

Mitochondrial Genetics of Alloplasmic Male-Sterile *Brassica napus* Lines

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Abstract

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Reliable and economical production of hybrid (F_1) varieties requires efficient means to control pollination. The predominant system for pollination control of most field crops today is cytoplasmic male sterility (CMS). Since CMS is a consequence of disturbed nuclear-mitochondrial interactions, the CMS trait is maternally inherited. A common way to produce CMS plants is to combine the nuclear genome from one species with the mitochondria from another; plants of such origin are described as alloplasmic CMS. This thesis describes the production and molecular characterization of a novel alloplasmic CMS system produced from rapeseed (*Brassica napus*) (+) *Arabidopsis thaliana* somatic hybrids.

A population of 170 *B. napus* (+) *A. thaliana* somatic hybrid lines was backcrossed to *B. napus* and 22 lines with male sterility and/or aberrant flower morphology were found. Nine of these were analysed for RFLP and found to contain nuclear and plastid DNA from *B. napus*, whilst the mitochondrial genomes were frequently recombined containing both *B. napus* and *A. thaliana* DNA. Besides the male-sterile trait, the majority of the lines displayed homeotic conversions of anthers to carpeloid organs. Vegetative growth was reduced to some extent in some of the lines. One CMS line segregated both fertile and sterile plants. Fertility co-segregated with molecular markers from *A. thaliana* chr III. By a dihaploidisation strategy we produced plants with a disomic addition of *A. thaliana* chr III, which had a stable inheritance of the fertile phenotype.

Global mitochondrial mRNA expression was analysed in the fertility-restored line, the corresponding CMS line, *B. napus* and *A. thaliana*. Run-on experiments showed that transcriptional activities were highly variable between *B. napus* and *A. thaliana* and that in the CMS line transcriptional activity was reduced for several ribosomal protein genes and increased for *orf139*. Steady-state levels were more homogenous in comparison to transcriptional activities showing that RNA turnover is an important regulatory mechanism. Increased transcript abundance of several genes was observed in the CMS line, often correlated with presence of longer transcripts.

Transcripts of three *A. thaliana* loci, *orf139*, *orf240a* and *orf294* accumulated in the CMS line, but with reduced levels in the restored line. The *orf139* and *orf294* transcripts accumulated differentially in a tissue and genotype-specific manner, while *orf240a* was constitutively expressed throughout the plant. Both *orf240a* and *orf294* transcripts can be polyadenylated, thus providing an explanation for their post-transcriptional regulation. Segregation analysis of sterile and fertile alloplasmic lines indicates that *orf139* and *orf240a* are less likely candidates to be responsible for the male-sterile phenotype, whereas the *orf294* can be CMS-associated. However, it is likely that more than one locus in the *A. thaliana* mt-DNA could encode CMS in the nuclear background of *B. napus*.

Keywords: *Arabidopsis thaliana*, rapeseed, cytoplasmic male sterility (CMS), restorer of fertility, cybrids, mitochondrial gene expression, RNA stability, mitochondrial open reading frames (*orfs*), species-specific factors.

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Abbreviations

List of selected abbreviations used in the text:

ATP	adenosine triphosphate
bp	basepairs
cDNA	complementary DNA
CHA	chemical hybridizing agents
chr	chromosome
CMS	cytoplasmic male sterility
CNM	conserved nonanucleotide motif
cRT-PCR	circular RT-PCR
<i>Fr</i>	fertility-restorer
GISH	genomic <i>in situ</i> hybridisation
GMS	genetic/genic male sterility
kb	kilo basepairs
kDa	kilo Dalton
<i>ms</i>	male-sterility
mt	mitochondrial
NMS	nuclear male sterility
nt	nucleotides
<i>orf</i>	open reading frame
PCR	polymerase chain reaction
PPR	pentatricopeptide repeat
RT-PCR	reverse transcriptase PCR
<i>Rf</i>	restorer of fertility
RFLP	restriction fragment length polymorphism
rRNA	ribosomal RNA
SSLP	simple sequence length polymorphism
tRNA	transfer RNA
UTR	untranslated region

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Papers I-IV

The present thesis is based on the following papers, which will be referred to by their Roman numerals:

I. Leino, M., Teixeira, R. Landgren, M. & Glimelius, K. 2003. *Brassica napus* lines with rearranged *Arabidopsis* mitochondria display CMS and a range of developmental aberrations. *Theoretical and Applied Genetics* 106, 1156-1163.

II. Leino, M., Thyselius, S., Landgren, M. & Glimelius, K. 2004. *Arabidopsis thaliana* chromosome III restores fertility in a cytoplasmic male-sterile *Brassica napus* line with *A. thaliana* mitochondrial DNA. *Theoretical and Applied Genetics* 109, 272-279.

III. Leino, M., Landgren, M. & Glimelius, K. 2005. Alloplasmic effects on mitochondrial transcriptional activity and RNA turnover result in accumulated transcripts of *Arabidopsis orfs* in cytoplasmic male sterile *Brassica napus*. *The Plant Journal*. Published online 31 March 2005. doi: 10.1111/j.1365-313X.2005.02389.x

IV. Leino, M., Sundström, J., Landgren, M. & Glimelius, K. 2005. Expression analysis of the *Arabidopsis* mitochondrial *orf139*, *orf240a* and *orf294* in different tissues of male-sterile and fertile alloplasmic *Brassica napus* lines. (manuscript)

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Introduction

The relationship between pollen and the fertilization of flowers for formation of grains and fruit has probably been known since the early establishment of agriculture. In parallel with the development of more sophisticated agricultural techniques, ways to control pollination have been sought, in order to produce more desirable agricultural products. By 2000BC the Assyrians practiced artificial pollination of date palm trees. The first report of plants with inhibited pollen production is probably Kölreuter's (1763), in which bisexual flowers were described with aborted anthers. In 1877 Darwin proposed male sterility as a strategy to force outbreeding. That male sterility could be the consequence of cytoplasmic genes –“plasmon” was suggested in the beginning of the 20th century when the first examples of non-Mendelian inheritance were described. By studying populations of *Satureja*, *Silene* and *Plantago*, Correns (1904, 1906) demonstrated that gynodioecy could be maternally inherited. The phenomenon of cytoplasmic male sterility (CMS) has been widely exploited for pollination control in the production of hybrid seed since the first report of its use for production of hybrid onion (Jones and Clarke, 1943).

With the development of tools for molecular genetics that arose in the 1970's and 1980's plant development could be examined on a molecular level. Not the least, the development and differentiation of flowers and flower organs in higher plants have been surveyed thoroughly lately. By the identification of meristem identity-genes, ABC-genes and ABC-regulators (reviewed by Coen and Meyerowitz, 1991; Theissen, 2001; Theissen and Saedler, 2001; Lohman & Weigel, 2002) a basic understanding of the genetic control of floral patterning has now been obtained. However, several important pieces of the puzzle are still missing, there among the impact of cytoplasmic genes.

Fertility regulating mechanisms in plant breeding

Production of hybrid seed

Heterosis, or hybrid vigour, was discovered and exploited by corn breeders over a century ago (Shull, 1908). The phenomenon is defined as the increased productivity, speed of development and fertility resulting from differences in parental gametes, although the molecular basis of heterosis is still a matter of debate (reviewed by Birchler *et al.*, 2003). By controlled crosses of inbred parental lines, hybrid (F₁) varieties are produced. The hybrid varieties combine the two desired traits of heterosis and heterogeneity, that otherwise are conflicting breeding objectives when breeding open-pollinating crops. Hybrid varieties are nowadays produced in a large number of crop species, e.g., corn, sorghum, pearl millet, wheat, sunflower, cotton, oil palm, rice, sugar beet, rapeseed and numerous vegetable crops (Maunder, 1999). Reliable and economical hybrid seed production depends on three biological requirements: 1) presence of hybrid vigour, 2) prevention of self-pollination of the female parent, 3) adequate pollination by the male parent (Wright, 1980). The latter two requirements can be referred to as pollination control, for which several different practices have been developed.

Manual, mechanical and chemical pollination control

The simplest way to prevent self-pollination of the female parent is to remove the pollen bearing organs of the plant. This procedure is feasible in monoecious species, like corn and cucumbers, where male and female flowers are positioned on separate parts of the plant. In corn the procedure, called detasseling, can even be mechanised (Wright, 1980). For crop plants with bisexual flowers manual emasculation, in this case removal of anthers, is extremely labourous. Nevertheless, manual emasculation is practised for hybrid seed production of several horticultural crops, e.g. tomato (Kalloo, 1993). Induction of male sterility by chemicals, referred to as gametocides, pollen suppressants or chemical hybridizing agents (CHA) has been known for over 50 years (Moore, 1950). In spite of some use in rice and wheat hybrid production, CHAs are only marginally used, mainly due to incomplete male sterility or to severe damage to the rest of the plant (Tu & Banga, 1998).

Genetical pollination control

In light of the above mentioned obstacles to prevention of self-pollination by manual, mechanical or chemical measures a better alternative is provided by genetical pollination control. This can be classified into two major categories, self-incompatibility and male sterility. Self-incompatibility acts through the inhibition of pollen germination or pollen growth in the style of carpels of the same plant from which pollen is derived. The incompatibility can either be gametophytic where the pollen ability to grow through the stigma is determined by the haploid genotype of the pollen grain, or sporophytic where the ability of pollen to penetrate the stigma is determined by the genotype of the pollen parent. Self-incompatibility is regulated by multiple alleles of single loci. Lately, several of the genes responsible for the reaction have been identified (reviewed by Charlesworth, 2000; Stone and Goring, 2001). Surprisingly, a common mechanism between plant families seems not to be the case. In theory, self-incompatibility provide the breeder with an ideal hybrid system since each plant can serve both as a pollen donor and as a producer of hybrid seed resulting from pollination by another plant. However, the system is not widely used, mainly due to significant difficulties in stabilising and maintaining the parental lines. Exceptions include some vegetable species belonging to *Brassica oleracea* (Gray, 1993; Ockendon & Smith, 1993; Chiang *et al.*, 1993; Crisp & Tapsell, 1993).

Male sterility is defined as the inability to produce functional anthers and/or pollen. The trait can be regulated either by nuclear or cytoplasmic genes. Nuclear male sterility (NMS), also termed genic or genetic male sterility (GMS) is usually caused by recessive alleles (*ms*). Mutations causing NMS are commonly found in nature and a large number of male sterility genes have been found in economically important genera, such as *Zea*, *Lycopersicon*, *Hordeum*, *Pisum*, *Capsicum*, *Gossypium*, *Glycine* and *Oryza* (reviewed by Horner and Palmer, 1995). The drawback with NMS is the impossibility to create 100% male sterile populations. The male sterile plants must be maintained by crossings with an isogenic line heterozygous for the male sterility gene resulting in offspring populations of 50%

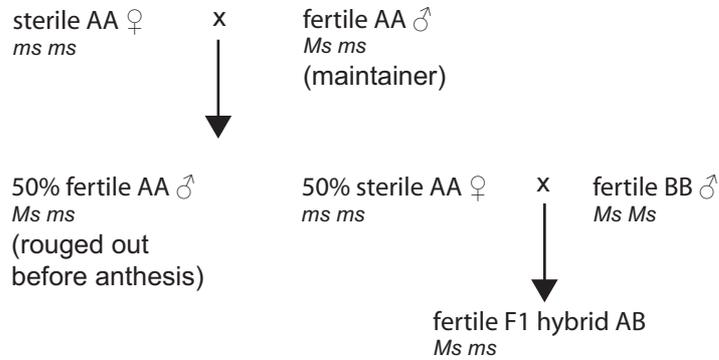
fertile and 50% sterile plants. Thus, in seed production plots the fertile plants have to be rogued out before anthesis (fig 1a). Unless the male sterile gene is linked to some kind of marker gene, these plants are difficult to identify. Even though several types of phenotypic markers, conditional lethal markers and partial male sterilities sensitive to environmental conditions have been produced (reviewed by Krishna Rao *et al.*, 1990) NMS is seldomly used in field scales. An important exception is NMS rice where the male sterility trait is linked to photoperiod-sensitivity (Yuan *et al.*, 1993).

A special form of NMS has been accomplished by genetic engineering. In 1990 Mariani *et al.* produced male sterile *Brassica napus* plants by fusing a tapetum specific promoter to an RNase gene – *Barnase* from *Bacillus amyloliquefaciens* or *RNase T1* from *Aspergillus oryzae*. By linking the male sterility gene to a selectable marker, in this case the *bar* gene that confers resistance to ammonium-glufosinate, plants not expressing the transgene can be eliminated. Thus, by treating the segregating 1:1 population of sterile and fertile plants that occur after crossing with the fertile maintainer line with the herbicide a pure population of sterile plants can be obtained. The system was further developed by the introduction of fertility restorer lines (Mariani *et al.*, 1992). These lines carry the *Barstar* gene under control of the same tapetum specific promoter. The *Barstar* gene, that is an *RNase* inhibitor, has the ability to counteract the effect of the *Barnase* gene so that fertility is restored. The utilisation of the barnase-barstar system in hybrid seed production is illustrated in figure 1b. Male-sterile transgenes developed in other systems promise that the techniques can be applied in more crop species (reviewed by Williams, 1995).

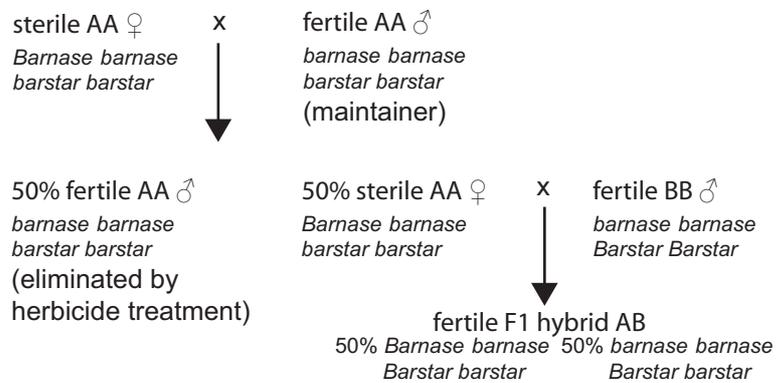
Male sterility can also be caused by cytoplasmic genes and is then termed cytoplasmic male sterility (CMS). CMS can result from spontaneous mutations (autoplasmic CMS) or, more commonly, by the combination of the nucleus from one species with the cytoplasm from another (alloplasmic CMS). CMS has been demonstrated in over 150 plant species (Kaul, 1988). As the cytoplasm in most species is inherited via the eggcell and not from pollen the trait is maternally inherited. This makes CMS extra valuable for production of hybrid seed since crossings with the maintainer line result in 100% sterile plants. CMS was first exploited in the production of hybrid onion (Jones and Clarke, 1943) and has since been extensively used in a number of crop species, including corn, sorghum, pearl millet, sugar beet, sunflower, rice and carrot.

