

This is an author produced version of a paper published in Animal. This paper has been peer-reviewed and is proof-corrected, but does not include the journal pagination.

Citation for the published paper:

Jansson, A. & Lindberg, J. E. (2012) Forage-only diet alters the metabolic response of horses in training. *Animal*. First View Article, Available on CJO 2012, http://dx.doi.org/10.1017/S1751731112000948

Access to the published version may require journal subscription. Published with permission from: Cambridge University Press.

Standard set statement from the publisher:

2.2 The author may post the AM version of the author's article in the Institutional Repository of the institution in which the author worked at the time the article was first submitted or (for appropriate journals) in PubMed Central or UK PubMed Central or arXiv, provided the posting is accompanied by a prominent statement that the article has been accepted for publication and will appear in a revised form, subsequent to peer review and/or editorial input by Cambridge University Press, in <Journal title> published by Cambridge University Press, together with a copyright notice in the name of the copyright holder (Cambridge University Press or the sponsoring Society, as appropriate). On publication the full bibliographical details of the article (volume: issue number (date), page numbers) must be inserted after the journal title, together with a link to the Cambridge website address for the Journal. [...] 2.4. The author may post the VoR version of the article (in PDF or HTML form) in the Institutional Repository of the institution in which the author worked at the time the article was first submitted, or (for appropriate journals) in PubMed Central or UK PubMed Central or arXiv, no sooner than one year after first publication of the article in the Journal, subject to file availability and provided the posting includes a prominent statement of the full bibliographical details, a copyright notice in the name of the copyright holder (Cambridge University Press or the sponsoring Society, as appropriate), and a link to the online edition of the Journal at Cambridge Journals Online.

Epsilon Open Archive http://epsilon.slu.se

1	A forage-only diet alters the metabolic response of horses in training
2	
3	A. Jansson and J. E. Lindberg
4	Dept of Animal Nutrition and Management, Swedish University of Agricultural
5	Sciences, SE-750 07 Uppsala, Sweden.
6	
7	Corresponding author: Anna Jansson. E-mail: Anna.Jansson@slu.se
8	
9	Running head: Exercise response in horses on forage-only diet.
10	
11	Abstract
12	Most athletic horses are fed a high-starch diet, despite the risk of health problems.
13	Replacing starch concentrate with high-energy forage would alleviate these health
14	problems, but could result in a shift in major substrates for muscle energy supply
15	from glucose to short-chain fatty acids (SCFA) due to more hindgut fermentation of
16	fibre. Dietary fat inclusion has previously been shown to promote aerobic energy
17	supply during exercise, but the contribution of SCFA to exercise metabolism has
18	received little attention. This study compared metabolic response to exercise and
19	lactate threshold (V <sub>La4</sub> ) in horses fed a forage-only diet (F) and a more traditional

20	high-starch, low-energy forage diet (FC). The hypothesis was that diet F would
21	increase plasma acetate concentration and increase $V_{\text{La4}}$ compared with diet FC. Six
22	Standardbred geldings in race training were used in a 29 day change-over
23	experiment. Plasma acetate, non-esterfied fatty acids (NEFA), lactate, glucose and
24	insulin concentrations and venous pH were measured in samples collected before,
25	during and after a treadmill exercise test (ET, day 25) and muscle glycogen
26	concentrations before and after ET. Plasma acetate concentration was higher before
27	and after exercise in horses on diet F compared with diet FC and there was a
28	tendency (P=0.09) for increased $V_{La4}$ on diet F. Venous pH and plasma glucose
29	concentrations during exercise were higher in horses on diet F than diet FC, as was
30	plasma NEFA on the day after ET. Plasma insulin and muscle glycogen
31	concentrations were lower for diet F, but glycogen utilisation was similar for the two
32	diets. The results show that a high-energy, forage-only diet alters the metabolic
33	response to exercise and, with the exception of lowered glycogen stores, appears to
34	have positive rather than negative effects on performance traits.

37 Keywords: acetate, blood pH, exercise, insulin, muscle glycogen

38

39

40	mp	lication

It is a serious animal welfare issue that diets associated with gastrointestinal disorders and abnormal and stereotypical behaviour are fed to horses today. There is an urgent need for diets that support the natural digestive function and behaviour of horses. Such diets would reduce welfare problems and healthcare costs for the horse industry.

46

## 47 Introduction

48 Current practice in feeding performance horses world-wide is to use fairly late-cut 49 forage, whereas for example in dairy production early-cut, high-energy forage is 50 used. The high energy requirements of many performance horses means that 51 supplementation with more energy-dense feeds is necessary and more than 40% of 52 the diet commonly consists of cereal-based, high-starch concentrates (Glade, 1983;

53	Redbo et al., 1998; Williamson et al., 2007). This is a serious animal welfare issue,
54	since low forage to concentrate ratios and high starch intake are associated with
55	reduced gut microbial stability (Willing et al., 2009), gastrointestinal disorders (Tinker
56	et al., 1997; Hudson et al., 2001; Luthersson et al., 2009) and abnormal and
57	stereotypical behaviour (Gillham <i>et al.</i> , 1994, Redbo <i>et al.</i> , 1998; Waters <i>et al.</i> ,
58	2002). In addition, rhabdomyolysis has been associated with high concentrate
59	intakes (MacLeay et al., 1999). We have shown in earlier studies that Standardbred
60	horses in training can maintain body weight and condition on high-energy, forage-
61	only diets (Connysson <i>et al.</i> 2006; Muhonen <i>et al.,</i> 2009; Connysson <i>et al.,</i> 2010).
62	We found no detrimental effect of increased crude protein (CP) intake (which may be
63	associated with the use of high-energy forages) on plasma lactate concentration and
64	pH during exercise, but urinary pH decreased and evaporative losses tended to
65	increase (Connysson et al., 2006). However, the effect on exercise performance of
66	forage-only diets compared with the traditional forage-concentrate diet has not yet
67	been studied.

