods Enzymol* 105 (1984) 121–126.
- [31] M. Hatamian, A. Rezaei Nejad, M. Kafi, M.K. Souri, K. Shahbazi, Interaction of lead and cadmium on growth and leaf morphophysiological characteristics of European hackberry (*Celtis australis*) seedlings, *Chemical and Biological Technologies in Agriculture* 7 (2020) 1–8.
- [32] B. Fattahi, K. Arzani, M.K. Souri, M. Barzegar, Effects of cadmium and lead on seed germination, morphological traits, and essential oil composition of sweet basil (*Ocimum basilicum* L.), *Ind. Crop. Prod.* 138 (2019) 111584.
- [33] S. Yadav, P. Modi, A. Dave, A. Vijapura, D. Patel, M. Patel, Effect of abiotic stress on crops, *Sustainable crop production* 3 (2020).
- [34] V. Murugaiyan, J. Ali, A. Mahender, U.M. Aslam, Z.A. Jewel, Y. Pang, C.M. Marfori-Nazarea, L.-B. Wu, M. Frei, Z. Li, Mapping of genomic regions associated with arsenic toxicity stress in a backcross breeding populations of rice (*Oryza sativa* L.), *Rice* 12 (2019) 1–14.
- [35] M.A. Farooq, R.A. Gill, F. Islam, B. Ali, H. Liu, J. Xu, S. He, W. Zhou, Methyl jasmonate regulates antioxidant defense and suppresses arsenic uptake in Brassica napus L., *Front. Plant Sci.* 7 (2016) 468.
- [36] H.F. Bakhat, Z. Zia, S. Fahad, S. Abbas, H.M. Hammad, A.N. Shahzad, F. Abbas, H. Alharby, M. Shahid, Arsenic uptake, accumulation and toxicity in rice plants: possible remedies for its detoxification: a review, *Environ. Sci. Pollut. Control Ser.* 24 (2017) 9142–9158.
- [37] S.R. Mousavi, Y. Niknejad, H. Fallah, D.B. Tari, Methyl jasmonate alleviates arsenic toxicity in rice, *Plant Cell Rep.* 39 (2020) 1041–1060.
- [38] A. Kumar, S. Basu, G. Kumar, Evaluating the effect of seed-priming for improving arsenic tolerance in rice, *J. Plant Biochem. Biotechnol.* (2021) 1–5.
- [39] S. Sharma, G. Anand, N. Singh, R. Kapoor, Arbuscular mycorrhiza augments arsenic tolerance in wheat (*Triticum aestivum* L.) by strengthening antioxidant defense system and thiol metabolism, *Front. Plant Sci.* 8 (2017) 259098.
- [40] F. Basit, M.M. Nazir, M. Shahid, S. Abbas, M.T. Javed, T. Naqqash, Y. Liu, G. Yajing, Application of zinc oxide nanoparticles immobilizes the chromium uptake in rice plants by regulating the physiological, biochemical and cellular attributes, *Physiol. Mol. Biol. Plants* 28 (2022) 1175–1190.
- [41] H. Singh, D. Kumar, V. Soni, Performance of chlorophyll a fluorescence parameters in *Lemma minor* under heavy metal stress induced by various concentration of copper, *Sci. Rep.* 12 (2022) 10620.
- [42] A. Füzy, R. Kovács, I. Cseresnyés, I. Parádi, T. Szili-Kovács, B. Kelemen, K. Rajkai, T. Takács, Selection of plant physiological parameters to detect stress effects in pot experiments using principal component analysis, *Acta Physiol. Plant.* 41 (2019) 1–10.
- [43] S.R. Mousavi, Y. Niknejad, H. Fallah, D.B. Tari, Methyl jasmonate alleviates arsenic toxicity in rice, *Plant Cell Rep.* 39 (2020) 1041–1060.
- [44] F. Wu, Q. Fang, S. Yan, L. Pan, X. Tang, W. Ye, Effects of zinc oxide nanoparticles on arsenic stress in rice (*Oryza sativa* L.): germination, early growth, and arsenic uptake, *Environ. Sci. Pollut. Control Ser.* 27 (2020) 26974–26981.
- [45] M. Hasanuzzaman, M.R.H. Raihan, A.A.C. Masud, K. Rahman, F. Nowroz, M. Rahman, K. Nahar, M. Fujita, Regulation of reactive oxygen species and antioxidant defense in plants under salinity, *Int. J. Mol. Sci.* 22 (2021) 9326.