Fertility can be restored by the introgression of specific nuclear genes to plants with CMS-inducing cytoplasm. This is sometimes referred to as genetic-cytoplasmic-male sterility. The nuclear fertility-restoring genes are alternatively symbolised by *Fr* (for fertility-restorer) or *Rf* (for restorer of fertility). For production of hybrid seed in species where the hybrid plants must be fertile, e.g. cereals, the male parent must carry an *Rf* allele. The system to use CMS and *Rf* genes for hybrid seed production is illustrated in figure 1c. In all CMS-inducing cytoplasm investigated so far, the CMS trait has been associated with the mitochondrial genome.

A Nuclear male-sterility (NMS)



B Transgenic Barnase/Barstar system



C Cytoplasmic male-sterility (CMS)

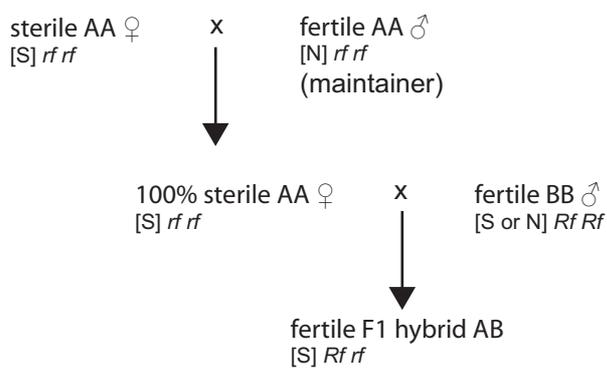


Figure 1. Use of different types of male sterility in hybrid seed production. A and B represents the gametes of two different parental lines. A) Use of NMS governed by the recessive male sterility gene *ms*. B) Use of the transgenic Barnase/Barstar system. C) Use of CMS. The genotype of the cytoplasm, N= normal and S=sterile, and the presence of dominant restorer of fertility, *Rf*, genes are indicated.

Plant mitochondrial genetics

Genome structure and content

The genomes of higher plant mitochondria differ significantly from those of other organisms due to their large size, from 222 kb of the *B. napus* mt-genome (Handa, 2003) to some species within the *Cucurbitaceae* where mt-genomes appear to extend to over 2000 kb (Ward *et al.*, 1981). With the complete mitochondrial genome sequence now available from *A. thaliana* (Unseld *et al.*, 1997), *B. napus* (Handa, 2003), sugar beet – *Beta vulgaris* (Kubo *et al.*, 2000), rice – *Oryza sativa* (Notsu *et al.*, 2002) and corn – *Zea mays* (Clifton *et al.*, 2004) some general features of plant mitochondrial genomes can be listed.

First, gene density must be considered very low. For example, the identified genes of *A. thaliana* account for only 10% of the genome. Introns, duplications, integrations of DNA of nuclear and plastid origin and large unidentified open reading frames account for another 30%. Consequently, 60% of the genome has no known function (Unseld *et al.*, 1997). Second, the gene content (table 1) is highly conserved, although some individual genes have been transferred to the nucleus in some species. Third, the identified genes are often ordered in clusters enabling co-transcription from the same promoter. This clustering has also been experimentally shown for some clusters in *B. napus* (Itani & Handa, 1998) and *A. thaliana* (Brandt *et al.*, 1992). However, the gene clusters are only occasionally conserved between species and the order among clusters usually shifts completely (fig 2). Fourth, each genome contains internal repeats. These repeated sequences could be responsible for recombination and the formation of subgenomic molecules. For example, a tricircular structure is suggested for *B. napus* (Palmer & Shields, 1984) and five circular molecules can be formed in *A. thaliana* (Klein *et al.*, 1994).

All five species have an identical set of 18 genes coding for subunits of complex I, III, IV and V. A pseudogene for the complexII gene *sdh4* is found in *A. thaliana* and *B. napus*, but not in the other species. In the liverwort *Marchantia polymorpha* this gene is intact (Oda *et al.*, 1992). The conserved *orf25* and *orfB* have recently been identified as the ATPsynthase subunits *atp4* (Heazlewood *et al.*, 2003) and *atp8* (Heazlewood *et al.*, 2003; Sabar *et al.*, 2003). Furthermore, six genes for cytochrome c biogenesis are present, some of which have been transferred to the nucleus in some species. Likewise, of the total of 14 genes encoding ribosomal subunits among these mt-genomes, each species has a different set. Noteworthy is the *rps14* gene that is functional in *B. napus*, but only a pseudogene in *A. thaliana*. Several tRNA genes are present, but not always the same ones. In none of the species does the set of tRNA genes account for all 20 amino acids found in proteins. The remaining tRNAs must be imported from the nucleus or in rare cases from the chloroplast (see e.g. Dietrich *et al.*, 1996). The three ribosomalRNA genes *rrn26*, *rrn18* and *rrn5* are found in all genomes, with the last two always in close physical association. Two additional genes, the maturase gene *mat-R* and *orfX* (*mttB*), proposed to encode a transporter protein (Bonnard and Grienberger, 1995) are also found in all species. In conclusion, plant mitochondrial genomes vary significantly in structure, size and sequence, but share a set of very well conserved genes.

Table 1. Comparison of gene content in higher plant mitochondrial genomes, *Arabidopsis thaliana* (*A. t.*), rapeseed – *Brassica napus* (*B. n.*), sugar beet – *Beta vulgaris* (*B. v.*), rice – *Oryza sativa* (*O. s.*) and corn NB – *Zea mays* (*Z. m.*)

Gene	<i>A. t.</i>	<i>B. n.</i>	<i>B. v.</i>	<i>O. s.</i>	<i>Z. m.</i>
NADH dehydrogenases (Complex I)					
<i>nad1</i> *	+	+	+	+	+
<i>nad2</i> *	+	+	+	+	+
<i>nad3</i>	+	+	+	+	+
<i>nad4</i>	+	+	+	+	+
<i>nad4L</i>	+	+	+	+	+
<i>nad5</i> *	+	+	+	+	+
<i>nad6</i>	+	+	+	+	+
<i>nad7</i>	+	+	+	+	+
<i>nad9</i>	+	+	+	+	+
Succinate dehydrogenase (Complex II)					
<i>sdh4</i>	Ψ	Ψ	-	-	-
Apocytochrome b (Complex III)					
<i>cob</i>	+	+	+	+	+
Cytochrome oxidase (Complex IV)					
<i>cox1</i>	+	+	+	+	+
<i>cox2</i>	+	+(2)	+	+	+
<i>cox3</i>	+	+	+	+	+
ATP synthase (Complex V)					
<i>atp1</i>	+	+	+	+	+(2)
<i>atp4 (orf25)</i>	+	+	+	+	+
<i>atp6</i>	+(2)	+	+	+	+
<i>atp8 (orfB)</i>	+	+	+	+	+
<i>atp9</i>	+	+	+	+	+
Cytochrome c biogenesis					
<i>ccmB</i>	+	+	+	+	+
<i>ccmC</i>	+	+	Ψ	+	+
<i>ccmFN</i>	-	-	+	+	+
<i>ccmFN1</i>	+	+	-	-	-
<i>ccmFN2</i>	+	+	-	-	-
<i>ccmFC</i>	+	+	+	+	+
Ribosomal proteins (small subunit)					
<i>rps1</i>	-	-	-	+	+
<i>rps2A</i>	-	-	-	+	+
<i>rps2B</i>	-	-	-	-	+
<i>rps3</i>	+	+	+	+	+
<i>rps4</i>	+	+	+	+	+
<i>rps7</i>	+	+	+	+	+
<i>rps11</i>	-	-	-	Ψ	-
<i>rps12</i>	+	+	+	+	+
<i>rps13</i>	-	-	+	+	+
<i>rps14</i>	Ψ	+	-	Ψ	-
<i>rps19</i>	Ψ	-	-	+	-

Gene	<i>A. t.</i>	<i>B. n.</i>	<i>B. v.</i>	<i>O. s.</i>	<i>Z. m.</i>
Ribosomal proteins					
(large subunit)					
<i>rpl2</i>	+	+	-	+	-
<i>rpl5</i>	+	+	+	+	-
<i>rpl16</i>	+	+	-	+	+
Other proteins					
<i>mat-R</i>	+	+	+	+	+
<i>orfX (mmtB)</i>	+	+	+	+	+
Ribosomal RNAs					
<i>rrn5</i>	+	+	+	+	+
<i>rrn18</i>	+	+	+	+	+
<i>rrn26</i>	+	+	+	+	+
Transfer RNAs					
<i>trnA</i>	-	-	-	-	Ψ
<i>trnR</i>	-	-	-	Ψ	Ψ
<i>trnN</i>	+	+	+	+	+
<i>trnD</i>	+	+	+	+	+
<i>trnC</i>	+	+	+	+	+
<i>trnE</i>	+	+	+	+	+
<i>trnQ</i>	+	+	+	+	+
<i>trnG</i>	+	+	+	-	-
<i>trnH</i>	+	+	+	+	+
<i>trnI</i>	+	+	+	+	+
<i>trnK</i>	+	+	+	+	+
<i>trnL</i>	-	-	-	-	Ψ
<i>trnM</i>	+	+	+	+	+
<i>trnJ</i>	+	+	+	+	+
<i>trnF</i>	-	-	+	+	+
<i>trnP</i>	+	+	+	+	+
<i>trnS</i>	+	+	+	+	+
<i>trnW</i>	+	+	+	+	+
<i>trnY</i>	+	+	+	+	+
<i>trnV</i>	-	-	Ψ	Ψ	Ψ

* trans-spliced . (+) present, (-) absent, (Ψ) pseudogene. Numbers in parenthesis indicate if more than one gene copy exists.

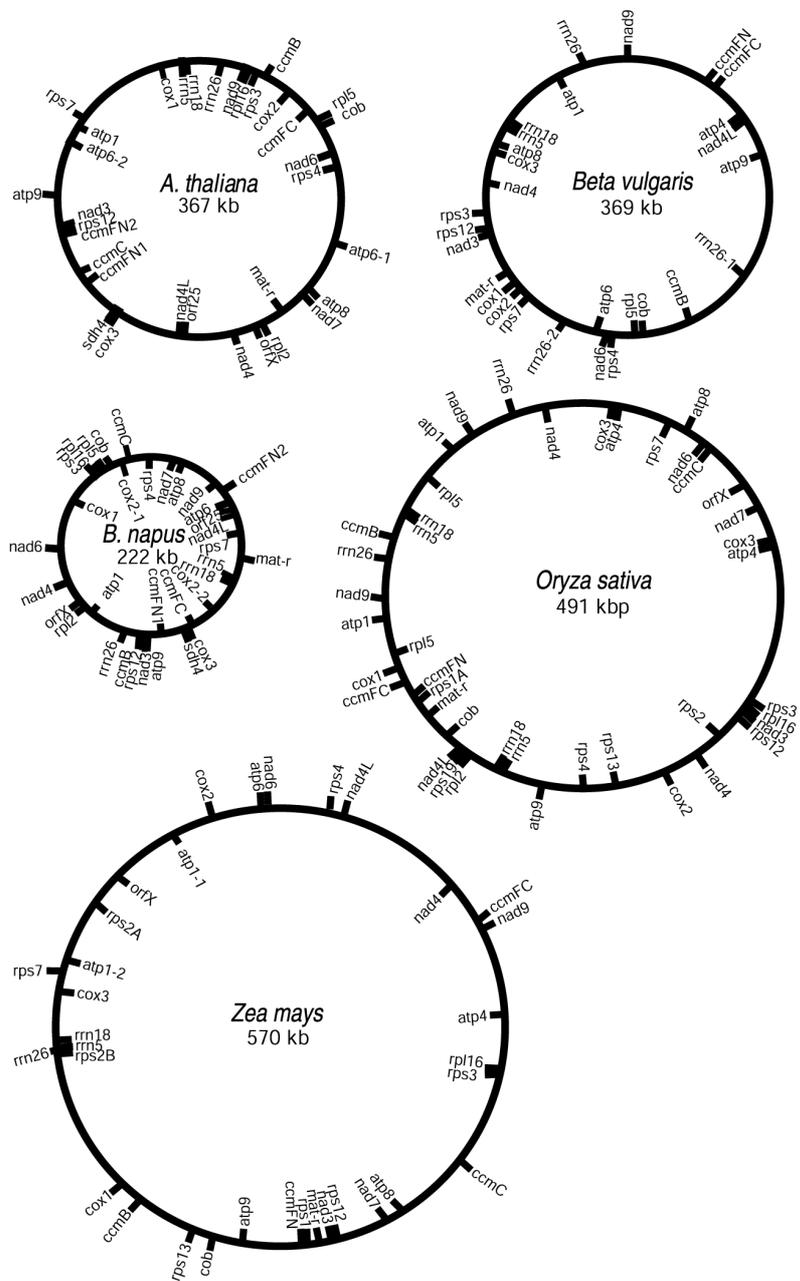


Figure 2. Circular maps of higher plant mt-genomes. For simplicity only protein coding genes and rRNA genes are indicated and trans-spliced exons omitted.

Regulation of gene expression

In addition to, or perhaps as a result of the complex mt-genomes in plants, regulation of gene expression has unique features. The multiple steps of transcriptional, post-transcriptional and translational control have been reviewed by Binder *et al.* (1996), Giegé and Brennicke (2001), Hoffmann *et al.* (2001) and Binder and Brennicke (2003).

