

**PUBLICATIONS** 

APPROVED: 09 December 2024 doi: 10.2903/sp.efsa.2025.EN-9181

## Pest survey card on non-EU Choristoneura spp.

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#### **Abstract**

This document provides the conclusions of the pest survey card that was prepared in the context of the EFSA mandate on plant pest surveillance (M-2020-0114) at the request of the European Commission. The full pest survey card for the non-EU *Choristoneura* spp. is published and available online in the EFSA Pest Survey Card gallery at the following link and will be updated whenever new information becomes available: https://efsa.europa.eu/plants/planthealth/monitoring/surveillance/non-eu-choristoneura

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**Keywords:** 1CHONG, budworms, delimiting survey, detection survey, polyphagous, risk-based surveillance, Union quarantine pest

**Requestor:** European Commission

Question number: EFSA-Q-2023-00622

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**Acknowledgements:** EFSA wishes to thank the following for the support provided to this scientific output: the University of Padova in the context of the grant GP/EFSA/PLANTS/2022/05-Lot 2 for the preparation, Niklas Bjorklund and Josep A. Jaques Miret for the review and ISA experts Alicia Culot and Giulia Mattion (in the context of procedure EOI/EFSA/2022/01) for the finalisation and publication of this survey card. Melanie Camilleri, at the time of the preparation of the document, was a Seconded National Expert at EFSA.

**Suggested citation:** EFSA (European Food Safety Authority), Ipekdal K, Rassati D, Battisti A, Bjorklund N, Camilleri M, Rzepecka D, Jaques Miret JA, Carotti L and Salinari F, 2025. Pest survey card on non-EU *Choristoneura* spp. EFSA supporting publication 2025:EN-9181. doi:10.2903/sp.efsa.2025.EN-9181. Available online: https://efsa.europa.eu/plants/planthealth/monitoring/surveillance/non-eu-choristoneura.

**ISSN:** 2397-8325

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# 1 Introduction

This pest survey card was prepared in the context of EFSA's mandate on plant pest surveillance (M-2020-0114), at the request of the European Commission. Its purpose is to guide the Member States in preparing data and information for non-EU Choristoneura spp. surveys. These are required to design risk-based pest surveys, in line with current international standards. The 11 taxa (nine species and two subspecies) addressed in this pest survey card belong to the genus Choristoneura and are regulated as Union quarantine pests. Choristoneura species overwinter as larvae, pupation occurs in summer and the adults emerge and oviposit in summer. They can be univoltine (C. conflictana, C. fumiferana, C. lambertiana, C. occidentalis occidentalis, C. orae, C. pinus, C. retiniana), bivoltine (C. parallela, C. rosaceana), or semivoltine (C. occidentalis biennis). Species in the C. fumiferana species complex (C. carnana, C. fumiferana, C. lambertiana, C. occidentalis, C. orae, C. pinus and C. retiniana) are restricted to conifers (Abies, Larix, Picea, Pinus, Pseudotsuga and Tsuga). The remaining species are polyphagous (Corylus, Malus, Pistacia, Populus, Prunus, Pyrus, Rubus and Vaccinium and several herbaceous species). While C. conflictana and C. rosaceana can also feed on some conifers, C. parallela is restricted to angiosperms. The import of the conifers above into the EU is prohibited. Detection and delimiting surveys should be based on adult trapping and visual examination for pest presence and symptoms (foliar damage, defoliation, larval webbing around leaves). However, for detection surveys in conifer forests, adult trapping is the most practical method. Identification to species level is possible through both morphological identification keys and barcoding, the latter is recommended because of the occurrence of seven native EU Choristoneura species, the regulation of two subspecies of C. occidentalis, and the possibility of hybridisation in the C. fumiferana species complex.