- [46] M. Saeed, U.M. Quraishi, R.N. Malik, Arsenic uptake and toxicity in wheat (*Triticum aestivum* L.): a review of multi-omics approaches to identify tolerance mechanisms, *Food Chem.* 355 (2021) 129607.
- [47] M. Kashif, A. Sattar, S. Ul-Allah, A. Sher, M. Ijaz, M. Butt, A. Qayyum, Silicon alleviates arsenic toxicity in maize seedlings by regulating physiological and antioxidant defense mechanisms, *J. Soil Sci. Plant Nutr.* 21 (2021) 2032–2040.
- [48] M. Gupta, M.A. Ahmad, Arsenate induced differential response in rice genotypes, *Ecotoxicol. Environ. Saf.* 107 (2014) 46–54.
- [49] Z. Khan, T.C. Thounaojam, R. Bhagawati, H. Upadhyaya, Impact of arsenic on the seedlings of Ranjit and Aijung, two most edible rice cultivars of Assam, India, *J. Stress Physiol. Biomed.* 18 (2022) 28–39.
- [50] M. Asgher, S. Ahmed, Z. Sehar, H. Gautam, S.G. Gandhi, N.A. Khan, Hydrogen peroxide modulates activity and expression of antioxidant enzymes and protects photosynthetic activity from arsenic damage in rice (*Oryza sativa* L.), *J. Hazard Mater.* 401 (2021) 123365.
- [51] H.-i. Jung, J. Lee, M.-J. Chae, M.-S. Kong, C.-H. Lee, S.-S. Kang, Y.-H. Kim, Growth-inhibition patterns and transfer-factor profiles in arsenic-stressed rice (*Oryza sativa* L.), *Environ. Monit. Assess.* 189 (2017) 1–11.
- [52] M.I.R. Khan, B. Jahan, M.F. AlAjmi, M.T. Rehman, N. Iqbal, M. Irfan, Z. Sehar, N.A. Khan, Crosstalk of plant growth regulators protects photosynthetic performance from arsenic damage by modulating defense systems in rice, *Ecotoxicol. Environ. Saf.* 222 (2021) 112535.
- [53] N.I. Elsheery, M.N. Helaly, S.F. El-Hefnawy, M.M. Elhamahmy, E.M. Abdelrazik, Y.B. Sardarov, P. Ahmad, M. Zivcak, M. Brestic, S.I. Allakhverdiev, 5-Aminolevulinic acid (ALA) reduces arsenic toxicity stress in wheat (*Triticum aestivum* L.), *J. Plant Growth Regul.* (2022) 1–20.
- [54] V. Anand, J. Kaur, S. Srivastava, V. Bist, V. Dharmesh, K. Kriti, S. Bisht, P.K. Srivastava, S. Srivastava, Potential of methyltransferase containing *Pseudomonas oleovorans* for abatement of arsenic toxicity in rice, *Sci. Total Environ.* 856 (2023) 158944.
- [55] S.N. Tan, J.W.H. Yong, Y.F. Ng, Arsenic exposure from drinking water and mortality in Bangladesh, *Lancet* 376 (2010) 1641–1642. [https://doi.org/10.1016/S0140-6736\(10\)62090-9](https://doi.org/10.1016/S0140-6736(10)62090-9).
- [56] J.W.H. Yong, S.N. Tan, Y.F. Ng, K.K.K. Low, S.F. Peh, J.C. Chua, A.A.B. Lim, Arsenic hyperaccumulation by *Pteris vittata* and *Pityrogramma calomelanos*: A comparative study of uptake efficiency in arsenic treated soils and waters, *Water Science and Technology* 61 (2010) 3041–3049. <https://doi.org/10.2166/wst.2010.223>.
- [57] T.S.W. Tow, Z.X. Eng, S.P. Wong, L. Ge, S.N. Tan, J.W.H. Yong, *Axonopus compressus* (Sw.) Beauv.: A potential biomonitor for molybdenum in soil pollution, *Int. J. Phytoremed.* 20 (2019) 1363–1368. <https://doi.org/10.1080/15226514.2016.1207599>.
- [58] A. van der Ent, A.J. Baker, R.D. Reeves, A.J. Pollard, H. Schat, Hyperaccumulators of metal and metalloids trace elements: facts and fiction, *Plant Soil* 362 (2013) 319–334. <https://doi.org/10.1007/s11104-012-1287-3>.
- [59] X. Shen, M. Dai, J. Yang, L. Sun, X. Tan, C. Peng, I. Ali, I. Naz, A critical review on the phytoremediation of heavy metals from environment: performance and challenges, *Chemosphere* 291 (2022) 1–11. <https://doi.org/10.1016/j.chemosphere.2021.132979>.