The large physical distances between genes suggest that several promoters are required for transcription. From run-on experiments (Finnegan & Brown, 1990; Mulligan *et al.*, 1991; Muise and Hauswirth, 1992; Giegé *et al.*, 2000) it is also clear that promoters have different strength, since differences in transcriptional activity are found for individual genes. By *in vitro* capping experiments with the enzyme guanylyltransferase, transcription initiation sites and corresponding promoter sequences can be identified (see e.g. Covello & Gray, 1991; Brown *et al.*, 1991; Binder & Brennicke, 1993). A motif of four nucleotides, CRTA, where R represents either a guanine or an adenine, is common to promoters identified this way in both monocots and dicots. By *in vitro* transcription studies the importance of nucleotides surrounding this motif has been further investigated (Binder *et al.*, 1995; Dombrowski *et al.*, 1999; Hoffmann & Binder, 2002). The region from at least 14 bases upstream and 4 bases downstream of the transcription initiation site seems to be involved. The combined *in vitro* capping and *in vitro* transcription studies have defined a dicot consensus nonanucleotide motif (CNM) promoter sequence, CRTAAGAGA. Searches for this sequence (including some variations) in the mt-genomic sequences of *A. thaliana* (Dombrowski *et al.*, 1998) and *B. napus* (Handa, 2003) resulted in two major conclusions: About half of the genes lacked this promoter motif and the locations of the CNM for specific genes are not always conserved between the two species. Consequently, there must be other, yet unidentified, promoter motifs. Recently, Kühn *et al.* (2005) identified a large number of *A. thaliana* promoters which differ from the CNM. Additionally, they clearly showed that multiple promoters for a single gene are often present. Although RNAPolymerases in plant mitochondria have been identified (Hedke *et al.*, 1997, 2000) cofactors mediating transcription initiation are still unknown. Cofactors have been shown to be species-specific in corn/teosinte (Newton *et al.*, 1995) and *Nicotiana* (Edqvist & Bergman, 2002). In both these cases particular promoters require a specific nuclear background for transcript initiation.

Transcription is only the first step in determining the steady-state population of translatable RNA. Equally important are post-transcriptional events. For example, transcript mapping shows that 5' and 3' processing most often takes place in plant mitochondrial transcripts. The enzymes responsible for processing are currently unknown, although conserved sequence elements marking processing sites have been suggested by Schuster & Brennicke (1989). In transcript 3'ends stem-loop structures are regularly found. In the pea *atp9* transcripts, such structures were found to act as processing signals by impeding exonucleolytic degradation from 3'ends (Dombrowski *et al.*, 1997). Stem-loop structures in Ogura CMS *orf138* transcript 3' regions were found to play roles both as processing and stabilizing signals (Bellaoui *et al.*, 1997). Recently it has been discovered that

polyadenylation of 3' ends results triggers degradation by a 3' to 5'-exoribonuclease activity. (Gagliardi and Leaver, 1999; Lupold *et al.*, 1999; Gagliardi *et al.*, 2001; Kuhn *et al.*, 2001; Perrin *et al.*, 2004a; Perrin *et al.*, 2004b). In *A. thaliana* two exoribonucleases, AtmtPNPase and AtRNaseII, have been cloned (Perrin *et al.*, 2004a). Mutants of AtmtPNPase result in accumulation of pretranscripts and polyadenylated transcripts (Perrin *et al.*, 2004a; 2004b).

Group II introns are found in many protein coding genes but vary from species to species (Binder *et al.*, 1996). The spliceosome and potential cofactors have not been identified. Trans-splicing of the exons of *nad1*, *nad2* and *nad5*, spread all over the genome, occur in every flowering plant hitherto examined (Malek *et al.*, 1997). A unique feature of plant mitochondrial gene expression is RNA editing, first detected by Covello & Gray (1989), Hiesel *et al.* (1989) and Gualberto *et al.* (1989). The editing events change Cs to Us in the RNA. Giegé & Brennicke (1999) undertook a global investigation of editing sites in *A. thaliana* mitochondria. A total of 441 editing sites was found, mostly located in coding regions, some in introns and leading or trailer sequences, but extremely rarely found elsewhere. In a similar investigation performed in *B. napus* Handa (2003) discovered 427 sites, 81% of which were common for both species. Compared to the average DNA sequence similarity for protein coding genes of 99.2%, diversification of editing seem higher than coding information. In neither of these surveys of editing sites, could a common sequence element for editing be found. Electroporation experiments indicate that cis-elements, 20 nt upstream of the editing site could have a function (Farré *et al.*, 2001). The role of editing for gene regulation has been extensively examined. The only clear role of editing to date is for a tRNA, encoding phenylalanine in *Oenothera*, which requires editing to excise from its precursor molecule (Marchfelder *et al.*, 1996). Mitochondrial gene expression is probably also regulated on a translational level, however the mechanisms regulating translation of transcripts are virtually unknown.

Cytoplasmic male sterility

CMS-associated genes

Molecular studies of CMS plants have correlated the trait with expression of putative genes in the mitochondrial genome (reviewed by Schnable & Wise 1998; Budar & Pelletier, 2001; Budar *et al.*, 2003; Hanson & Bentolila, 2004). Although mitochondrial CMS-associated genes have been suggested in numerous systems clear evidence for a link to the male-sterile phenotype is often lacking. However, in some cases, shown in figure 3, strong correlations between the gene and the CMS-phenotype have been demonstrated.

Of the CMS-associated loci hitherto found, some shared similarities are worth noting. First, open reading frames (*orfs*) are found, comprised of novel sequences of unknown origin combined with sequences of known mitochondrial genes. Examples include the *Brassica pol* and *nap orf224* and *orf222*, which contain parts of the *atp8* gene (Singh & Brown, 1991, Brown, 1999), as does the sunflower PET1 *orf522* (Laver *et al.*, 1991), the Sorghum A3 *orf107*, which contains a piece of *atp9* (Tang *et al.*, 1996), the rice Boro *orf79* (Akagi *et al.*, 1994) and wheat

thimophevi orf256 (Rathburn & Hedcoth, 1991), which contain small fragments of *cox1* and the *urf13-T* in corn CMS-T, which contains a small fragment from *rrn26* (Dewey *et al.*, 1986). Other loci are even more complex, such as the *Brassica tour* orf263 which contains parts of *nad5a* and *atp6* (Landgren *et al.*, 1996), the *Petunia pcf* (orf402) which contains parts of *atp9* and *cox2* (Young and Hanson, 1987), and the orf77 of corn CMS-S, which contains several small pieces from the *atp9* gene (Zabala *et al.*, 1997).

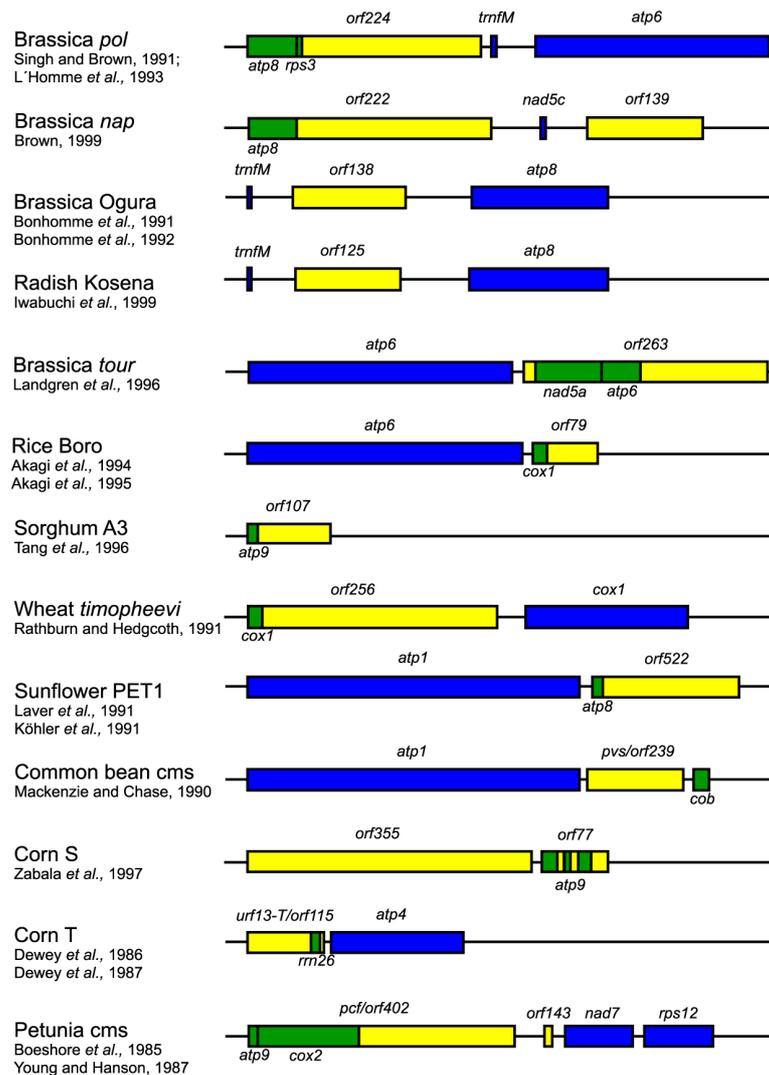


Figure 3. Genes associated with CMS and corresponding references. CMS-genes are denoted by the current convention of number of codons, with the exception of the historical convention of sunflower *orf522* that rather encodes 522 nucleotides. The alternative names for common bean *orf239*, corn T *orf115* and petunia *orf402* are also indicated. Boxes indicate *orfs*. Blue indicates standard mt-genes. Yellow indicates CMS-associated genes with fragments of standard genes indicated by green.

A second common feature is close physical association to, and in some cases co-transcription with, standard mitochondrial genes. In fact, a common strategy for identifying CMS-associated loci is to look for aberrant transcription patterns in Northern analysis using standard mt-gene probes. Surprisingly often, CMS-associated loci are found close to ATP synthase subunit genes. This is the case for corn CMS-T, in which the *urf13-T* is positioned upstream of *atp4* (Dewey *et al.*, 1986, 1987), the *Brassica pol orf224* positioned upstream of and co-transcribed with the *atp6* gene (Singh and Brown, 1991), the *Brassica Ogura orf138* (Bonhomme *et al.*, 1991, 1992) and the radish Kosena *orf125* (Iwabuchi *et al.*, 1999) which are positioned upstream of *atp8* and co-transcribed with this gene, the *Brassica tour orf263* (Landgren *et al.*, 1996) and rice Boro *orf79* (Akagi *et al.*, 1994) which are positioned downstream of and co-transcribed with *atp6*, and the sunflower PET1 *orf522* which is positioned downstream of and co-transcribed with *atp1* (Köhler *et al.*, 1991). Apart from these examples, the wheat *thimophevii orf256* is located upstream of and co-transcribed with *cox1*. The *orf77* of corn CMS-S is also closely positioned to *cox1* but do not appear co-transcribed with it (Zabala *et al.*, 1997). The *pvs* locus in *Petunia* is positioned upstream of and co-transcribed with both *nad3* and *rps12* (Pruitt and Hanson, 1991).

Besides these two common, but not compulsory, requisites CMS loci have few other features in common. The parts of the *orfs* not containing fragments from standard mt-genes have limited sequence similarity with the exception of some pairwise similarities of *Brassica pol orf224* and *nap orf 222*, the *Brassica Ogura orf138* and radish Kosena *orf125*, and the rice Boro *orf79* and sorghum A3 *orf107*.

Since efficient methods for transforming mitochondria are still lacking the ultimate proof for a role of CMS-associated genes remains to be demonstrated. Attempts to provide proof by transforming plants with CMS-associated *orfs* linked to a mitochondrial targeting sequence have been made. Chaumont *et al.* (1995) transformed tobacco with the *urf13* from CMS-T corn, but did not obtain male-sterile plants. Wintz *et al.* (1995) transformed *Petunia* with the CMS-associated *pcf* gene, but did neither obtain male-sterile plants. He *et al.* (1996) managed to obtain sterile tobacco plants upon transformation with the common bean *pvs* CMS-associated gene. However, male-sterility was found independent whether or not the gene was mitochondrially targeted. All in all, experiments with nuclear transformation of CMS-associated genes must be considered inconclusive.

An interesting observation from the fully sequenced mt-genomes of *A. thaliana* (Unselde *et al.*, 1997) and corn (Clifton *et al.*, 2004) is that mitochondrial genomes of fertile plants contain many more *orfs* with properties typical for CMS than would occur by random. In the *A. thaliana* mt-genome Marienfeld *et al.* (1997) found 8 *orfs* with a mosaic structure involving fragments of known genes. An interesting hypothesis (see e.g. Budar & Pelletier, 2001) is that these *orfs* can cause CMS, but are suppressed by nuclear regulators (*Rf* genes). In alloplasmic plants the putative CMS-genes are exposed to a nuclear background, which has not evolved or maintained the corresponding restorer genes. Consequently, this would allow the CMS gene and phenotype to be expressed.

Nuclear fertility-restorer genes

That CMS is the result of disturbed nuclear-mitochondrial interactions is clearly demonstrated by the restoration of flower morphology and fertility through introgression of certain nuclear restorer of fertility – *Rf* genes. In the case of alloplasmic CMS *Rf* genes are often found in the nuclear genome of the cytoplasmic donor species. Restored lines can thus be produced by introgression of parts of nuclear DNA from this species to the CMSline. Restoration has been accomplished in, for example, *Nicotiana* alloplasmic CMS lines (Burns *et al.*, 1978; Gerstel *et al.*, 1978), wheat with *T. thimophevii* cytoplasm (Livers, 1964) and numerous *Brassica juncea* interspecific hybrids (Prakash *et al.*, 2001; Pathania *et al.*, 2003; Banga *et al.*, 2003).

Rf genes have been demonstrated to affect the CMS-associated genes by several different modes of action. In common bean the *Fr* locus has the ability to shift the stoichiometric relationship in the tripartite mt-genome so that the molecule containing the CMS-associated *pvs* gene is suppressed to substoichiometric levels (Mackenzie and Chase, 1990; Janska *et al.*, 1998; Sarria *et al.*, 1999). Therefore, transcription of the *pvs* gene is indirectly reduced as there are fewer DNA molecules encoding it. This mechanism has not been reported from any other CMS-system, although a nuclear influence on mitochondrial genome structure has been reported in, for example, *Nicotiana* (Håkansson and Glimelius, 1991) and *A. thaliana* (Abdelnoor *et al.*, 2003). An influence of *Rf* genes on transcriptional level is more common. In corn the non-allelic restorer genes *Rf1*, *Rf8* and *Rf** all have the ability to process the CMS-associated T-*urf13* transcript (Dill *et al.*, 1997; Wise *et al.*, 1999). Likewise, in Brassica with *pol* and *nap* cytoplasm the allelic genes *Rfp* and *Rfn* have the ability to process the *orf224-atp6* and *orf222-nad5c-orf139* co-transcripts, respectively (Li *et al.*, 1998; Menassa *et al.*, 1999; Brown *et al.*, 1999). The *Rf3* gene in sorghum (Tang *et al.*, 1996) and *Rf3* gene of corn CMS-S (Wen & Chase, 1999) also appear to process CMS-associated transcripts. Interestingly, these *Rf* loci often also process other mitochondrial transcripts.