From a physiological point of view the horse is adapted to continuous grazing of a forage-only diet (low in starch content) and has a well-developed symbiosis, with

70	the hindgut microbiota fermenting the forage fibres. This results in the production of
71	energy-yielding substrates in the form of short-chain fatty acids (SCFA) and the
72	proportion of acetate increases and propionate decreases when forage:concentrate
73	ratio is increased (Hintz et al., 1971; Willard et al., 1977). This suggests that horses
74	are well-adapted to rely on fat metabolism and aerobic energy supply at rest, but
75	probably also during exercise. The adaptation to, and importance of, aerobic
76	substrate utilisation during exercise in Thoroughbred and Standardbred horses has
77	been documented in a number of studies (Lindholm and Phiel, 1974; Wilson et al.,
78	1987; Essén-Gustavsson et al., 1989) and is also confirmed by the correlation
79	between the performance of Standardbred horses and plasma lactate threshold
80	$(V_{La4})$ (Persson, 1983). Equine muscle also shows high plasticity with respect to its
81	adaptation to energetic demands as a result of exercise training (Voiton et al., 2007).
82	Therefore, it appears reasonable to assume that the substrate profile for efficient
83	muscle energy metabolism in horses will comprise glucose, long-chain FA from the
84	diet or from body lipid stores (Pagan et al., 2002; Geor, 2006) and SCFA from
85	hindgut fermentation of dietary fibre (Palmgren-Karlsson et al., 2002). The aim of this
86	study was to compare the effects of a high-energy forage-only diet with those of a

50:50 (DM basis) forage:concentrate (starch-rich) diet in terms of lactate threshold ( $V_{La4}$ ), muscle glycogen concentration and metabolic plasma profile in Standardbred horses in training. The hypothesis was that the forage-only diet would increase plasma acetate concentration and  $V_{La4}$ .

91

### 92 Materials and methods

93

### 94 Horses

Six Standardbred geldings in race training were used, aged 6.5 ± 0.4 years (mean ± 95 SD). The average number of races in which the horses had competed was  $27 \pm 8$ 96 97 and the average racing record was 77.3 ± 0.8 s/1000 m. They had an initial body weight (BW) of 515 ± 21 kg. The horses were kept at a training camp for harness 98 99 racing 20 km south of Uppsala, Sweden. They were housed in individual stalls on 100 wood shavings during the night and were kept together in a sand/clay paddock 101 between 08:00-15:00 h on days without training. All horses had passed a flexure test prior to the study and were regarded as healthy. The experiment was approved by 102

103 the Uppsala local ethics committee and was conducted in the period October-104 December 2007.

105

### 106 Experimental design

107 *Diets* 

The horses were offered a forage-only diet (F) consisting of early-cut haylage 108 109 (timothy, meadow fescue mixture) (Table 1) and a mixed diet (FC) consisting of late-110 cut haylage (timothy, meadow fescue mixture) supplemented with concentrate (50:50 111 dry matter basis) in a change-over design experiment with 29-day experimental 112 periods. Feed allowance was based on individual BW and was 13-17.4 kg haylage 113 and 180-240 g sugar (only to ensure complete intake of the salt, mineral and vitamin 114 supplements) for diet F and 6.3-8.4 kg haylage, 5.3-7.1 kg oats, 0.9-1.2 kg soy bean meal, 0.18-0.24 kg wheat bran and 90-120 g sugar for diet FC. The diets were 115 116 estimated to be iso-caloric and iso-nitrogenous, and provided energy and nutrients 117 according to requirements specified by NRC (1989). Horses on both diets were 118 offered a mineral and vitamin supplement (51 ± 2 g/day, Miner Röd, Krafft, 119 Falkenberg, Sweden), NaCl (36  $\pm$  1 g/day) and those on diet FC ground chalk

120	(calcium carbonate, $34 \pm 1$ g/day) to meet mineral and vitamin requirements specified
121	by NRC (1989). Water was provided ad libitum from graded buckets. The forage
122	allowance was fed in the afternoon and the concentrate and mineral and vitamin
123	supplement (diet FC) at 15.00, 23.00 and 06.00 h. With diet F, the mineral and
124	vitamin supplement was fed at 23.00 and 06.00. Diet FC was introduced gradually
125	during the experimental period (on days 1 and 2, horses were fed 50% of the F diet
126	and 50% of the FC diet, and then the FC diet was increased by 10% per day until the
127	full ration was reached on day 7). Diet F was introduced abruptly on day 1.
128	
129	Training
130	The horses were given sub-maximal warm-up (3.5-5 km, heart rate < 200 beats/min)
131	and intensive training (heart rate > 200 beats/min) on an oval or straight field track
132	(approx. 0.6% incline) on days 2, 5, 7, 9, 13, 17, 21, 25 and 29 in order to maintain
133	but not improve fitness. The intensive training consisted of interval training (4
134	intervals of 600 m) or 1600-2000 m heats and the training protocol was exactly the

- 135 same in both periods.
- 136

#### 137 Exercise test

138 On day 25 of each experimental period, the horses performed a standardised 139 incremental exercise test (ET) on a treadmill (Säto, Knivsta, Sweden) located at a 140 veterinary clinic 25 km from the training camp. The horses were transported to the 141 clinic by trailer. All horses had prior experience of exercise on this treadmill (minimum 142 2 occasions). 143 Prior to ET, the horses were kept together in the paddock from 07:30 to 10:30 h 144 and were then offered 1 kg of forage (diet F) and 1 kg of oats (diet FC). After this, 145 water but no feed was offered until the horses returned to the stable at about 24:00 h. 146 This design was chosen because practical experience indicated that many horses do

147 not consume any feed while at the racetrack. On the day after ET, all horses were

148 fed at 06.00 h as usual, but were kept in their stalls until 11:00 h.