## 2 The survey preparation

Table 1 addresses the key questions that are relevant for preparing a pest survey. First, the plant pest needs to be characterised in terms of its life cycle and biology. Then, the structure and size of the target population needs to be characterised and these analyses should be tailored to the situation in each Member State. Figure 1 gives examples of the components of a target population for the non-EU *Choristoneura* spp. and is not necessarily exhaustive. Finally, the detection process needs to be characterised in terms of the sequence of detection and identification methods required for the survey.



Table 1: Preparation of surveys for the non-EU *Choristoneura* spp.

SURVEY QUESTION	SECTIONS	KEY INFORMATION
WHAT?	1. The pest and its biology	Non-EU <i>Choristoneura</i> spp. develop through egg, larva, pupa and adult stages. Adults emerge and oviposit in summer except the bivoltine <i>C. rosaceana</i> which also oviposits in spring.
WHERE?	2. Target population	Most of the non-EU <i>Choristoneura</i> spp. hosts are conifers. Crops such as <i>Corylus</i> , <i>Malus</i> , <i>Pistacia</i> , <i>Populus</i> , <i>Prunus</i> , <i>Pyrus</i> , <i>Rubus</i> and <i>Vaccinium</i> , along with some herbaceous species, are major hosts of <i>C. conflictana</i> , <i>C. parallela</i> and <i>C. rosaceana</i> . Based on this host grouping and on the recommended detection method (trapping), detection survey strategies differ between conifer forests and the other host situations. Delimiting surveys should focus on the species-specific hosts occurring in the survey area. Epidemiological units: areas that contain at least one individual host plant (forest, agricultural and urban areas).
		(ports, airports, nurseries, gardens, urban parks and plant importing companies).  Inspection unit: area covered by pheromone
		trap or individual host plants.
HOW?	3. Detection and identification	Recommended method: adult trapping pheromone traps for detection surveys in conifer forests, combined with visual examination in other host situations (e.g. nurseries, orchards); adult trapping and visual examination for delimiting surveys. In all cases identification should be confirmed by molecular methods.
WHEN?		Although exact timing of the preferred survey period varies depending on the species, the adults can usually be detected in summer and the larvae all year around.

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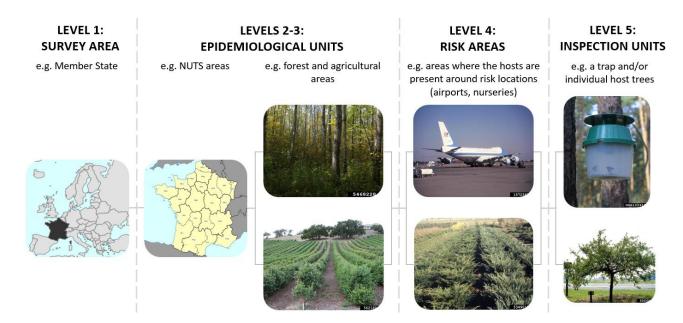


Figure 1: Example of the hierarchical structure of the target population for non-EU *Choristoneura* spp. (Sources: Eurostat, 2022 (levels 1–2); Steven Katovich, Bugwood.org (level 3, top); Gerald Holmes, Strawberry Center, Cal Poly San Luis Obispo, Bugwood.org (levels 3, bottom and 4, top); Norbert Frank, University of West Hungary, Bugwood.org (level 4, bottom); Landesforstpräsidium Sachsen, Bugwood.org (level 5, top); T. Davis Sydnor, The Ohio State University, Bugwood.org (level 5, bottom))



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## 3 From survey preparation to survey design

Figure 2 shows the next steps after the survey preparation for designing statistically sound and risk-based detection and delimiting surveys of the non-EU *Choristoneura* spp. Guidance on the selection of type of survey, related survey preparation and design, is provided in the EFSA general guidelines for pest surveys<sup>1</sup>.