Other modes of restorer action have been reported. In a restored CMS-line of *Nicotiana tabaccum* with *N. repanda* cytoplasm the transcript initiation site of the CMS-associated locus is affected (Edqvist and Bergman, 2002). Restorer effects on editing have been reported for *atp6* transcripts of rice Boro CMS (Iwabuchi *et al.*, 1993) and sorghum A3 (Howad & Kempken, 1997) but the relationship of editing to fertility restoration is unclear. In sterile and fertility-restored lines of sunflower PET1, transcripts are identical but preferably degraded in the restored line due to polyadenylation of 3'ends (Monéger *et al.*, 1994; Gagliardi & Leaver, 1999). Finally, restorer genes can also act on the translational or post-translational level and inhibit accumulation of the CMS-associated protein. This action of restorer genes has been demonstrated in the similar *Brassica* Ogura (Bellaoui *et al.*, 1999) and Radish/*Brassica* Kosena (Koizuka *et al.*, 2000) CMS systems.

The molecular identity of *Rf* genes was elusive until 1996, when the *Rf2* gene restoring CMS-T corn was identified as an aldehyde dehydrogenase that assembles in the mitochondrial matrix (Cui *et al.*, 1996; Liu *et al.*, 2001). If this gene is a true

restorer gene has been debated (Touzet, 2002; Schnable, 2002). First, *Rf2* does not affect the chimeric gene or its protein. Second, *Rf2* has an important physiological role also in plants with normal fertile cytoplasm, as heterozygous *rf2* plants have impaired anther formation. Lately, four more restorer genes, that do affect levels of the CMS-associated protein, have been cloned in a *Petunia* CMS system (Bentolila *et al.*, 2002), in rice Boro (Kazama and Toriyama, 2003; Komori *et al.*, 2004), in *Brassica* Ogura (Brown *et al.*, 2003; Desloire *et al.*, 2003), and *Raphanus/Brassica* Kosenka (Koizuka *et al.*, 2003). The latter two are identical. All these genes bear a PPRmotif (pentatricopeptide repeat), a gene family recently characterized by Lurin *et al.* (2004) of organellar targeted, and probably RNA binding, proteins. In *A. thaliana* 441 PPR genes have been found to date and the gene family is probably well represented also in other plant species. Interestingly, the restorer genes cloned so far, all belong to the same subgroup of PPRgenes and have similar motifs.

Mechanisms of CMS

In spite of the progress made in identifying mitochondrial CMS-loci and nuclear restorer genes, the interactions leading to male sterility remains puzzling. The translation of *Brassica* Ogura *orf138* yields a 19kD protein (Grelon *et al.*, 1994), the Radish Kosenka *orf125* is translated into a 17 kDa protein (Iwabuchi *et al.*, 1999), the wheat *thimophevii orf 256* to a 7 kDa protein (Song & Hedgcoth, 1994), the sunflower PET1 *orf522* to a 15-16 kDa protein (Horn *et al.*, 1991; Monéger *et al.*, 1994), the common bean *pvs* to a 27 kDa protein (Abad *et al.*, 1995), the corn CMS-T *urf13* to a 13 kDa protein (Wise *et al.*, 1987) and the *Petunia pcf* to a 19.5 kDa protein (Nivison & Hanson, 1989). These proteins are generally hydrophobic, suggesting association with mitochondrial membranes. The membrane association has indeed been shown for the *Brassica* Ogura ORF138 (Grelon *et al.*, 1994), the sunflower ORF522 (Horn *et al.*, 1996), and the corn CMS-T URF13 (Dewey *et al.*, 1987). Expression of URF13 is also associated with susceptibility to toxin from the corn fungal pathogen *Cochliobolus heterostrophus* (reviewed by Levings, 1993). In presence of the toxin the protein forms a pore in the mitochondrial membrane. Whether this pore, is formed also in sterile anthers is not known. Interestingly, expression of a novel mitochondrial gene due to altered processing in *Citrus jambhiri* also leads to membrane pore formation and susceptibility to the fungal pathogen *Alternaria alternata* (Ohtani *et al.*, 2002). The membrane association and/or the co-transcription with subunits of ATP synthase (Complex V) could also suggest disturbed ATP production. In an alloplasmic *Nicotiana* CMS system reduced ATP/ADP ratios have been observed in flower buds in the sterile line (Bergman *et al.* 2000). In the alloplasmic CMS-lines of *Brassica napus* described in this thesis, Teixeira *et al.* (2005a) found lower ATP levels in comparison to the fertile line. Lowered ATP synthase activity has also been reported in sunflower PET1 (Sabar *et al.*, 2003). Thus, impaired energy production could be one explanation for the developmental aberrations. Balk & Leaver (2001) showed that programmed cell death (PCD) occurs in premature tapetal cells of sunflower PET1 developing anthers. The authors propose that insufficient energy access may activate the PCD-associated proteins. However, why lowered ATP levels should specifically disturb pollen development and not other energy demanding processes, like germination, remains mysterious.

CMS-associated polypeptides are generally constitutively expressed, with a noteworthy exception: In common bean, the PVS-ORF239 proteins accumulate in microspore cell walls and callose layers and seem to toxify microspore development (Abad *et al.*, 1995; Sarria *et al.*, 1998). Restorer genes have also been shown to act in a tissue-specific manner. For example, the *Rfp* restorer of Brassica *pol* CMS processes the *orf224* more efficiently in the inner three whorls of the flower (Menassa *et al.*, 1999), the *Rfn* of Brassica *nap* CMS reduces the *orf222* in developing anthers (Geddy *et al.*, 2005) and the *orf522* of PET1 sunflower is most efficiently degraded in floral tissue (Monéger *et al.*, 1994). Although the general feature of CMS is inhibited pollen production, a wide range of different morphological aberrations is found among different CMS systems. In some systems, such as *Petunia*, sunflower PET1 and corn CMS-T, only the pollen production in anthers is affected, whereas in other systems stamens undergo homeotic conversions of floral organs. In Brassica Ogura, especially grown in cold temperatures, conversions of stamens to carpeloid structures were found (Polowick & Sawhney, 1987), in Brassica *juncea* with *Enarthrocarpus lyratus* cytoplasm petaloid anthers are formed (Banga *et al.*, 2003), in *Nicotiana* interspecific hybrids or cybrids a range of flower modifications occur (Gerstel *et al.*, 1978; Kofer *et al.*, 1991, Farbos *et al.*, 2001), in CMS carrot both petaloid and carpeloid stamens are formed (Linke *et al.*, 1999) and in CMS wheat pistilloid stamens are found (Murai *et al.*, 2002). Interestingly, the homologues of the floral homeotic gene APETALA3 in *A. thaliana* conferring anther identity (Coen and Meyerowitz, 1991) are found affected in several CMS-systems (Murai *et al.*, 2002; Linke *et al.*, 2003; Geddy *et al.*, 2005; Teixeira *et al.*, 2005b). The link between CMS-associated genes, their products, mitochondrial function and pollen/flower formation, however, remains unclear.

Aims of the study

The overall aim of the project was to study the genetic mechanisms involved in the nuclear-mitochondrial interaction leading to CMS in *Brassica napus* (+) *Arabidopsis thaliana* somatic hybrids. Specific aims were to:

- produce stable *B. napus* CMS lines from a population of *B. napus* (+) *A. thaliana* somatic hybrids and to characterize genotypes and phenotypes of these lines;
- produce stable restorer of fertility lines through introgression of nuclear *A. thaliana* DNA and to characterize the nuclear DNA involved in fertility restoration;
- develop methods for high-throughput expression profiling of mitochondrial genes;
- analyze species-specific effects on mitochondrial transcriptional and post-transcriptional processes;
- analyze developmental and molecular regulatory mechanisms of putative CMS-associated genes.

Results and discussion

Establishment of a CMS-system (I, II)

Forsberg *et al.* (1998) performed protoplast fusions between *Brassica napus* and *Arabidopsis thaliana*. From these experiments approximately 200 lines derived from individual calli were obtained. After a backcross to *B. napus* a BC₁ population of 170 lines was screened for plants with male sterility and/or aberrant flower morphologies. Twenty-two lines were identified. Nine lines with good vigour and seed set were selected and backcrossed further to obtain a BC₃ generation. These lines were characterized with RFLP and specific nuclear and organellar probes and found to have nuclear and plastidic DNA from *B. napus*, whilst mitochondrial DNA was recombined with varying portions of *A. thaliana* and *B. napus* mtDNA.

Male-sterile plants could be classified into two major categories, one group with stamens of reduced size and limited pollen production and one group with stamens converted into feminized organs, including ovules and stigmas, that were completely unable to produce pollen. Both groups displayed reduced petal size, which was especially true for the second group. The first group shared clear morphological similarities with the *nap*, *pol* (Fan & Stefansson, 1986) and *tour* (Liu *et al.*, 1996) CMS-inducing cytoplasms in *B. napus* whereas the second group resembles cold-grown plants of the original type of Ogura CMS in *B. napus* (Polowick and Sahwney, 1987). The second group was more common among the screened lines. Detailed investigations of two lines in group two (4:19 and 41:17) showed that morphological differences of flower meristems can be noticed from stage 4 and onwards during flower development (Teixeira *et al.* 2005b). Interestingly, the floral homeotic genes APETALA3 and PISTILLATA, responsible for proper stamen and petal development are downregulated in these lines (Teixeira *et al.* 2005b, Carlsson *et al.*, manuscript in preparation). Some reduction in vegetative development and later flowering time were also observed in these lines. The phenotypes were stably maternally inherited between generations and have to date been backcrossed for 12 generations without any noticeable shift in phenotype.

In the first backcrossed generation of CMS-line 4:19, one plant with partially restored male-fertility was observed. This line was subjected to recurrent backcrosses with the maintainer line *B. napus* cv. Hanna and analysed for inheritance of the restoration trait. The restorer trait appeared not to be inherited in a Mendelian ratio, nor was it possible to stabilise the trait via selfings. In a segregating population mapping with RFLP markers was performed. These results showed that restoration co-segregated with markers covering the whole *A. thaliana* chromosome III. From these results we concluded that the restored plants possessed a monosomic addition of the foreign chromosome. Thus, we utilized a dihaploidisation strategy to stabilize the line. Haploid plants were produced from microspore cultures, screened with SSLP-markers for presence of the *A. thaliana* chromosome and doubled with colchicine. One dihaploid line had a restored phenotype that was stably inherited after selfing and by GISH we showed that this

line possessed a disomic addition of the *A. thaliana* chromosome III. The *A. thaliana* chr III contains over 5000 genes (The Arabidopsis Genome Initiative, 2000), but some putative candidate loci for the restorer gene/s could be mentioned. PPR genes are to date the best candidate family for restorer genes. On chr III 94 PPR genes can be identified, out of which 48 are predicted by Predotar (Small *et al.*, 2004) to be targeted to mitochondria (Lurin *et al.*, 2004). At least one of these genes shares high homology with the cloned restorer genes from *Petunia* and Ogura/Kosena radish (Desloire *et al.*, 2003). The *CHM* gene, affecting stoichiometric shifting in the *A. thaliana* mitochondrial genome is also positioned on chr III (Abdelnoor *et al.*, 2003). Additionally, Elo *et al.* (2003) have performed bioinformatic studies revealing a large gene cluster of putative mitochondrial DNA and RNA binding proteins on chr III.

The restored line, the CMS line, and the two parental species have served as research material (fig. 3) in this thesis. As the full mitochondrial genome sequence from both *B. napus* (Handa, 2003) and *A. thaliana* (Unseld *et al.*, 1997) as well as the full nuclear genome sequence of *A. thaliana* (The Arabidopsis Genome Initiative, 2000) are available the material is especially suitable for mitochondrial genetic studies. In addition, the finding of several lines with different male sterile or aberrant flower morphologies and heterogenous mitochondrial genomes offers the possibility to correlate certain flower abnormalities with a certain mitochondrial DNA composition.

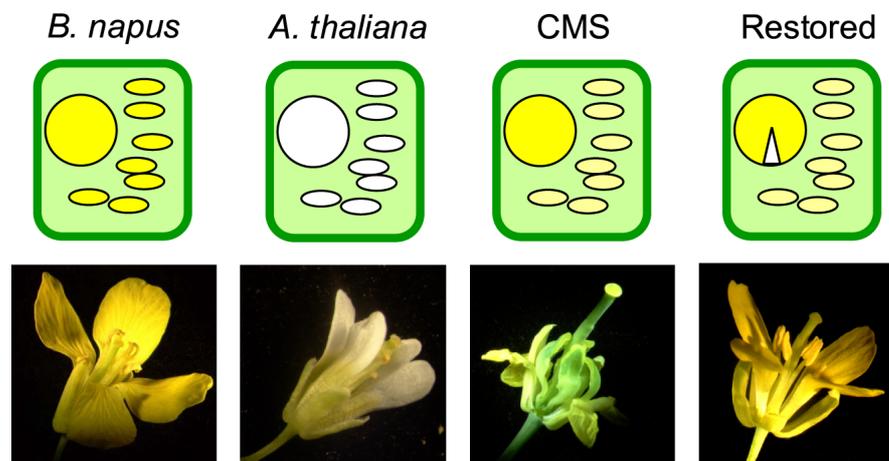


Figure 4. Schematic presentation of the nuclear-mitochondrial composition in the plant material used in this thesis. Note that *B. napus* (maintainer line) and the CMS line are isogenic in respect of the nuclear genome, whilst the CMS and restored lines are isogenic in respect of mitochondrial genomes.