Exercise testing consisted of two phases. The first phase (warm-up, walk at 2 m/s for 3 minutes, trot 6 m/s for 5 min, trot 9.5 m/s for 5 min and walk 2 m/s for 3 min, no incline) was performed 25 min prior to the second phase, which consisted of 5 minutes of walk (0% incline) and then a stepwise increase in velocity (6.0, 7.0, 8.0, 9.0, 9.5 m/s) at an incline of 6.3% every second minute until the horses reached or 154 exceeded a heart rate of 200 beats/min. Four horses reached 9.0 m/s and two 155 reached 9.5 m/s. The horses walked for 5 min on the treadmill after the final step. 156 157 Sampling 158 Feed and water intake, body weight and condition Samples of concentrate were collected from each batch used in every period and 159 160 samples of forage from each new bale opened and kept frozen at -20°C until analysis. The forage samples were pooled and analysed as one sample per feed and 161 162 period. Feed leftovers were weighed and eliminated every day. Daily nutrient and 163 energy intake was calculated using feed intake data and analysis of feeds and 164 leftovers. Water intake (drinking) was measured on days 19 to 28. Body weight was recorded before each training session, before the warm-up 165 prior to ET, immediately after ET, on the day after ET (at 11.00 h) and on days 27-29 166 (afternoon). Changes in body condition were assessed by a simple recording of 167 whether the ribs were easy to palpate or not and whether they were visible during 168 169 motion.

170

## 171 Exercise test

172	Heart rate was recorded by a heart frequency meter (Polar RS800, Kempele,
173	Finland) during exercise and manually post-exercise with a stethoscope. Heart rate
174	was measured before warm-up, during warm-up (9.5 m/s), immediately after warm-
175	up, after 25 minutes of rest, during each incremental step, after 5 minutes walking on
176	the treadmill and 10, 25, 55 and 85 minutes after the end of walking.
177	Breathing frequency was recorded before warm-up, directly after warm-up, after
178	25 minutes of rest, after 5 minutes walking on the treadmill and 10, 25, 55 and 85
179	minutes after the end of walking. Rectal temperature was recorded before warm-up,
180	directly after warm-up, after 25 minutes of rest and 10, 25, 55, 85 minutes after the
181	end of walking.
181 182	end of walking. A jugular catheter was introduced under local anaesthesia (Carbocaine 20 mg/ml,
182	A jugular catheter was introduced under local anaesthesia (Carbocaine 20 mg/ml,
182 183	A jugular catheter was introduced under local anaesthesia (Carbocaine 20 mg/ml, Astra Zeneca AB, Sweden) before transport to the clinic and blood samples (20
182 183 184	A jugular catheter was introduced under local anaesthesia (Carbocaine 20 mg/ml, Astra Zeneca AB, Sweden) before transport to the clinic and blood samples (20 ml/sample) were collected in Li-heparinised tubes before warm-up, immediately after

188 samples were kept chilled until centrifuged and frozen at -20C° for later analysis.
189 Samples collected for analysis of blood pH, TCO<sub>2</sub> and HCO<sub>3</sub> were analysed within
190 10 min of collection.

Muscle biopsies were taken from *m. gluteus medius* at a depth of approximately 6 cm according to the method described by Lindholm and Piehl (1974). A local anaesthetic (Carbocain 20 mg/ml, Astra Zeneca AB, Sweden) was applied to the area and a nose twitch was used. Biopsies were taken before warm-up and immediately after ET, frozen in liquid nitrogen and stored at -80°C until analysis.

196

#### 197 Analyses

Preparation and conventional chemical analyses (dry matter (DM), nitrogen (N), neutral detergent fibre (NDF), acid detergent fibre (ADF), lignin, water-soluble carbohydrate (WSC) and ash) of feeds were performed as described by Palmgren-Karlsson *et al.* (2000). Minerals were analysed with inductively coupled plasma optical emission spectrometry (ICP-OES) (SS-EN 14538:2006, Ametek Spectro, Kleve, Germany). The metabolisable energy content of the forage was estimated by an *in vitro* method (Lindgren, 1979).

# 206 Analyses of plasma, blood and muscle

207	Plasma insulin was analysed using an ELISA method (Mercodia equine insulin kit,
208	Mercodia, Uppsala, Sweden). For quantitative determination of non-esterified fatty
209	acids (NEFA), an enzymatic colorimetric method was used (ACS-ACOD method,
210	Wako Chemicals GmbH, Neuss, Germany). Plasma urea, acetate and glucose
211	concentrations were analysed with an enzymatic colorimetric/UV-method (Boehringer
212	Mannheim/R-Biopharm, Darmstadt, Germany) and lactate using an ELISA method
213	(R-Biopharm GmbH, Darmstadt, Germany). Venous pH, TCO $_2$ and HCO $_3$ were
214	analysed using an i-STAT®1 analyser (Abbot Laboratories, Abbot Park, Illinois, USA).
215	Total plasma protein (TPP) concentration was measured by refractometer (Atago,
216	Sur-Ne, Tokyo, Japan) in samples taken before warm-up, before ET, 55 min after
217	walking and the day after walking.
218	Muscle samples for glycogen analysis were freeze-dried and dissected free
219	from visible blood, connective tissue and fat under a microscope before analysis. A
220	sample of about 1-2 mg muscle fibre was boiled in 1 M HCl and glucose residues
221	were determined by fluorescence (Lowry and Passonneau, 1973).

### 223 Calculations and statistical analysis

The velocity at plasma lactate concentration 4 mmol/l ( $V_{La4}$ ) was calculated individually from an exponential curve fitted by Microsoft Office Excel 2007.

226	All data were subjected to analysis of variance (GLM procedure in the Statistical
227	Analysis Systems package 9.1) (SAS Institute Inc. Cary, NC, USA) using the
228	following model: $Y_{i j k} = \mu + \alpha_i + \beta_j + \gamma_k + \epsilon_i + (\beta\gamma)_{j k} + e_{i j k i}$ , where $Y_{i j k}$ is the
229	observation, $\mu$ the mean value, $\alpha_i$ the effect of animal, $\beta_j$ the effect of diet, $\gamma_k$ the
230	effect of sample, $\epsilon_I$ the effect of period, $(\beta\gamma)_{j\ k}$ the effect of interaction between diet
231	and sample and $e_{i \ j \ k \ l}$ the residuals; $e_{i \ j \ k \ l}$ ~IND (0, $\delta^2$ ). The value used for
232	significance within and between treatments was P<0.05. Values are presented as
233	least square mean $\pm$ standard error of the mean.