- [60] M. Li, Q. Heng, Z. Wang, Y. Jiang, X. Wang, X. He, J.W.H. Yong, T.M. Dawoud, S.U. Rahman, J. Fan, Phytoremediation efficiency of poplar hybrid varieties with diverse genetic backgrounds in soil contaminated by multiple toxic metals (Cd, Hg, Pb, and As), *Ecotoxicology and Environmental Safety* 283 (2024) 116843. <https://doi.org/10.1016/j.ecoenv.2024.116843>.
- [61] M.N.H. Sani, M. Amin, A.B. Siddique, S.O. Nasif, B.B. Ghaley, L. Ge, F. Wang, J.W.H. Yong, Waste-derived nanobiochar: a new avenue towards sustainable agriculture, environment, and circular bioeconomy, *Sci. Total Environ.* 905 (2023) 166881. <https://doi.org/10.1016/j.scitotenv.2023.166881>.
- [62] H. Shaghaleh, M. Azhar, M. Zia-ur-Rehman, Y.A. Hamoud, A.A.A. Hamad, M. Usman, M. Rizwan, J.W.H. Yong, H.F. Alharby, A.J. Al-Ghamdi, B.M. Alharbi, Effects of agro based organic amendments on growth and cadmium uptake in wheat and rice crops irrigated with raw city effluents: Three years field study, *Environ. Pollut.* 344 (2024) 123365. <https://doi.org/10.1016/j.envpol.2024.123365>.
- [63] Saleem, K., Asghar, M. A., Javed, H. H., Raza, A., Seleiman, M. F., Ullah, A., Rahman, A., Iqbal, S., Hanif, A. Imran, S., Nadeem, S. M., Du, J., Kocsy, G., Riaz, A., Yong, J. W. H (2023) Alleviation of arsenic toxicity-induced oxidative stress in lemon grass by methyl jasmonate. *South African Journal of Botany* 160: 547-559 <https://doi.org/10.1016/j.sajb.2023.07.034>.
- [64] L. Liu, Q. Zhang, L. Hu, J. Tang, L. Xu, X. Yang, J.W.H. Yong, X. Chen, Legumes can increase cadmium contamination in neighboring crops, *PLOS ONE* 7 (2012) e42944. <https://doi.org/10.1371/journal.pone.0042944>.
- [65] F.F. Qureshi, M.A. Ashraf, R. Rasheed, I. Hussain, M. Rizwan, M. Iqbal, J.W.H. Yong, Microbial-assisted alleviation of chromium toxicity in plants: A critical review, *Plant Stress* 11 (2024) 100394. <https://doi.org/10.1016/j.plstress.2024.100394>.
- [66] M. Ma, Y. Liu, C. Bai, J.W.H. Yong, The significance of chloroplast NAD(P)H dehydrogenase complex and its dependent cyclic electron transport in photosynthesis, *Front. Plant Sci.* 12 (2021) 661863. <https://doi.org/10.3389/fpls.2021.661863>.
- [67] J.W.H. Yong, Y.F. Ng, S.N. Tan, A.Y.L. Chew, Effect of fertilizer application on photosynthesis and oil yield of *Jatropha curcas* L., *Photosynthetica* 48 (2010) 208–218. <https://doi.org/10.1007/s11099-010-0026-3>.

- [68] D. Wu, C. Chen, Y. Liu, L. Yang, J.W.H. Yong, Iso-osmotic calcium nitrate and sodium chloride stresses have differential effects on growth and photosynthetic capacity in tomato, *Scientia Horticulturae* 312 (2023) 111883. <https://doi.org/10.1016/j.scienta.2023.111883>.
- [69] Liu, J., Caspersen, S., Yong, J.W.H. (2022). Growing together gives more rice and aquatic food. *eLife* (2022) <https://elifesciences.org/articles/77202>.
- [70] R. Wei, C. Chen, M. Kou, Z. Liu, Z. Wang, J. Cai, W. Tan, Heavy metal concentrations in rice that meet safety standards can still pose a risk to human health, *Commun Earth Environ* 4 (2023) 84. <https://doi.org/10.1038/s43247-023-00723-7>.
- [71] L.M. Nunes, G. Li, W.Q. Chen, A.A. Meharg, P. O'Connor, Y.G. Zhu, Embedded health risk from arsenic in globally traded rice, *Environ. Sci. Technol.* 56 (2022) 6415–6425. <https://doi.org/10.1021/acs.est.1c08238>.