Mitochondrial DNA composition (I, III)

Nine of the CMS-lines were characterized for mitochondrial genome composition using RFLP analysis. Almost any gene probe - restriction enzyme combination resulted in polymorphism between *A. thaliana* and *B. napus*, a finding reflecting the large differences in non-coding sequence and structure between the two species. The RFLP pattern for each CMS line was compared to the parental lines to determine from which parental species a specific gene region was derived (fig 5a). This methodology is not foolproof as fragments visually estimated to be of the same size could be wrongly interpreted to be similar. However, two general conclusions could be drawn from this investigation. The CMS lines contain both *A. thaliana* and *B. napus* mtDNA and frequent recombinations have occurred. In fact none of the nine lines showed an identical RFLP patterning. Of special interest is the pattern obtained with the *cox2* gene probe, for which a very high degree of polymorphism between lines was found (fig 5b). The *cox2* gene is located in the repeat regions in *B. napus* (Handa, 2003) and close to repeat I in *A. thaliana* (Unselde *et al.*, 1997). The repeat regions could be involved in intragenomic recombination and the formation of subcircular molecules. Whether novel subcircular molecules also formed when the two species were combined is not known, but all the same quite possible. In the presence of the *A. thaliana* chr III in the restored line a shift in genome structure in this region was observed. Nuclear influence on genome structure has also been reported in alloplasmic *Nicotiana* lines (Håkansson and Glimelius, 1991) and in CMS lines of common bean (Janska *et al.*, 1998). In *A. thaliana* a gene influencing genome structure, CHM, has been cloned (Abdelnoor *et al.*, 2003). This gene is indeed positioned on chr III, but it is not known whether this gene is specifically responsible for the structural shifts observed in the *cox2* region.

Novel recombined hybrid-specific fragments were found occasionally. In the investigation of the nine CMS lines with ten gene probes recombinations were found, in one or more lines, with the probes *atp1*, *atp9*, *cob*, *cox2* and *nad5a*. Three of the lines, 4:19, 41:17 and 14:103, were analysed for RFLP and 34 gene probes (III and unpublished results). Additional recombined fragments were found, in one or more lines with the probes *nad1*, *cox3*, *atp6*, *ccmB*, *rpl16*, *rps7*, *rps3*, *rps4*, *rps12* and *orfX*. In the extensive analysis of mitochondrial recombinations after somatic hybridisation between different Brassicaceae species performed by Landgren and Glimelius (1990, 1994) and Landgren *et al.* (1994) hotspots for recombinations were found, for example, with *atp1* and *atp9*. As suggested by Clark *et al.* (1986) the recombination events could preferably take place at sites of short repeated sequences. In fact, both the *B. napus* (Handa, 2003) and *A. thaliana* mt-genomes (Unselde *et al.*, 1997) are filled with short repeats. We have not sequenced the rearranged fragments, but with the full genome sequences available this is easily feasible and could determine if a certain particular sequence motif facilitates recombination.

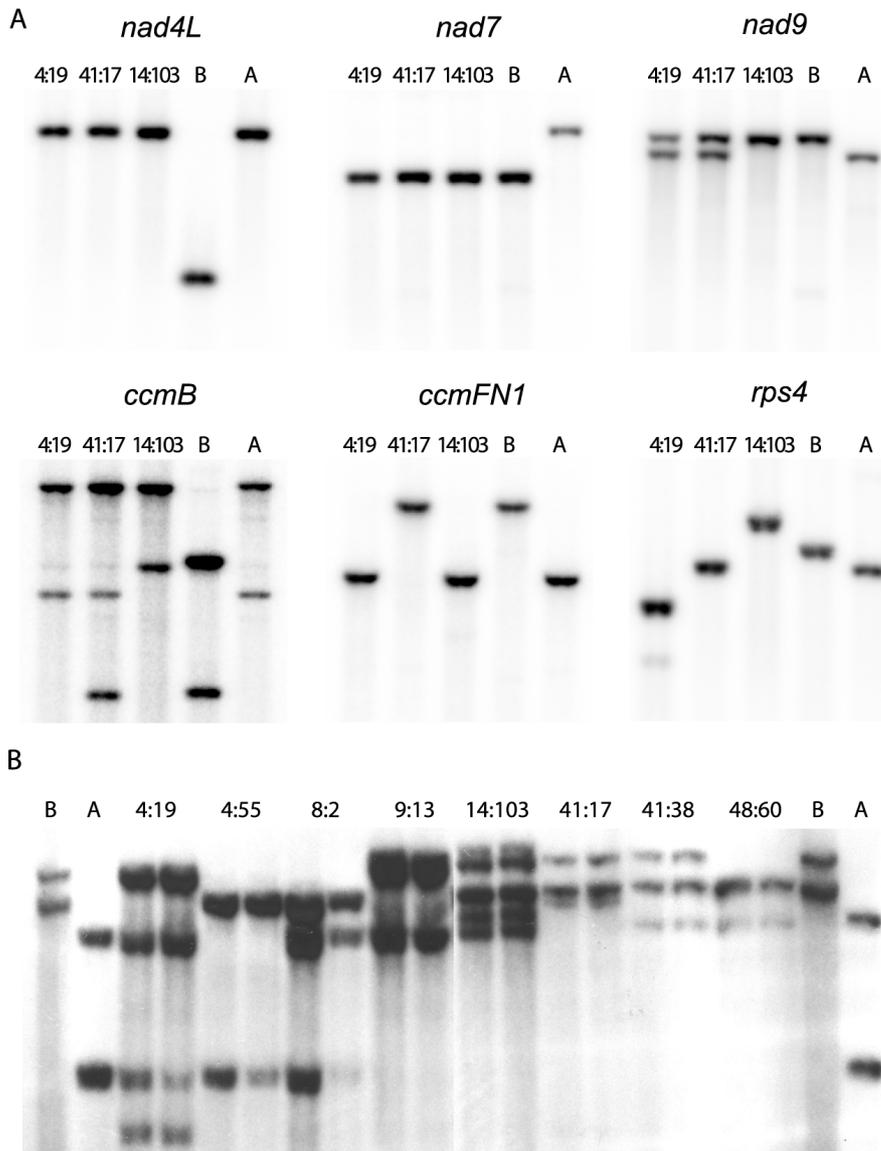


Figure 5. Examples of RFLP hybridisation patterns of the parental and different CMS lines obtained with different gene probes. (a) The three different CMS lines 4:19, 41:17 and 14:103 and the two parental species, *B. napus* (B) and *A. thaliana* (A). Notice presence of gene copies from either or both parental species in the CMS lines as well as recombined fragments. (b) RFLP hybridization pattern obtained with the gene probe *cox2* from eight different CMS lines. DNA was analysed from two individual plants of each line.

Species-specific control of gene expression (III)

To investigate mitochondrial gene expression and RNA turnover in the different lines a macroarray-system was set up. The system is based on the methods described by Giegé *et al.* (2000), but refined and supplemented by additional gene probes. Using sequence specific primers, sequences were PCR amplified from all identified protein coding and rRNA coding genes as well as several *orfs* in the *A. thaliana* mitochondrial genome. The PCR products were dotted onto nylon membranes using a pipetting robot in order to produce a high number of identical filters. The filters could then be used for DNA/RNA profiling studies by hybridization of differentially labeled nucleic acids from the set of fertile and sterile lines.

Transcriptional activity is genome-specific and nucleary regulated

By disrupting mitochondrial preparations in the presence of [α - 32 P]G/UTP and extending previously initiated transcripts, labelled run-on transcripts were produced. These transcripts were used as probes on the filters and by quantifying the hybridisation signal from each gene the relative transcriptional activities could be estimated. In all lines the rRNA genes were the most actively transcribed. This finding is in contrast with the investigation performed by Giegé *et al.* (2000) in *A. thaliana*, who found rRNA genes no more transcriptionally active than other mitochondrial genes. However, it is in accordance with the investigations performed in corn (Finnegan and Brown, 1990; Mulligan *et al.*, 1991; Muise and Hauswirth, 1992) in which the rRNA genes were more active than the protein coding mitochondrial genes. An important difference in the methodology used by us and by Giegé *et al.* (2000) is our use of RNase to remove adjacent single stranded RNA before the quantification. As mitochondrial transcripts regularly extend extensively outside the reading frames this procedure reduces the influence of differential transcript lengths, which cannot be normalized for. Additionally, we used flower bud mitochondria, whereas Giegé *et al.* (2000) used cell culture mitochondria. These very different plant materials may account for some of the differences observed.

Interestingly, large differences were found in transcriptional activity for individual genes between the lines, a finding which correlates well with the comparison of *B. napus* and *A. thaliana* mitochondrial genomes made by Handa (2003). Handa's study demonstrated large structural shifts in genome structure between the two lines, although the gene content was basically identical. Consequently, promoters for a large number of the investigated genes seem to have shifted. The CMS and restored line had mainly similar transcriptional activities as expected due to their identical mitochondrial genomes. However, in comparison to the parental species, some interesting alterations in transcriptional activity were observed. Several of the ribosomal protein subunit genes had very reduced transcriptional activities and the *orf139* had highly increased activity. RFLP analysis demonstrated that these genes were exclusively inherited from *A. thaliana* and as a consequence they would be under the control of *A. thaliana*-specific promoters that work differently, and in most cases less efficient, in the *B. napus*.

nuclear background. In addition to promoter strength transcriptional activity can also be regulated by selective DNA amplification in the mitochondrial genome (Muisse & Hauswirth, 1995). We could, however, only detect marginal alterations of gene copy numbers by probing the filters with end-labelled mtDNA.

RNA turnover regulates transcript steady-state levels

Steady-state levels of transcripts from individual genes were also investigated in order to compare with the transcriptional activities. Transcript abundance was estimated by end-labelling RNA isolated from the same mitochondrial preparations used in the run-on assays and hybridising this RNA to identical filters. From these results it was obvious that steady-state levels of RNA were not directly proportional to transcriptional activity i.e. promoter strength. Thus, RNA steady-state levels seem to be the result of transcriptional activity as well as differential degradation rates. This finding is in accordance with the studies performed by Mulligan *et al.* (1991) and Giegé *et al.* (2000) in corn and *A. thaliana* respectively, who also demonstrated the influence of post-transcriptional regulation. The steady-state levels of individual genes shifted less than transcriptional activities between *B. napus* and *A. thaliana*. In contrast, the CMS line had higher transcript abundance than the parental lines for several genes, including *nad4*, *cox3*, *atp1*, *atp9*, *ccmC*, *ccmFC*, *rpl2*, *rpl16*, *Ψsdh4*, *orf294* and *orf240a*. Northern analysis revealed that the elevated steady-state levels often were correlated with presence of transcripts longer than those found in the parental lines. RFLP analysis revealed that the DNA for encoding these genes in the CMS line was inherited from *A. thaliana*. Although we cannot exclude the possibility that the novel transcripts are a consequence of genome rearrangements they are more likely due to alloplasmic incompatibilities of transcriptional regulation. Similar observations were reported in alloplasmic tobacco (Håkansson & Glimelius, 1991), maize (Wen & Chase, 1999) and *Brassica* (Li *et al.*, 1998). Alloplasmic-induced transcripts can result both from novel transcript initiation sites, such as demonstrated with the *cox2* gene of maize (Newton *et al.*, 1995) and the *atp1* of tobacco (Edqvist and Bergman, 2002) and from differences in processing ability, as shown for restorer effects of *T-urf13* in maize (Dill *et al.*, 1997), *nap orf222* and *pol orf224* in *B. napus* (Li *et al.*, 1998; Menassa *et al.*, 1999; Brown *et al.*, 1999) and the CMS-associated loci in sorghum (Tang *et al.*, 1996) and corn CMS-S (Wen and Chase, 1999). Furthermore, Northern analysis revealed more clearly quantitative differences between lines for low abundant transcripts. The *A. thaliana orf139*, *orf240a* and *orf294* were more abundant in the CMSline than in *A. thaliana* and the fertility-restored line. Thus, we focused on control of expression of these putative CMS-associated genes.

Characterization of putative CMS-associated genes (III, IV)

In order to characterize the post-transcriptional and developmental regulation of *orf139*, *orf240a* and *orf294* different analyses were performed. Mapping of the transcript ends was performed with circular RT-PCR. This method, first utilised in plant mitochondria by Kuhn & Binder (2002) utilises the ability of T4 RNA ligase to form junctions of transcript 5' and 3' ends that can be detected by PCR and

sequencing. RT-PCR on cDNA primed with dT-oligos was utilized for the mapping of putative polyadenylation sites (Gagliardi & Leaver, 1999). Additionally we performed RT-PCR and quantitative real-time RT-PCR experiments on RNA isolated from different plant organs as well as *in situ* hybridisation to examine the possible developmental regulation of the transcripts. Finally, to investigate whether expression of the three *orfs* correlated with the male-sterile phenotype we investigated a population of both sterile, semi-sterile and fertile alloplasmic lines. The results of these experiments are summarized in Table 3.

Table 3. Summary of transcript characteristics for *orf139*, *orf240a* and *orf294*. (A) *A. thaliana*, (C) CMS, (R) restored

feature	gene		
	<i>orf139</i>	<i>orf240a</i>	<i>orf294</i>
transcriptional activity	very high (C, R) high (A)	high (A) moderate (C, R)	moderate (A, C, R)
steady-state levels	extremely low (A) very low (R) low (C)	very low (A) low (R) moderate (C)	very low (A, R) moderate (C)
5' ends	identical in C, R unknown in A	identical	similar in C, R shorter in A
3' ends	heterogenous (C, R) unknown in A*	identical	heterogenous, extend into <i>atp1</i>
CNM near 5' ends	yes	no	yes
editing	-	yes (A, C, R)	-
polyadenylation	- *	yes (A)	yes (A, C, R)
mosaic structure	no	yes, part of <i>rps3</i>	no
similarities to identified genes	no	yes, to <i>orf222</i> and <i>orf224</i> in <i>nap</i> and <i>pol B. napus</i> CMS	no
tissue-specific expression levels	yes	-	yes
co-segregation with male-sterility	no	no	possibly

*a polyadenylated transcript end 596 nt upstream of the start codon was found in *A. thaliana*.

From the run-on and kinase labelling experiments it was determined that *orf139* is very actively transcribed but degraded to very low steady state levels. In the CMS and restored line a common type of transcript 5' end mapped 1093 nt upstream of the start codon, close to a consensus promoter sequence. The 3' ends were more heterogenous. We thus propose that the *orf139* transcripts possess relatively stable 5' ends, but are actively degraded from 3' ends. Unfortunately, we were unable to map the transcript ends in *A. thaliana* by the cRT-PCR experiment,

probably due to the low transcript abundance. However, by RT-PCR experiments with a forward primer 956 nt upstream of the startcodon a polyadenylated 3' end at a position 596 nt upstream of the startcodon was found. This processing/polyA site is absent in the CMS and restored line (unpublished results) and thus mediated by species-specific factors. The *orf139* does not have typical CMS features such as fragments of standard genes or co-transcription with standard genes. The finding of a fertile line among the alloplasmic lines, which expresses the *orf* makes it an unlikely candidate to be responsible for the male-sterile phenotype. On the other hand, other facts that pinpoint a function for this gene region are the presence of a CNM promoter, the presence of a small non-messenger RNA (Marker *et al.*, 2002) in the 5'UTR of the longest transcripts and a tissue-specific transcript accumulation, i.e., in carpels of *A. thaliana*, in flower buds of the CMSline and in roots, buds and carpels of the restored line.