234

### 235 **Results**

236 General observations

All horses completed the study. No health problems were observed, except for one horse which developed a fever two days into period 1 (resulting in three missed

239	training sessions). There were leftovers on both diets (Table 2) and forage intake
240	corresponded to 69% of the allowance on both diets, resulting in a
241	forage:concentrate ratio of 40:60 for diet FC. A comparison of the in vitro digestible
242	organic matter (IVDOM) of the individual leftovers and the feed samples collected on
243	days 20-22 showed consistently lower IVDOM in the leftovers compared with the
244	feed for diet F, but not diet FC (Diet F: feed=78.3%, leftovers=73.7-76.9%; diet FC:
245	feed=66.9%, leftovers=63.7-69.6%), indicating selective feed intake in all individuals
246	on diet F.
247	Body condition was similar in four individuals on both diets but two individuals
248	had higher body condition on diet FC (horse 1 changed from 'ribs visible at motion' to
249	'not visible but easy to palpate' and horse 2 changed from 'ribs not visible at motion
250	but easy to palpate' to 'not visible, not easy to palpate'). Mean BW was higher
251	(P=0.0006) for horses on diet F (519.0 $\pm$ 0.5 kg) than diet FC (516.0 $\pm$ 0.5 kg). Water
252	intake was higher with diet F than diet FC (30.0 $\pm$ 0.5 vs 19.0 $\pm$ 0.5 l/day, P<0.0001).
253	

254 Exercise test

255	There was no significant difference in $V_{\mbox{\tiny La4}}$ between diets, although there was a
256	tendency (P=0.086) for higher V <sub>La4</sub> in horses on diet F (8.0 $\pm$ 0.1 $vs$ 7.6 $\pm$ 0.1 m/s).
257	Plasma lactate concentration was higher (P<0.05) in horses on diet FC compared
258	with diet F at the end of the 5-min walk and 10 min after the walk on the treadmill
259	(diet FC: 8.1 ± 0.4 and 4.4 ± 0.4 $vs$ diet F: 5.7 ± 0.4 and 3.2 ± 0.4 mmol/l). Venous
260	pH (Figure 1), TCO <sub>2</sub> and HCO <sub>3</sub> (P=0.0001) in connection with exercise were higher
261	in horses on diet F compared with diet FC (pooled data for TCO <sub>2</sub> : 31.2 $\pm$ 0.2 vs 30.3
262	$\pm$ 0.2 mmol/l and HCO <sub>3</sub> : 30.0 $\pm$ 0.2 vs 29.1 $\pm$ 0.2 mmol/l). Acetate concentration was
263	higher with diet F than diet FC (P=0.0001) and plasma NEFA concentration on the
264	day after ET was higher (P<0.05) with diet F compared with diet FC (Figure 2).
265	Plasma insulin concentration was higher (P=0.0001) in horses on diet FC compared
266	with diet F (Figure 2) and glucose concentration was higher (P<0.05) with diet F
267	immediately after warm-up and at 9 m/s, but lower (P<0.05) on the day after ET
268	(Figure 2). Muscle glycogen content was higher (P<0.05) in horses on diet FC than
269	diet F both before (644 $\pm$ 22 <i>vs</i> 560 $\pm$ 22 mmol glucosyl units/kg dry weight) and after
270	exercise (546 $\pm$ 22 vs 473 $\pm$ 22 mmol glucosyl units/kg dry weight). There were no
271	differences between the diets in heart rate (109 $\pm$ 1 and 111 $\pm$ 1 beats/min in F and

272	FC, respectively), breathing frequency (62 $\pm$ 3 and 68 $\pm$ 3 beats/min in F and FC,
273	respectively), or rectal temperature (38.5 $\pm$ 0.1 and 38.5 $\pm$ 0.1 beats/min in F and FC,
274	respectively), before, during and after exercise (pooled data).
275	There were no differences in BW between diets before (diet FC: 509 $\pm$ 1 and
276	diet F: 511 $\pm$ 2 kg) and after warm-up and ET (diet FC: 501 $\pm$ 1.0 and diet F: 500 $\pm$ 1
277	kg). The pre-exercise BW was not recovered until day 28 on both diets. Mean TPP
278	during ET day and on the day after was lower with diet F than diet FC (64.6 $\pm$ 0.6 $\nu s$
279	67.0 ± 0.6 g/l).

## 281 **Discussion**

Horses fed the forage-only (F) diet showed an altered metabolic response during exercise compared with horses on the forage-concentrate diet and few, if any, effects likely to impair performance were observed. There was an increase in plasma acetate concentration on the F diet, as expected. Moreover, with this diet plasma lactate concentration was significantly lower post-exercise and there was a tendency for higher  $V_{La4}$ , indicating that aerobic energy utilisation may have improved. In addition, venous pH increased during exercise in horses on the F diet, which could counteract the acidosis induced by the exercise typically performed by racehorses.
This indicates that high-energy, forage-only diets are an interesting alternative to
conventional diets.

292

293 The increased plasma levels of acetate in horses on the F diet can be explained by 294 more extensive hindgut fermentation due to higher fibre intake, and subsequent 295 production of SCFA (Hintz et al., 1971; Willard et al., 1977). SCFA absorbed from the 296 hindgut can be used as substrates in body tissue metabolism. Propionate may be 297 primarily used to produce glucose in the gluconeogenetic pathway, while acetate and 298 butyrate can be transformed to acetyl-CoA and then utilised as substrates for aerobic energy metabolism in the tri-carboxylic pathway in the muscle (Voiton et al., 2007). 299 The latter also applies to long-chain fatty acids (LCFA) that are metabolised through 300 β-oxidation in the muscle. It has been suggested that long-term feeding of 301 supplemental fat to exercising horses increases the mobilisation and speed of 302 mobilisation of free fatty acids (FA), increasing the speed of uptake into muscle of 303 304 free FA, lowering lactic acid production, imparting a glycogen-sparing effect and 305 increasing pre-exercise muscle glycogen levels (Potter et al., 1992; Harris and

306	Harris, 2005). However, there is a large variation in the effects reported in the
307	literature and only a few studies report direct benefits on performance traits (Geor,
308	2006). Pagan et al. (2002) used a stable glucose isotope and showed that a high-fat
309	diet can be glucose-sparing by increasing FA oxidation from body lipid stores during
310	exercise, suggesting that performance could be improved. In the present study no
311	clear effect of the F diet on performance was observed, but there was a lowering of
312	post-exercise plasma lactate concentration, confirming previous findings for a high-
313	fibre diet (Palmgren-Karlsson <i>et al.</i> , 2002).