#### **DETECTION SURVEYS: SUBSTANTIATION OF PEST FREEDOM** Characterise the plant pest Survey preparation Characterise the target population Define the Characterise method(s) for inspection units structure (environmental suitability, host pest detection and **EFSA Pest Survey Card** range, spread capacity to define idemiological units and risk factors) per detection identification method Survey design Define the size of host Set the target Set the overall population for each confidence level and subdivision of the target method sensitivity EFSA Pest Survey design prevalence population RiBESS+ software Allocate Sample size inspections, (inspections, samples, tests) samples, tests Survey implementation Report results Integrate survey Select NPPO survey instructions (data to collect) including survey Conduct the design with survey survey design and survey instructions sites (data to collect)

#### **DELIMITING SURVEYS: DELIMITATION OF INFESTED ZONES** Characterise the plant pest Survey preparation Characterise spread Characterise method(s) for Define the inspection Outbreak capacity, host plants units per detection **EFSA Pest Survey Card** report population, risk method identification factors Survey design Define the Set the target Define structure and Set the overall -source of infestation confidence level and size of host population potentially infested zone method sensitivity for each survey band design prevalence **EFSA Pest Survey** survey band width Guidelines Allocate RiBESS+ software inspections, mples, tests to (inspections, samples, tests) survey bands Survey implementation Report results Integrate survey Survey bands Select NPPO survey instructions including survey until the design with survey survey design and sites

Figure 2: Steps required for the preparation, design and implementation of detection and delimiting surveys, in accordance with the methodology for statistically sound and risk-based surveillance<sup>1</sup>

delimited

(data to collect)

assumptions

<sup>&</sup>lt;sup>1</sup> EFSA (European Food Safety Authority), Lázaro E, Parnell S, Vicent Civera A, Schans J, Schenk M, Cortiñas Abrahantes J, Zancanaro G and Vos S, 2020. General guidelines for statistically sound and risk-based surveys of plant pests. EFSA supporting publication 2020:EN-1919. 65 pp. doi:10.2903/sp.efsa.2020.EN-1919 <a href="https://efsa.onlinelibrary.wiley.com/doi/10.2903/sp.efsa.2020.EN-1919">https://efsa.onlinelibrary.wiley.com/doi/10.2903/sp.efsa.2020.EN-1919</a>



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## **Relevant EFSA outputs**

- General guidelines for statistically sound and risk-based surveys of plant pests: https://efsa.onlinelibrary.wiley.com/doi/10.2903/sp.efsa.2020.EN-1919
- Pest survey card on non-EU Choristoneura spp.: <a href="https://efsa.europa.eu/plants/planthealth/monitoring/surveillance/non-eu-choristoneura">https://efsa.europa.eu/plants/planthealth/monitoring/surveillance/non-eu-choristoneura</a>
- Pest categorisation of non-EU *Choristoneura* spp. : https://doi.org/10.2903/j.efsa.2019.5671
- Index of the EFSA Plant Pest Survey Toolkit: <a href="https://efsa.europa.eu/plants/planthealth/monitoring/surveillance/index">https://efsa.europa.eu/plants/planthealth/monitoring/surveillance/index</a>
- EFSA Pest Survey Card gallery: https://efsa.europa.eu/plants/planthealth/monitoring/surveillance/gallery
- Pest survey cards: what, when, where and how to survey?
   https://efsa.europa.eu/plants/planthealth/monitoring/surveillance/video-pest-survey-card
- The statistical tool RiPEST: https://r4eu.efsa.europa.eu/app/surveillance
- The RiPEST manual: <a href="https://zenodo.org/doi/10.5281/zenodo.8335472">https://zenodo.org/doi/10.5281/zenodo.8335472</a>
- The statistical tool RiBESS+: <a href="https://r4eu.efsa.europa.eu/app/ribess">https://r4eu.efsa.europa.eu/app/ribess</a>
- The RiBESS+ manual: https://zenodo.org/doi/10.5281/zenodo.664465
- The RiBESS+ video tutorial: <a href="https://youtu.be/qYHqrCiMxDY">https://youtu.be/qYHqrCiMxDY</a>