Orf240a has several features consistent with its being a true gene, including distinct transcripts, which appear to be constitutively transcribed, editing of the transcripts and a location far from other identified genes. Editing of transcripts is very rare in regions of non-coding mtDNA (Giegé & Brennicke, 1999). The comparison of transcriptional activity and steady-state levels indicated that the gene is less stable in *A. thaliana* than in the other materials. Since the cRT-PCR analysis showed that *orf240a* transcripts are identical in all three lines differential processing could not be the cause of stability differences. A difference in polyadenylation-mediated transcript degradation probably provides the best explanation for the differences in transcript stability between lines. Indeed, we could demonstrate the presence of polyA-tails in *A. thaliana* transcripts. Even though we lack quantitative data of polyadenylation in the investigated material, a hypothesis is that in the CMSline, polyadenylation is less efficient. Alternatively, polyadenylated transcripts are less efficiently degraded. *Orf240a* has a high predicted amino acid similarity with *orf222* and *orf224* in the *B. napus* CMS-inducing cytoplasms *nap* and *pol*. Furthermore *orf240a* contains a small part of the standard gene, *rps3*, a characteristic typical for CMS-associated genes. However, among the alloplasmic lines, one fertile line expressing *orf240a* was found. Although it cannot be excluded that this fertile line have retained nuclear restorer genes from *A. thaliana* acting on a translational level, *orf240a* is therefore less likely CMS-associated.

Run-on experiments and Northern analysis revealed that *orf294* transcripts are both differentially processed and degraded among the lines. In the CMS and restored line transcripts are generally about 400 nt longer than in *A. thaliana*. The cRT-PCR analysis revealed that this difference is due to longer transcript 5' ends. One cRT-PCR clone from *A. thaliana* extends several hundred nt farther upstream close to a CNM promoter. This cRT-PCR clone could represent a *de novo* transcript with the remaining transcript ends resulting from differential processing. Several polyadenylation sites were found in transcript 3' ends. Thus, polyadenylation-mediated degradation is a possible regulatory mechanism. 3' ends were found that extended into *atp1*. The possible co-transcription with *atp1* is a typical CMS-associated characteristic. One interesting speculation is that *orf294-atp1* co-transcripts interfere with production of ATP1 and complex V of

respiratory chain. Reduced ATP production has been reported in this CMS system (Teixeira *et al.*, 2005a) as well as in *Nicotiana* CMS (Bergman *et al.*, 2000) and could interfere with cell proliferation in flower meristems (Farbos *et al.*, 2001, Teixeira *et al.*, 2005b). The developmental patterns suggest a genotype and tissue-specific regulation since accumulation seems to occur preferentially in carpels of *A. thaliana*, roots of the restored line and less in petals of all three lines. Additionally, *in situ* hybridisation showed that *orf294* is expressed in early flower meristems of *A. thaliana* and to some extent in the CMS-line, but not in the restored line. Among the alloplasmic lines transcription of *orf294* was exclusively found in sterile lines. However, it was also absent in one sterile and one semi-sterile line. This does not exclude *orf294* from being CMS-associated, but implying presence of other CMS-associated genes in these sterile lines.

To summarize, *orf139* and *orf240a* seem not to be CMS-associated, although the possibility of retained nuclear restorer genes acting on a translational level in the fertile line expressing *orf139* and *orf240a* transcripts cannot be ignored. Nevertheless, *orf139* and *orf240a* may well have other functions in plant mitochondria. The *orf294* may be causing the CMS-phenotype. However, the exception with the sterile and semi-sterile lines not expressing *orf294*, implies that other CMS-inducing genes are present in these lines. Taken together these results reveal the complexity of alloplasmic induced mitochondrial gene expression aberrations and the difficulty to associate particular transcripts with the CMS-trait. The diverse range of flower phenotypes among the *B. napus* (+) *A. thaliana* alloplasmic lines indicate that several CMS-associated, or flower morphology affecting loci, in the *A. thaliana* mt-genome are present and differently retained in the different lines. The *A. thaliana* mt-genome contains 460 *orfs* longer than 60 codons (Unseld *et al.*, 1997). The expression pattern, or alloplasmic effects on expression of the vast majority of these *orfs* remain unknown. The alloplasmic influenced accumulation of *orf139*, *orf240a* and *orf294* transcripts provide examples of how putative genes, without known function, can be activated when removed from their nuclear suppressors.

Conclusions

The main conclusions from the results presented in this thesis follow.

- Introgression of *A. thaliana* mitochondrial DNA to the nuclear background of *B. napus* results in cytoplasmic male-sterile plants.
- The cytoplasmic male-sterile lines often display homeotic conversions of anthers to carpeloid structures, reduced petal size and some disturbances in vegetative growth.
- Introgression of *A. thaliana* chr III into CMS line 4:19 restores anther development and pollen production.
- *A. thaliana* and *B. napus* have different promoter strength for a large number of individual genes.

- As demonstrated in a CMS-line, the transcriptional activity of *A. thaliana* mt-DNA is often altered in the nuclear background of *B. napus*. In particular, several ribosomal protein subunit genes have reduced transcriptional activities and *orf139* has increased transcriptional activity.
- Copy-number alteration of individual mt-genes is not a major mechanism for gene expression regulation.
- Post-transcriptional events largely counter-balance differences in transcriptional activity.
- Novel mitochondrial transcripts were formed in the CMS line, probably due to processing aberrations.
- Transcripts of several *A. thaliana* mitochondrial *orfs* accumulate in the nuclear background of *B. napus*.
- In *A. thaliana* and the fertility restored line *orf240a* and *orf294* transcripts are more efficiently degraded in comparison to the CMS-line. This degradation could be polyadenylation-mediated as polyA-tails were found in *A. thaliana orf240a* transcripts and in *orf294* transcripts from all three lines.
- Transcripts of *orf139* and *orf294* accumulate differently in mitochondria dependent on genotype and tissue.
- The *orf139* and *orf240a* are less likely candidates to be responsible for the male-sterile phenotype, whereas the *orf294* can be CMS-associated. However, it is likely that more than one locus in the *A. thaliana* mt-DNA could encode CMS in the nuclear background of *B. napus*.

Future perspectives

This thesis describes the establishment and characterization of a *B. napus* (*A. thaliana*) CMS system. The system was utilized to generate information about the genetics of CMS. Additionally, the unique combination of two fully sequenced mitochondrial genomes provides possibilities to study species-specific factors regulating mitochondrial gene expression. I suggest the following avenues of research as examples of how to exploit the material for further investigations.

- As visualised in the CMS-line, a large number of novel transcripts are produced as a result of alloplasmic incompatibilities, e.g. for *atp9*, *cox3* and *ccmC*. Mapping of these transcripts would indicate if insufficient processing of precursor transcripts is a general phenomenon and if this can be related to specific sequence motifs.
- Alloplasmic effects on transcriptional activity were found, most obvious for several of the ribosomal protein genes. Mapping of the *de novo* transcript 5' ends and promoter elements would indicate if these genes are regulated by a special kind of promoter motif that would be driven by species-specific transcription factors.
- Even though neither *orf139*, *orf240a* or *orf294* might be causing CMS their function is still intriguing. Polysome analysis to indicate if the transcripts are translationally active would be an initial step. As a second

step production of antibodies against the putative polypeptides and determination of the cellular localization and developmental regulation of the proteins could be performed.

- The results indicate that more than one locus in the *A. thaliana* mt-genome might result in CMS when transferred to the *B. napus* nuclear background. In paper IV we used 16 sterile and 8 fertile lines. More fertile cybrid lines are available. Mapping of the genomes with more *A. thaliana* specific probes and a larger population of plants could reveal other putative CMS-loci.
- Although the restorer element consists of the whole *A. thaliana* chr III, the full sequence of it is available. Some particularly interesting genes have already been mentioned. Transformation of the CMS-line with these putative restorer genes and corresponding analyses of mitochondrial gene expression and flower development would provide means to identify genes involved in the nuclear-mitochondrial interaction resulting in CMS. To limit the number of putative restorer genes narrowing the amount of *A. thaliana* would be useful. However, translocations of *A. thaliana* DNA to *B. napus* chromosomes in this material are extremely rare, and would probably require X-ray treatment.
- The combination of *A. thaliana* mt-DNA with the *B. napus* nuclear background obviously resulted in CMS. It would be interesting to see if the reversed combination – *B. napus* mtDNA in *A. thaliana* nuclear background also would result in CMS. An alternative strategy would be to create alloplasmic *A. thaliana* CMS using a cytoplasmic donor species more closely related to *A. thaliana*, for instance *A. lyrata*. With the wealth of information and tools for *A. thaliana* available, an *A. thaliana* CMS system would provide extraordinary good opportunities to study especially the nuclear genes involved in the expression of male sterility.

References

- Abad, A.R., Mehrtehs, B.J. & Mackenzie, S.A. 1995. Specific expression in reproductive tissues and fate of a mitochondrial sterility-associated protein in cytoplasmic male-sterile bean. *Plant Cell* 7, 271-285.
- Abdelnoor, R.V., Yule, R., Elo, A., Christensen, A.C., Meyer-Gauen, G. & Mackenzie, S.A. 2003. Substoichiometric shifting in the plant mitochondrial genome is influenced by a gene homologous to *MutS*. *Proceedings of the National Academy of Sciences, USA* 100, 5968-5973.
- Akagi, H., Nakamura, A., Sawada, R., Oka, M. & Fujimura, T. 1995. Genetic diagnosis of cytoplasmic male-sterile cybrid plants of rice. *Theoretical and Applied Genetics* 90, 948-951.
- Akagi, H., Sakamoto, M., Shinjyo, C., Shimada, H., & Fujimura, T. 1994. A unique sequence located downstream from the rice mitochondrial *atp6* may cause male-sterility. *Current Genetics* 25, 52-58.
- The Arabidopsis Genome Initiative. 2000. Analysis of the genome sequence of the flowering plant *Arabidopsis thaliana*. *Nature* 408, 796-815.
- Balk, J. & Leaver, C.J. 2001. The PET1-CMS mitochondrial mutation in sunflower is associated with premature programmed cell death and cytochrome *c* release. *The Plant Cell* 13, 1803-1818.
- Banga, S.S., Deol, J.S., Banga, S.K. 2003. Alloplasmic male-sterile *Brassica juncea* with *Enarthrocarpus lyratus* cytoplasm and the introgression of gene(s) for fertility restoration from cytoplasm donor species. *Theoretical and Applied Genetics* 106, 1390-1395.
- Bellaoui, M., Grelon, M., Pelletier, G. & Budar, F. 1999. The restorer *Rfo* gene acts post-translationally on the stability of the ORF138 Ogura CMS-associated protein in reproductive tissues of rapeseed cybrids. *Plant Molecular Biology* 40, 893-902.
- Bellaoui, M., Pelletier, G. & Budar, F. 1997. The steady-state level of mRNA from the Ogura cytoplasmic male sterility locus in *Brassica* cybrids is determined post-transcriptionally by its 3' region. *The EMBO Journal* 16, 5057-5068.
- Bentolila S., Alfonso, A.A. & Hanson, M.R. 2002. A pentatricopeptide repeat-containing gene restores fertility to cytoplasmic male-sterile plants. *Proceedings of the National Academy of Sciences, USA* 99, 10887-10892.
- Bergman, P., Edqvist, J., Farbos, I. & Glimelius, K. 2000. Male-sterile tobacco displays abnormal mitochondrial *atp1* transcript accumulation and reduced floral ATP/ADP ratio. *Plant Molecular Biology* 42, 531-544.
- Binder, S. & Brennicke, A. 1993. Transcription initiation sites in *Oenothera* mitochondria. *Journal of Biological Chemistry* 268, 7849-7855.
- Binder, S. & Brennicke, A. 2003. Gene expression in plant mitochondria: transcriptional and post-transcriptional control. *Philosophical Transactions of the Royal Society London B* 358, 181-189.
- Binder, S., Hatzack, F. & Brennicke, A. 1995. A novel pea mitochondrial *in vitro* transcription system recognizes homologous and heterologous mRNA and tRNA promoters. *Journal of Biological Chemistry* 270, 22182-22189.
- Binder, S., Marchfelder, A. & Brennicke, A. 1996. Regulation of gene expression in plant mitochondria. *Plant Molecular Biology* 32, 303-314.
- Birchler, J.A., Auger, D.L. & Riddle, N.C. 2003. In search of the molecular basis of heterosis. *The Plant Cell* 15, 2236-2239.
- Boeshore, M.L., Hanson, M.R. & Izhar, S. 1985. A variant mitochondrial-DNA arrangement specific to petunia stable sterile somatic hybrids. *Plant Molecular Biology* 4, 125-132.
- Bonhomme, S., Budar, F., Ferault, M. & Pelletier, G. 1991. A 2.5 kb *NcoI* fragment of Ogura radish mitochondrial-DNA is correlated with cytoplasmic male sterility in *Brassica* cybrids. *Current Genetics* 19, 121-127.
- Bonhomme, S., Budar, F., Lancelin, D., Small, I., Defrance, M.C. & Pelletier, G. 1992. Sequence and transcript analysis of the *NcoI* 2.5 Ogura-specific fragment correlated with