315 Waller and Lindinger (2007) reported profound plasma alkalosis, i.e. a decrease in 316 venous plasma hydrogen concentration and an increase in plasma TCO<sub>2</sub> and HCO<sub>3</sub>-, after post-exercise oral administration of sodium acetate. These results were 317 318 confirmed in the present study, where venous pH, TCO<sub>2</sub> and HCO<sub>3</sub>- concentrations 319 were higher in horses on the F diet. The alkalising effect of a forage-only diet may 320 also be a direct result of the increased plant cell and organic acid intake typical of 321 herbivores (Houpt, 1989). Plant cells contain organic anions (citric, oxalic, malonic 322 and fumaric acids) that are electrically balanced by potassium and other cations.

323 Citrate and other organic anions are oxidised to CO<sub>2</sub> and H<sub>2</sub>O but their breakdown 324 requires hydrogen ions, which are derived from the hydration of carbon dioxide. 325 Thus, as organic anions are oxidised, HCO<sub>3</sub> is produced, which has an alkalotic 326 effect on body fluid pH (Houpt, 1989). The possibility of counteracting the acidosis 327 induced by intensive exercise by oral supplementation (generally with salts of  $HCO_3$ -) 328 has been studied for a long time, but the effects on performance are not unequivocal 329 (Kelso et al., 1987; Lawrence et al., 1987; Schuback et al., 2002). This may be the 330 first study to show that there is a natural way of achieving this effect, although a 331 positive effect on performance remains to be proven.

332

Plasma glucose levels were higher during exercise in horses on the F diet. The origin and importance of this elevation in glucose concentration for exercise performance is not clear, but endurance during both maximal (Lacombe *et al.*, 2001) and submaximal (Farris *et al.*, 1998) exercise is improved by glucose infusion, indicating that high blood glucose availability could improve performance. Alterations in glucose metabolism (both in tissue uptake and release from the liver) are likely to occur in horses on forage-only diets, based on the low insulin levels observed here and

340	earlier (Connysson et al., 2010), and might be the reason for the elevated glucose
341	levels during exercise. However, on the day after exercise tests the plasma glucose
342	concentration was lower in horses on the F diet and the NEFA concentration was
343	higher. The importance of this is also unclear, but it might reflect the demand for
344	glucose for resynthesis of glycogen and low dietary glucose availability (total daily
345	intake of WSC and starch was approximately 860 g and 3100 g/day on diet F and
346	FC, respectively) and increased utilisation of body fat for maintenance.
347	
348	The low WSC intake might also have affected muscle glycogen synthesis (Lacombe
349	et al., 2004). Muscle glycogen content was lower (-13%) before and after ET in
350	horses on the F diet, while relative glycogen depletion was similar. It is known that
351	glycogen synthase is activated by insulin (Devlin and Horton, 1985) and the low
352	insulin levels on the F diet might have been a limiting factor for glycogen synthesis.
353	Further studies are needed to determine whether total muscle glycogen storage
354	capacity is reduced on a forage-only diet or whether it is only the rate of synthesis
355	that is reduced. It is important to note that all horses had trained approximately 96 h
356	before ET and glycogen recovery might not have been complete. It is well-

357 documented that post-exercise recovery of glycogen stores takes several days in 358 horses (Snow et al., 1987; Hyyppä et al., 1997, Lacombe et al., 2004). While an 359 exercise-induced reduction in muscle glycogen content of 80% has been shown to 360 impair endurance during high-intensity exercise (Lacombe et al., 2001), the 361 importance of a smaller reduction, such as that induced by the F diet in the present 362 study, is not known. Interestingly, oral acetate supplementation of a typical hay-grain 363 diet has been shown to enhance the rate of glycogen re-synthesis during the initial 4-364 h recovery period after muscle glycogen depletion (Waller et al., 2009), but our data 365 suggest that increased acetate availability does not maximally support glycogen 366 repletion between training sessions. However, a recent study (Essén-Gustavsson et al., 2012) of horses on forage-only diets with different CP content showed that high 367 CP forage increased muscle glycogen content compared with a forage providing the 368 CP intake recommended by NRC (2007). This shows that CP is important for 369 370 glycogen content and might have increased the glycogen content in horses on diet FC in the present study. 371

372

373 One argument against using forage-only or high-forage diets for performance horses 374 is the risk of an unwanted increase in BW. This is probably based on the common 375 perception that fibre is 'bulk' and perhaps also on knowledge of the water-holding 376 capacity of plant fibres. In a study on riding horses (Ellis et al., 2002), a forage-only 377 diet increased body weight and heart rate during submaximal exercise, suggesting 378 that performance might be impaired. However, it should be noted that the energy 379 content of the forage in that study was not high enough to support the energy needs 380 of horses in race training and, accordingly, the digestibility was not high enough, 381 thereby causing 'bulk weight gain'. The present study showed no (ET days) or a 382 limited (3 kg in pooled data) increase in BW in horses fed a high-energy forage diet. 383 There was also no effect on heart rate and breathing frequency. It is likely that differences in chemical composition between forages are the reason for the 384 differences in BW change in these studies. In the present study, the forage in diet F 385 was early-cut and had high fibre digestibility, as reported by Ragnarsson and 386 Jansson (2011) for the same batch of forage. 387

388

389 Part of the increase in BW observed on a forage-only diet could also be due to increased plasma volume, as indicated by the lowered TPP. It has been suggested 390 391 that the hindgut serves as a fluid reservoir and that fibre-rich diets increase this 392 reservoir (Meyer, 1987). The change in TPP with altered fibre intake could therefore 393 reflect changes in the equilibrium between the gut and the extracellular fluid. It has 394 been shown that increased forage intake lowers TPP (Danielsen et al., 1995) and 395 also that signs of dehydration (increased TPP) following feed deprivation are delayed 396 in horses on a forage-only diet compared with those on a mixed diet (Connysson et 397 al., 2010). However, BW recovery was similar on both diets in the present study. The 398 results also showed that it may take two to three days for horses transported to an exercise event to recover their body weight. The loss of BW on ET day was due to 399 lack of feed intake (less than 60% of the allowance) and to fluid and faeces losses 400 401 during transportation and exercise.