- cytoplasmic male sterility in Brassica cybrids. *Molecular and General Genetics* 235, 340–348.
- Bonnard, G. & Grienenberger, J.M. 1995. A gene proposed to encode a transmembrane domain of an ABC transporter is expressed in wheat mitochondria. *Molecular and General Genetics* 246, 91-99.
- Brandt, P., Sünkel, S., Unseld, M., Brennicke, A. & Knoop, V. 1992. The nad4L gene is encoded between exon c of nad5 and ORF25 in the Arabidopsis mitochondrial genome. *Molecular and General Genetics* 236, 33-38.
- Brown, G.G. 1999. Unique aspects of cytoplasmic male sterility and fertility restoration in *Brassica napus*. *Journal of Heredity* 90, 351–356.
- Brown, G.G., Auchincloss, A.H., Covello, P.S. Gray, M.W., Menassa, R. & Singh, M. 1991. Characterization of transcription initiation sites on the soybean mitochondrial genome allows identification of a transcription-associated sequence motif. *Molecular and General Genetics* 228, 345-355.
- Brown, G.G., Formanova, N., Jin, H., Wargachuk, R., Dendy, C., Patil, P., Laforest, M., Zhang, J., Cheung, W.Y., Landry, B.S. 2003. The radish *Rfo* restorer gene of Ogura cytoplasmic male sterility encodes a protein with multiple pentatricopeptide repeats. *The Plant Journal* 35, 262-272.
- Budar, F. & Pelletier, G. 2001. Male sterility in plants: occurrence, determinism, significance and use. *Comptes Rendus de l'Académie des Sciences - Series III - Sciences de la Vie* 324, 543-550
- Budar, F., Touzet, P. & De Paepe, R. 2003. The nucleo-mitochondrial conflict in cytoplasmic male sterilities revisited. *Genetica* 117, 3-16.
- Burns J.A., Gerstel, D.U. & Sand, S.A. 1978. Cytoplasmic male sterility in Nicotiana, restoration of fertility, and the nucleolus. II. *N. debneyi* cytoplasm. *Genetics* 90, 151-159.
- Carlsson, J., Teixeira, R., Sundström, J., Lagercrantz, U. & Glimelius, K.. Cytoplasmic male sterile *Brassica napus* displays altered expression in a diverse range of nuclear genes. (manuscript in preparation)
- Charlesworth, D. 2000. Unlocking the secrets of self-incompatibility. *Current Biology* 10, R184-R186.
- Chaumont, F., Bernier, B., Buxant, R., Williams, M.E., Levings III, C.S. and Boutry, M. 1995. Targeting the maize T-urf13 product into tobacco mitochondria confers methomyl sensitivity to mitochondrial respiration. *Proceedings of the National Academy of Sciences, USA* 92, 1167-1171.
- Chiang, M.S., Chong, C., Landry, B.S. & Crête, R. 1993. Cabbage. In: Kalloo, G. & Bergh, B.O. (eds.) *Genetic improvement of vegetable crops*. Pergamon Press, Oxford.
- Clark, E., Schnabelrauch, L., Hanson, M.R. & Sink, K.C. 1986. Differential fate of plastid and mitochondrial genomes in *Petunia* somatic hybrids. *Theoretical and Applied Genetics* 72, 748-755
- Clifton, S. W., Minx, P., Fauron, C. M.-R., Gibson, M. Allen, J. O., Sun, H., Thompson, M., Barbazuk, W. B., Kanuganti, S., Tayloe, C., Meyer, L., Wilon, R. K. & Newton, K. J. 2004. Sequence and comparative analysis of the maize NB mitochondrial genome. *Plant Physiology* 136, 3486-3503.
- Coen, E.S. & Meyerowitz, E.M. 1991. The war of the whorls: genetic interactions controlling flower development. *Nature* 353, 31-37.
- Correns, C. 1904. Experimentelle untersuchungen über die Gynodioecie. *Berichte der Deutschen Botaischen Gesellschaft* 22, 506-517.
- Correns, C. 1906. Die Vererbung der geschlechtsformen bei den gynodiöcischen Pflanzen. *Berichte der Deutschen Botaischen Gesellschaft* 24, 459-474.
- Covello, P.S. & Gray, M.W. 1989. RNA editing in plant mitochondria. *Nature* 341, 662-666.
- Covello, P.S. & Gray, M.W. 1991. Sequence analysis of wheat mitochondrial transcripts capped *in vitro*: definitive identification of transcription initiation sites. *Current Genetics* 20, 245-251.
- Crisp, P. & Tapsell, C.R. 1993. Cauliflower. In: Kalloo, G. & Bergh, B.O. (eds.) *Genetic improvement of vegetable crops*. Pergamon Press, Oxford.

- Cui, X., Wise, R.P. & Schnable, P.S. 1996. The *rf2* nuclear restorer gene of male-sterile T-cytoplasm maize. *Science* 272, 1334-1336.
- Darwin, C. 1877. *The different forms of flowers on plants of the same species*. John Murray, London.
- Desloire, S.H., Gherbil, W., Laloui, S., Marhadour, V., Clouet, V., Cattolico, L., Falentin, C., Giancola, S., Renard, M., Budar, F., Small, I., Caboche, M., Delourme, R., Bendahmane, A. 2003. Identification of the fertility restoration locus, *Rfo*, in radish, as a member of the pentatricopeptide-repeat protein family. *EMBO Reports* 4, 588-594.
- Dewey, R.E., Levings III, C.S. & Timothy, D.H. 1986. Novel recombinations in the maize mitochondrial genome produce a unique transcriptional unit in the Texas male-sterile cytoplasm. *Cell* 44, 439-449.
- Dewey, R.E., Timothy, D.H. & Levings, C.S. 1987. A mitochondrial protein associated with cytoplasmic male sterility in the T-cytoplasm of maize. *Proceedings of the National Academy of Sciences, USA* 84, 5374-5378.
- Dietrich, A., Small, I., Cosset, A., Weil, J.H. & Maréchal-Drouard, L. 1996. Editing and import: strategies for providing plant mitochondria with a complete set of functional transfer RNAs. *Biochimie* 78, 518-529.
- Dill, C.L., Wise, R.P. & Schnable, P.S. 1997. *Rf8* and *Rf** mediate unique T-*urf13*-transcript accumulation, revealing a conserved motif associated with RNA processing and restoration of pollen fertility in T-cytoplasm maize. *Genetics* 147, 1367-1379.
- Dombrowski, S., Brennicke, A. & Binder, S. 1997. 3'-Inverted repeats in plant mitochondrial mRNAs are processing signals rather than transcription terminators. *EMBO Journal* 16, 5069-5076.
- Dombrowski, S., Hoffmann, M., Guha, C. & Binder, S. 1999. Continuous primary sequence requirements in the 18-nucleotide promoter of dicot plant mitochondria. *Journal of Biological Chemistry* 274, 10094-10099.
- Dombrowski, S., Hoffmann, M., Kuhn, J., Brennicke, A. & Binder, S. 1998. On the mitochondrial promoters in *Arabidopsis thaliana* and other flowering plants. In: Möller, I.M., Gardeström, P., Glimelius, K. & Glaser, E. (eds.) *Plant mitochondria: From gene to function*. Backhuys Publishers, Leiden, Netherlands. pp 165-170.
- Edqvist, J. & Bergman, P. 2002. Nuclear identity specifies transcriptional initiation in plant mitochondria. *Plant Molecular Biology* 49, 59-68.
- Elo, A., Lyznik, A., Gonzalez, D.O., Kachman, S.D. & Mackenzie, S.A. 2003. Nuclear genes that encode mitochondrial proteins for DNA and RNA metabolism are clustered in the *Arabidopsis* genome. *Plant Cell* 15, 1619-1631.
- Fan, Z. & Stefansson, B.R. 1986. Influence of temperature on sterility of two cytoplasmic male-sterility systems in rape (*Brassica napus* L.). *Canadian Journal of Plant Science* 66, 229-234.
- Farbos, I., Mouras, A., Bereterbide, A. & Glimelius, K. 2001. Defective cell proliferation in the floral meristem of alloplasmic plants of *Nicotiana tabacum* leads to abnormal floral organ development and male sterility. *The Plant Journal* 26, 131-142.
- Farré, J.C., Leon, G., Jordana, X. & Arraya, A. 2001. *cis* Recognition elements in plant mitochondrion RNA editing. *Molecular and Cellular Biology* 21, 6731-6737.
- Finnegan P.M. & Brown, G.G. 1990. Transcriptional and post-transcriptional regulation of RNA levels in maize mitochondria. *The Plant Cell* 2, 71-83.
- Forsberg, J., Dixelius, C., Lagercrantz, U. & Glimelius, K. 1998. UV dose-dependent DNA elimination in asymmetric hybrids between *Brassica napus* and *Arabidopsis thaliana*. *Plant Science* 131, 65-76.
- Gagliardi, D. & Leaver, C.J. 1999. Polyadenylation accelerates the degradation of the mitochondrial mRNA associated with cytoplasmic male sterility in sunflower. *EMBO Journal* 18, 3757-3766.
- Gagliardi, D., Perrin, R., Maréchal-Drouard, L., Grienberger, J.-M. & Leaver, C.J. 2001. Plant mitochondrial polyadenylated mRNAs are degraded by a 3' to 5'-exoribonuclease activity, which proceeds unimpeded by stable secondary structures. *Journal of Biological Chemistry* 276, 43541-43547.

- Geddy, R., Mahé, L. & Brown, G.G. 2005. Cell-specific regulation of a *Brassica napus* CMS-associated gene by a nuclear restorer with related effects on a floral homeotic gene promoter. *The Plant Journal* 41, 333-345.
- Gerstel, D.U., Burns, J.A., Burk, L.G. 1978. Cytoplasmic male sterility in *Nicotiana*, restoration of fertility, and the nucleolus. *Genetics* 89, 157-169.
- Giegé, P. & Brennicke, A. 1999. RNA editing in *Arabidopsis* mitochondria effects 441 C to U changes in ORFs. *Proceedings of the National Academy of Sciences, USA*, 96, 15324-15329.
- Giegé, P., Hoffmann, M., Binder, S. & Brennicke, A. 2000. RNA degradation buffers asymmetries of transcription in *Arabidopsis* mitochondria. *EMBO Reports* 1, 164-170.
- Giegé, P. & Brennicke, A. 2001. From gene to protein in higher plant mitochondria. *Comptes Rendus de l'Académie des Sciences - Series III - Sciences de la Vie* 324, 209-217.
- Gray, A. R. 1993. Broccoli. In: Kalloo, G. & Bergh, B.O. (eds.) *Genetic improvement of vegetable crops*. Pergamon Press, Oxford.
- Grelon, M., Budar, F., Bonhomme, S. & Pelletier, G. 1994. Ogura cytoplasmic male sterility (CMS)-associated *orf138* is translated into a mitochondrial membrane polypeptide in male-sterile *Brassica* cybrids. *Molecular and General Genetics* 243, 540-547.
- Gualberto, J.M., Weil, J.H. & Grienberger, J.M. 1989. RNA editing in wheat mitochondria results in the conservation of protein sequences. *Nature* 341, 660-662.
- Handa, H. 2003. The complete nucleotide sequence and RNA editing content of the mitochondrial genome of rapeseed (*Brassica napus* L.): comparative analysis of the mitochondrial genomes of rapeseed and *Arabidopsis thaliana*. *Nucleic Acids Research* 28, 2571-2576.
- Hanson, M. R. & Bentolila, S. 2004 Interactions of mitochondrial and nuclear genes that affect male gametophyte development *Plant Cell*, 16, 154-169.
- He, S., Abad, A.R., Gelvin, S.B. & Mackenzie, S.A. 1996. A cytoplasmic male sterility-associated mitochondrial protein causes pollen disruption in transgenic tobacco. *Proceedings of the National Academy of Sciences, USA* 93, 11763-11768.
- Heazlewood, J.L., Whealan, J. & Millar, A.H. 2003. The products of the mitochondrial *orf25* and *orfB* genes are F₀ components in the plant F₁F₀ ATPsynthase. *FEBS Letters* 540, 201-205.
- Hedke, B., Borner, T. & Weihe, A. 1997. Mitochondrial and chloroplast phage-type RNA polymerases in *Arabidopsis*. *Science* 277, 809-811.
- Hedke, B., Borner, T. & Weihe, A. 2000. One RNA polymerase serving two genomes. *EMBO Reports* 1, 435-440.
- Hiesel, R., Wissinger, B., Schuster, W. & Brennicke, A. 1989. RNA editing in plant mitochondria. *Science* 246, 1632-1643.
- Hoffmann, M. & Binder, S. 2002. Functional importance of nucleotide identities within the pea *atp9* mitochondrial promoter sequence. *Journal of Molecular Biology* 320, 943-950.
- Hoffmann, M., Kuhn, J., Däschner, K. & Binder, S. 2001. The RNA world of plant mitochondria. *Progress in Nucleic Acid Research and Molecular Biology* 70, 119-154.
- Horn, R., Kohler, R.H. & Zetsche, K. 1991. A mitochondrial 16 kDa protein is associated with cytoplasmic male sterility in sunflower. *Plant Molecular Biology* 17, 29-36.
- Horn, R., Hustedt, J.E.G., Horstmeyer, A., Hahnen, J., Zetsche, K. & Friedt, W. 1996. The CMS-associated 16 kDa protein encoded by *orfH522* in the PET1 cytoplasm is also present in other male-sterile cytoplasms of sunflower. *Plant Molecular Biology* 30, 523-528.
- Horner, H.T. & Palmer, R.G. 1995. Mechanisms of genic male sterility. *Crop Science* 35, 1527-1535.
- Howad, W. & Kempken, F. 1997. Cell tyoe-specific loss of *atp6* RNA editing in cytoplasmic male sterile *Sorghum bicolor*. *Proceedings of the National Academy of Sciences, USA* 94, 11090-11095.
- Håkansson, G & Glimelius, K. 1991. Extensive nuclear influence on mitochondrial transcription and genome structure in male-fertile and male-sterile alloplasmic *Nicotiana* materials. *Molecular and General Genetics* 229, 380-388.