402

403 There is also anecdotal information that forage intake capacity is limited in
404 performance horses. Horses in the present study consumed forage corresponding to
405 1.95% of BW and four of the horses maintained similar body condition on both diets.

406	It is possible that the other two horses would have been maintained on a forage diet
407	with slightly higher energy content (around 11 MJ ME/kg dry matter). In all horses on
408	the forage-only diet, the maintenance of energy intake and body condition was to
409	some extent due to selection of the forage offered, as reflected in lower digestibility of
410	leftovers from this diet. This contributed to higher energy intake on the forage-only
411	diet and was probably an attempt to maximise energy intake.
412	
413	Another concern about a forage-dominated diet for athletic horses is the increased
414	heat increment of feeding. In the present study this was probably reflected in the
415	higher water intake on diet F. However, body temperature was similar, as would have
416	been expected in a homeothermic animal, and it was not possible to draw any
417	conclusions about evaporative fluid losses during exercise from the results, since
418	only BW was measured before and after exercise (including sweat, faecal and
419	respiratory losses).
420	
421	No feed-related clinical or behavioural disturbances very observed during the study.

422 Exercise temperament, novel object reaction, voluntary motion and post-exercise

423	feeding (Jansson, 2010) were also evaluated in the present study and numerical, but
424	not statistically significant, differences were detected, with the exception of
425	observations on post-exercise feed intake, which was more common on diet F.
426	However, subjectively more aggression was observed in horses fed diet FC,
427	especially in the afternoon when horses were fetched from the paddock (one at a
428	time) to be put in their boxes, where feed was available. Therefore, our impression is
429	that the FC diet might have affected behaviour (making horses more active and
430	reactive), although the recording system used could not verify this.

In conclusion, the present study indicates that a high-energy, forage-only diet alters
the metabolic response to exercise and, with the exception of lowered glycogen
stores, appears to have positive rather than negative effects on performance traits of
Standardbred horses.

436

# 437 Acknowledgements

438	The authors want to thank HästKraft AB and Lantmännen Box for providing the
439	forages and trainer Anna Svensson and the staff at Mälarkliniken, Sigtuna, Sweden,
440	for assistance during the study.
441	
442	Source of funding
443	This research received financial support from Stiftelsen Svensk Hästforskning,
444	Sweden.
445	
446	References
447	Connysson M, Muhonen S, Lindberg JE, Essén-Gustavsson B, Nyman G, Nostell K
448	
-	and Jansson A 2006. Effects on exercise response, fluid and acid-base balance of
449	and Jansson A 2006. Effects on exercise response, fluid and acid-base balance of protein intake from forage-only diets in Standardbred horses. Equine Veterinary
449	protein intake from forage-only diets in Standardbred horses. Equine Veterinary
449 450	protein intake from forage-only diets in Standardbred horses. Equine Veterinary Journal Supplement 36, 648-653.

454	Danielsen K, Lawrence LM, Siciliano P, Powell D, Thomson K 1995. Effects of diet
455	on weight and plasma variables in endurance exercised horses. Equine Veterinary
456	Journal Supplement 18, 372-377.
457	Devlin JT and Horton ES 1985. Effects of prior high-intensity exercise on glucose
458	metabolism in normal and insulin-resistant men. Diabetes 34, 973-979.
459	Ellis JM, Hollands T and Allen DE 2002. Effect of forage on body weight and
460	performance. Equine Veterinary Journal Supplement 34: 66-70.
461	Essén-Gustavsson B, McMiken D, Karlstrom K, Lindholm A, Persson S and Thornton
462	J 1989. Muscular adaptation of horses during intensive training and detraining.
463	Equine Veterinary Journal 21, 27-33.
464	Essén-Gustavsson B, Connysson M and Jansson A 2010. Effects of crude protein
465	intake from forage-only diets on muscle amino acids and glycogen levels in horses in
466	training. Equine Veterinary Journal 38(42), 341-346.

Farris JW, Hinchcliff KW, McKeever KH, Lamb DR and Thomson DL 1998. Effects of
tryptophan and of glucose on exercise capacity of horses. Journal of Applied
Physiology 85, 807-816.

470 Geor RJ 2006. The role of nutritional supplements and feeding strategies in equine

471 athletic performance. Equine and Comparative Exercise Physiology 3, 109-119.

472 Gillham SB, Dodman NH, Shuster L, Kream RK and Rand W 1994. The effect of diet

473 on cribbing behaviour and plasma  $\beta$ -endorphins in horses. Applied Animal Behaviour

- 474 Science 41, 147-153.
- 475 Glade MJ 1983. Nutrition and performance of racing Thoroughbreds. Equine

476 Veterinary Journal 15, 31-36.

477 Harris PA and Harris RC 2005. Ergogenic potential of nutritional strategies and

478 substances in horses. Livestock Production Science 92, 147-165.

479 Hintz HF, Argenzio RA and Schryver HF 1971. Digestion coefficients, blood glucose

- 480 levels and molar percentages of VFA in intestinal fluid of ponies fed varying forage-
- 481 grain ratios. Journal of Animal Science 33, 992.

Houpt RT 1989. Water balance and excretion. In Dukes's Physiology of Domestic
Animals 10th edition (ed Swenson MJ), pp. 496. Comstock, Cornell University Press,
Ithaca, New York, USA.

Hudson JM, Cohen ND, Gibbs PG and Thompson JA 2001. Feeding practices
associated with colic in horses. Journal of the American Veterinary Medical
Association 219, 1419-1425.

488 Hyyppä S, Räsanen LA, Pösö AR 1997. Resynthesis of glycogen in skeletal muscle

489 from Standardbred trotters after repeated bouts of exercise. American Journal of

490 Veterinary Research 58, 162-166.

491 Jansson, A. 2010. Effects of diet on behaviour of Standardbred horses in training. In

492 The Impact of Nutrition on the Health and Welfare of Horses (eds Ellis AD, Longland

493 AC, Coenen M and Miraglia N), EAAP Publication 128, 88.

494 Kelso TB, Hodgson DR, Witt EH, Bayly WM, Grant BD and Gollnick PD 1987.

495 Bicarbonate administration and muscle metabolism during high-intensity exercise. In

496 Equine Exercise Physiology 2 (eds Gillespie JR and Robinson NE), pp. 438-447.

497 ICEEP Publications, Davis CA, USA.

498	Lacombe VA, Hinchcliff KW, Geor RJ and Baskin CR 2001. Muscle glycogen
499	depletion and subsequent replenishment affect anaerobic capacity of horses. Journal
500	of Applied Physiology 91, 1782-1790.

Lacombe VA, Hinchcliff KW, Kohn CW, Devor ST and Taylor LE 2004. Effects of feeding meals with various soluble-carbohydrate content on muscle glycogen synthesis after exercise in horses. American Journal of Veterinary Research 65, 916-23.

Lawrence LM, Miller PA, Bechtel PJ, Kane RA, Kurcz EV and Smith JS 1987. The
effect of sodium bicarbonate ingestion on blood parameters in exercising horses. In
Equine Exercise Physiology 2 (eds Gillespie JR and Robinson NE), pp. 448-455,
ICEEP Publications, Davis CA , USA.
Lindgren E 1979. The nutritional value of roughages determined in vivo and by
laboratory methods. Report 45:63, Department of Animal Nutrition and Management,
Swedish University of Agricultural Sciences, Sweden.

512	Lindholm A and Piehl K 1974. Fibre composition, enzyme activity and concentrations
513	of metabolites and electrolytes in muscles of standardbred horses. Acta Veterinaria
514	Scandinavica 15, 287-309.

515 Lowry OH and Passonneau JV 1973. A Flexible System for Enzymatic Analysis. pp.

516 1-291. Academic Press, NY, USA.

517 Luthersson N, Nielsen, KH, Harris P and Parkin TD 2009. Risk factors associated

518 with equine gastric ulceration syndrome (EGUS) in 201 horses in Denmark. Equine

519 Veterinary Journal 41, 625-630.

520 MacLeay JM, Sorum SA, Marsh WE, Sorum MD 1999. Epidemiologic analysis of

521 factors influencing exertional rhabdomyolysis in Thoroughbreds. American Journal of

522 Veterinary Research. 60, 1562-1566.

523 Meyer H 1987. Nutrition of the equine athlete. In Equine Exercise Physiology 2, (eds

524 Gillespie JR and Robinson NE), pp. 644-673. ICEEP Publications, Davis CA, USA.

525	Muhonen S, Lindberg J E, Bertilsson J and Jansson A 2009. Effects on fluid balance
526	and exercise response in Standardbred horses feed silage, haylage and hay.
527	Comparative Exercise Physiology 5, 133-142.
528	NRC (National Research Council) 1989. Nutrient Requirements of Horses. 5th
529	edition. National Academic Press, Washington, DC, USA.
530	NRC (National Research Council) 2007. Nutrient Requirements of Horses. 6th
531	edition. National Academic Press, Washington, DC, USA.
532	Pagan J, Geor RJ, Harris PA, Hoekstra K, Gardner S, Hudson C and Prince A 2002.
533	Effects of fat adaptation on glucose kinetics and substrate oxidation during low-
534	intensity exercise. Equine Veterinary Journal Supplement 34, 33-38.
535	Palmgren-Karlsson C, Lindberg JE and Rundgren M 2000. Associative effects on
536	total tract digestibility in horses fed different ratios of grass hay and whole oats.
537	Livestock Production Science 65, 143-153.

538	Palmgren-Karlsson C, Jansson A, Essén-Gustavsson B and Lindberg JE 2002.
539	Effect of molassed sugar beet pulp on nutrient utilisation and metabolic parameters
540	during exercise. Equine Veterinary Journal Supplement 34, 44-49.
541	Persson SGB 1983. Analysis of fitness and state of training: Evaluation of exercise
542	tolerance and fitness in the performance horse. In Equine Exercise Physiology 1,
543	(eds Snow DH, Persson SGB and Rose RJ), pp. 441-457. Granta Publications
544	Cambridge, UK.
545	Potter GD, Hughes SL, Julen TR and Swinney DDL 1992. A review of research on
546	digestion and utilization of fat by the equine. Pferdeheilkunde Sonderausgabe 119-
547	123.
548	Ragnarsson S and J.E. Lindberg 2008. Nutritional value of timothy haylage in
549	Icelandic horses. Livestock Science 113, 202-208.
550	Ragnarsson S and Jansson A 2011. A comparison of grass haylage digestibility and
551	metabolic plasma profile in Icelandic and Standardbred horses. Journal of Animal

552 Physiology and Animal Nutrition 95(3), 273-279.

Redbo I, Redbo-Torstensson P, Ödberg FO, Hedendahl A and Holm J 1998. Factors
affecting behavioural disturbances in race-horses. Journal of Animal Science 66,
475-481.

- Schuback K, Essén-Gustavsson B and Persson SG 2002. Effect of sodium
  bicarbonate administration on metabolic responses to maximal exercise. Equine
  Veterinary Journal Supplement 34, 539-44.
- 559 Snow DH, Harris RC, Harman JC and Marlin DJ 1987. Glycogen repletion following
- 560 different diets. In Equine Exercise Physiology 2 (eds Gillespie JR and Robinson NE),
- 561 pp. 701-710. ICEEP Publications, Davis CA, USA.
- 562 Tinker MK, White NA, Lessard P, Thatcher CD, Pelzer KD, Davis B and Carme DK
- 563 1997. Prospective study of equine colic risk factors. Equine Veterinary Journal 29,
  564 454-458.
- Voiton DM, Navet R, Lacombe VA, Sluse F, Essen-Gustavsson B, Hinchcliff KW,
  Rivero JLL, Serteyn D and Valberg S 2007. Muscle energetic in exercising horses.
  Equine and Comparative Exercise Physiology 4, 105-118.

568	Waller AP, Geor RJ, Spriet LL, Heighenhasuer GJ and Lindinger MI 2009. Oral
569	acetate supplementation after prolonged moderate intensity exercise enhances early
570	muscle glycogen resynthesis in horses. Experimental Physiology 94, 888-898.
571	Waller A and Lindinger M 2007. The effect of oral sodium acetate administration on
572	plasma acetate concentration and acid-base state in horses. Acta Veterinaria
573	Scandinavica 49, doi:10.1186/1751-0147-49-38.
574	Waters AJ, Nicol CJ and French NP 2002. Factors influencing the development of
575	stereotypic and redirected behaviours in young horses: findings of a four year
576	prospective epidemiological study. Equine Veterinay Journal 34, 572-579.
577	Willard JG, Willard JC, Wolfram SA and Baker JP 1977. Effect of diet on cecal pH
578	and feeding behavior of horses. Journal of Animal Science, 45, 87-93.
579	Williamson A, Rogers CW and Firth EC 2007. A survey of feeding, management and
580	faecal pH of Thoroughbred racehorses in the North Island of New Zealand. New
581	Zealand Veterinary Journal 55, 337-341.

582	Willing B, Vörös A, Roos S, Jones C, Jansson A and Lindberg JE 2009. Changes in
583	faecal bacteria associated with concentrate and forage-only diets fed to horses in
584	training. Equine Veterinary Journal 41, 908-914.
585	Wilson RG, Thornton JR, Inglis S and Ainscow J 1987. Skeletal muscle adaptation in
586	racehorses following high intensity interval training. In Equine Exercise Physiology 2,
587	(eds Gillespie JR and Robinson NE), pp. 367-375. ICEEP Publications, Davis CA,
588	USA.

	Forage		Concentrate
	Diet F	Diet FC	Diet FC
DM	80	78	90
ME	10.4	8.8	11.4
Ash	75	56	37
Crude protein	104	61	174
Neutral detergent fibre	605	600	203
Acid detergent fibre	363	370	117
Lignin	51	69	27
EEC-fat	19	15	60
WSC <sup>2</sup>	79	147	
Free glucose	35	22	3
Free fructose	31	57	0
Fructans	4	51	5
Starch	0	0	358
Yeast	<u>~</u> 2.5	<u>&lt;</u> 3.6	-
Mould	<2.0	<u>&lt;</u> 2	-

## 590 microbial composition (cfu<sup>1</sup>/g fresh matter) of feeds in the experimental diets

<sup>1</sup>Colony-forming units, <sup>2</sup>Water-soluble carbohydrates

- 593 Table 2. Daily feed allowance, feed intake (kg) and nutrient (g) and estimated metabolisable
- 594 energy intake (MJ ME) during 29 days on a high-energy, forage-only diet (F) and a mixed
- 595 forage-concentrate diet (FC) (LSmeans ± SE)

	Diet F	Diet FC
Forage allowance	14.56 ± 0.04	7.00 ± 0.04
Forage intake	10.07 ± 0.09	4.88 ± 0.09
Concentrate allowance	$0.29 \pm 0.03$	7.31 ± 0.03
Concentrate intake	0.29 ± 0.04	7.07 ± 0.04
CP intake <sup>1</sup>	1132 ± 87	1467 ± 66
NDF intake	6588 ± 507	3885 ± 270
Starch intake	0	2503 ± 108
WSC intake	861 ± 66	605 ± 60
Energy intake <sup>2</sup>	110 ± 6	116 ± 6

<sup>1</sup>Corresponds to 113-146% of the requirements for very heavy exercise suggested by NRC
(2007), <sup>2</sup>Corresponds to 90-94% of the requirements for very heavy exercise suggested by
NRC (2007).

- 610 Figure legends
- 611
- 612

613 Figure 1. Venous pH before, during and after an incremental exercise test. Values 614 (LSmeans ± SE) for six Standardbred geldings on a high-energy, forage-only diet (F; 615 diamonds) and a 40:60 forage-concentrate diet (FC; squares). BW=before warm-up, AW=after warm-up, 25W=25 min after warm-up, v6-v9=incremental exercise test at 616 617 velocities 6, 7, 8 and 9 m/s (treadmill incline 6.3%), 5w=after 5 min walk, 10, 25, 55, 618 85 min after the walk. The effect of diet was significant (ANOVA, P=0.0001). \* 619 indicates significant difference (P<0.05) between diets for single samples. 620 621 Figure 2. Plasma glucose<sup>a</sup>, insulin<sup>a</sup>, acetate<sup>a</sup> and non-esterified fatty acid (NEFA) 622 concentrations before, during and after an incremental exercise test. Values 623 (LSmeans ± SE) from six Standardbred geldings on a high-energy, forage-only diet 624 (F; diamonds) and a 40:60 forage-concentrate diet (FC; squares). BW=before warm-625 up, AW=after warm-up, 25W=25 min after warm-up, v6-v9=incremental exercise test 626 at velocities 6, 7, 8 and 9 m/s (treadmill incline 6.3%), 5w=after 5 min walk, 10, 25, 627 55, 85 min after the walk and DA=day after at 11.00. a =no analyses for v6-v8 628 available. The effect of diet was significant for acetate and insulin (ANOVA, P<0.0001). \* indicates significant difference (P<0.05) between diets for single 629

- 630 samples.
- 631