- Itani, K. & Handa, H. 1998. Rapeseed mitochondrial *ccb206*, a gene involved in cytochrome *c* biogenesis, is co-transcribed with the *nad3* and *rps12* genes: organization, transcription, and RNA editing of the *nad3/rps12/ccb206* locus. *Current Genetics* 34, 318-325.
- Iwabuchi, M., Kyoizuka, J. & Shimamoto, K. 1993. Processing followed by complete editing of an altered mitochondrial *atp6* RNA restores fertility of cytoplasmic male sterile rice. *The EMBO Journal* 12, 1437-1446.
- Iwabuchi, M., Koizuka, N., Fujimoto, H., Sakai, T. & Imamura, J. 1999. Identification and expression of the kosenia radish (*Raphanus sativus* cv. Kosenia) homologue of the Ogura radish CMS-associated gene, *orf138*. *Plant Molecular Biology* 39, 183-188.
- Janska, H., Sarria, R., Woloszynska, M., Arrieta-Montiel, M. & Mackenzie, S. 1998. Stoichiometric shifts in the common bean mitochondrial genome leading to male sterility and spontaneous reversion to fertility. *The Plant Cell* 10, 1163-1180.
- Jones, H.A. & Clarke, A.E. 1943. Inheritance of male sterility in the onion and the production of hybrid seed. *Proceedings of the American Society for Horticultural Sciences* 43, 189-194.
- Kaloo, G. 1993. Tomato. In: Kaloo, G. & Bergh, B.O. (eds.) *Genetic improvement of vegetable crops*. Pergamon Press, Oxford, pp. 645-666
- Kaul, M.L.H. 1988. Male sterility in higher plants. *Monographs on Theoretical and Applied Genetics Vol 10*. Springer Verlag, Berlin, pp. 1005.
- Kazama, T. & Toriyama, K. 2003. A pentatricopeptide repeat-containing gene that promotes the processing of aberrant *atp6* RNA of cytoplasmic male-sterile rice. *FEBS Letters* 544, 99-102.
- Klein, M., Eckert-Ossenkopp, U., Schmiedeberg, I., Brandt, P., Unsel, M., Brennicke, A. & Schuster, W. 1994. Physical mapping of the mitochondrial genome of *Arabidopsis thaliana* by cosmid and YAC clones. *The Plant Journal* 6, 447-455.
- Kofer, W., Glimelius, K., Bonett, H.T. 1991. Modifications of mitochondrial DNA cause changes in floral development in homeotic-like mutants of tobacco. *The Plant Cell* 3, 759-769
- Koizuka, N.R., Imai, R., Fujimoto, H., Hayakawa, T., Kimura, Y., Kohno-Muraase, J., Sakai, T., Kawasaki, S. & Imamura, J. 2000. Genetic characterization of a pentatricopeptide repeat protein gene, *orf687*, that restores fertility in the cytoplasmic male-sterile Kosenia radish. *The Plant Journal* 34, 407-415.
- Koizuka, N.R., Imai, R., Iwabuchi, M., Sakai, T. & Imamura, J. 2003. Genetic analysis of fertility restoration and accumulation of ORF125 mitochondrial protein in the kosenia radish (*Raphanus sativus* cv. Kosenia) and a *Brassica napus* restorer line. *Theoretical and Applied Genetics* 100, 949-955.
- Komori, T., Ohta, S., Murai, N., Takakura, Y., Kuraya, Y., Suzuki, S., Hiei, Y., Imaseki, H. & Nitta, N. 2004. Map-based cloning of a fertility restorer gene, *Rf-1*, in rice (*Oryza sativa* L.) *The Plant Journal* 37, 315-325.
- Krishna Rao, M., Uma Devi, K. & Arundhati, A. 1990. Applications of genic male sterility in plant breeding. *Plant Breeding* 105, 1-25.
- Kubo, T., Nishizawa, S., Sugawara, A., Itchoda, N., Estiati, A. and Mikami, T. 2000. The complete nucleotide sequence of the mitochondrial genome of sugar beet (*Beta vulgaris* L.) reveals a novel gene for tRNA(Cys)(GCA). *Nucleic Acids Research* 28, 2571-2576.
- Kuhn, J. & Binder, S. 2002. RT-PCR analysis of 5' to 3'-end-ligated mRNAs identifies the extremities of *cox2* transcripts in pea mitochondria. *Nucleic Acids Research* 30, 439-446.
- Kuhn, J., Tengler, U. & Binder, S. 2001. Transcript lifetime is balanced between stabilizing stem-loop structures and degradation-promoting polyadenylation in plant mitochondria. *Molecular and Cellular Biology* 21, 731-742.
- Kühn, K., Weihe, A. and Börner, T. 2005. Multiple promoters are a common feature of mitochondrial genes in Arabidopsis. *Nucleic Acids Research* 33, 337-346.
- Köhler, R.H., Horn, R., Lössl, A. & Zetsche, K. 1991. Cytoplasmic male sterility in sunflower is correlated with the co-transcription of a new open reading frame with the *atpA* gene. *Molecular and General Genetics* 227, 369-376.

- Kölreuter, D. J. D. 1763. Vorläufige Nachricht von einigen das Geschlecht der Pflanzen betreffenden Versuchen und Beobachtungen. Fortsetzung 1. *Ostwalds Klassiker der exakten Wissenschaften Nr. 41*. Engelmann, Leipzig.
- Landgren, M. & Glimelius, K. 1990. Analysis of chloroplast and mitochondrial segregation in three different combinations of somatic hybrids produced within *Brassicaceae*. *Theoretical and Applied Genetics* 80, 776-784.
- Landgren, M. & Glimelius, K. 1994. A high frequency of intergenomic mitochondrial recombination and an overall biased segregation of *B. campestris* or recombined *B. campestris* mitochondria were found in somatic hybrids within *Brassicaceae*. *Theoretical and Applied Genetics* 87, 854-862.
- Landgren, M., Sundberg, E. & Glimelius, K. 1994. Biased mitochondrial segregation, independent of cell type used for fusion and of hybrid nuclear content, was found in *Brassica napus* (+) *Brassica oleracea* somatic hybrids. *Plant Science* 103, 51-57.
- Landgren, M., Zetterstrand, M., Sundberg, E., and Glimelius, K. 1996. Alloplasmic male-sterile *Brassica* lines containing *B. tournefortii* mitochondria express an ORF 3' of the *atp6* gene and a 32 kDa protein. *Plant Molecular Biology* 32, 879-890.
- Laver, H.K., Reynolds, S.J., Moneger, F., & Leaver, C.J. 1991. Mitochondrial genome organization and expression associated with cytoplasmic male sterility in sunflower (*Helianthus annuus*). *The Plant Journal* 1, 185-193.
- Levings III, C.S. 1993. Thoughts on cytoplasmic male sterility in cms-T maize. *The Plant Cell* 5, 1285-1290.
- L'Homme, Y. & Brown, G.G. 1993. Organizational differences between cytoplasmic male sterile and male fertile *Brassica* mitochondrial genomes are confined to a single transposed locus. *Nucleic Acids Research* 21, 1903-1909.
- Li, X.-Q., Jean, M., Landry, B.S. & Brown, G.G. 1998. Restorer genes for different forms of *Brassica* cytoplasmic male sterility map to a single nuclear locus that modifies transcripts of several mitochondrial genes. *Proceedings of the National Academy of Sciences, USA* 95, 10032-10037.
- Linke, B., Nothnagel, T. & Borner, T. 1999. Morphological characterization of modified flower morphology of three novel alloplasmic male sterile carrot sources. *Plant Breeding* 118, 543-548.
- Linke, B., Nothnagel, T. and Borner, T. 2003. Flower development in carrot CMS plants: Mitochondria affect the expression of MADS box genes homologous to GLOBOSA and DEFICIENS. *The Plant Journal* 34, 27-37.
- Liu, F., Cui, X., Horner, H.T., Weiner, H. & Schnable, P.S. 2001. Mitochondrial aldehyde dehydrogenase activity is required for male fertility in maize. *The Plant Cell* 13, 1063-1078.
- Liu, J.-H., Landgren, M. & Glimelius, K. 1996. Transfer of the *Brassica tournefortii* cytoplasm to *B. napus* for the production of cytoplasmic male sterile *B. napus*. *Physiologia Plantarum* 96, 123-129.
- Livers, R.W. 1964. Fertility restoration and its inheritance in cytoplasmic male-sterile wheat. *Science* 144, 420.
- Lohmann, J.U. & Weigel, D. 2002. Building Beauty: The genetic control of floral patterning. *Developmental Cell* 2, 135-142.
- Lupold, D.S., Caoile, A.G.F.S. & Stern, D.B. 1999. Polyadenylation occurs at multiple sites in maize mitochondrial *cox2* mRNA and is independent of editing status. *The Plant Cell* 11, 1565-1577.
- Lurin, C., Andrés, C., Aubourg, S., Bellaoui, M., Bitton, F., Bruyère, C., Caboche, M., Debast, C., Gualberto, J., Hoffmann, B., Lecharny, A., Le Ret, M., Martin-Magniette, M.-L., Mireau, H., Peeters, N., Renou, J.-P., Szurek, B., Taconnat, L. & Small, I. 2004. Genome-wide analysis of Aabidopsis pentatricopeptide repeat proteins reveals their essential role in organelle biogenesis. *The Plant Cell* 16, 2089-2103.
- Mackenzie, S.A. & Chase, C.D. 1990. Fertility restoration is associated with loss of a portion of the mitochondrial genome in cytoplasmic male-sterile common bean. *The Plant Cell* 2, 905-912.

- Malek, O., Brennicke, A. & Knoop, V. 1997. Evolution of trans-splicing plant mitochondrial introns in pre-Permian times. *Proceedings of the National Academy of Sciences, USA* 94, 553-558.
- Marchfelder, A., Brennicke, A. & Binder, S. 1996. RNA editing is required for efficient excision of tRNAPhe from precursors in plant mitochondria. *Journal of Biological Chemistry* 271, 1898-1903.
- Mariani, C., De Bucheleer, M., Truttner, S., Leemans, J. & Goldberg, R. 1990. Induction of male-sterility in plants by a chimaeric ribonuclease gene. *Nature* 347, 737-741.
- Mariani, C., Gossele, V., De Bucheleer, M., De Block, M., Goldberg, R., De Greef, W. & Leemans, J. 1990. A chimaeric ribonuclease inhibitor gene restores fertility to male sterile plants. *Nature* 357, 384-387.
- Marienfeld, J.R., Unseld, M., Brandt, P. & Brennicke, A. 1997. Mosaic open reading frames in the *Arabidopsis thaliana* mitochondrial genome. *Biological Chemistry* 378, 859-862.
- Marker, C., Zemann, A., Terhörst, T., Kiefmann, M., Kastenmayer, J. P., Green, P., Bachelierie, J-P., Brosius, J. & Hüttenhofer, A. 2002. Experimental Rnomics: Identification of 140 candidates for small non-messenger RNAs in the plant *Arabidopsis thaliana*. *Current Biology* 12, 2002-2013.
- Maunder, A.B. 1999. Logistics of seed production and commercialisation. In: Coors, J.G. and Pandey, S. (eds.) *Genetics and exploitation of heterosis in crops*. American society of agronomy and crop science society of America, Madison, WI, USA.
- Menassa, R., L'Homme, Y. & Brown, G.G. 1999. Post-transcriptional and developmental regulation of a CMS-associated mitochondrial gene region by a nuclear restorer gene. *The Plant Journal* 17, 491-499.
- Monéger, F., Smart, C.J. & Leaver, C.J. 1994. Nuclear restoration of cytoplasmic male sterility in sunflower is associated with the tissue-specific regulation of a novel mitochondrial gene. *EMBO Journal* 13, 8-17.
- Moore, R.H. 1950. Several effects of maleic hydrazide on plants. *Science* 112, 152-153.
- Muise, R.C. & Hauswirth, W.W. 1992. Transcription in maize mitochondria: effects of tissue and mitochondrial genotype. *Current Genetics* 22, 235-242.
- Muise, R.C. & Hauswirth, W.W. 1995. Selective DNA amplification regulates transcript levels in plant mitochondria. *Current Genetics* 28, 113-121.
- Mulligan, R.M., Leon, P. & Walbot, V. 1991. Transcriptional and posttranscriptional regulation of maize mitochondrial gene expression. *Molecular and Cellular Biology* 11, 533-543.
- Murai, K., Takumi, S., Koga, H. & Ogihara, Y. 2002. Pistillody, homeotic transformation of stamens into pistil-like structures, caused by nuclear-cytoplasm interaction in wheat. *The Plant Journal* 29, 169-181.
- Newton, K.J., Winberg, B., Yamato, K., Lupold, S. & Stern, D.B. 1995. Evidence for a novel mitochondrial promoter preceding the *cox2* gene of perennial teosintes. *EMBO Journal* 14, 585-593.
- Nivison, H.T. & Hanson, M.R. 1989. Identification of a mitochondrial protein associated with cytoplasmic male sterility in *Petunia*. *The Plant Cell* 1, 1121-1130
- Notsu, Y., Masood, S., Nishikawa, T., Kubo, N., Akiduki, G., Nakazono, M., Hirai, A. & Kadowaki, K. 2002. The complete sequence of the rice (*Oryza sativa* L.) mitochondrial genome: frequent DNA sequence acquisition and loss during the evolution of flowering plants. *Molecular Genetics and Genomics*, 268, 434-445.
- Ockendon, D.J. & Smith, B.M. 1993. Broccoli. In: Kalloo, G. & Bergh, B.O. (eds.) *Genetic improvement of vegetable crops*. Pergamon Press, Oxford.
- Oda, K., Yamato, K., Ohta, E., Nakamura, Y., Takemura, M., Nozato, N., Akashi, K., Kanegae, T., Ogura, Y., Kohchi, T. & Ohyama, K. 1992. gene organization deduced from the complete sequence of liverwort *Marchantia polymorpha* mitochondrial DNA. *Journal of Molecular Biology* 223, 1-7.
- Ohtani, K., Yamamoto, H. & Akimitsu, K. 2002. Sensivity to *Alternaria alternata* toxin in citrus because of altered mitochondrial RNA processing. *Proceedings of the National Academy of Sciences, USA* 99, 2439-2444.

- Palmer, J.D. & Shields, C.R. 1984. Tripartite structure of the *Brassica campestris* mitochondrial genome. *Nature* 307, 437-440.
- Pathania, A., Bhat, S.R., Dinesh Kumar, V., Ashutosh, Kirti, P.B., Prakash, S. & Chopra, V.L. 2003. Cytoplasmic male sterility in alloplasmic *Brassica juncea* carrying *Diplotaxis catholica* cytoplasm: molecular characterization and genetics of fertility restoration. *Theoretical and Applied Genetics* 107, 455-461
- Perrin, R., Meyer, E.H., Zaepfel, M., Kim, Y.-J., Mache, R., Grienenberger, J.-M., Gualberto, J.M. & Gagliardi, D. 2004a. Two exoribonucleases act sequentially to process mature 3'-ends of *atp9* mRNAs in *Arabidopsis* mitochondria. *Journal of Biological Chemistry* 279, 25440-25446.
- Perrin, R., Lange, H., Grienenberger, J.-M. & Gagliardi, D. 2004b. AtmtPNPase is required for multiple aspects of the 18S rRNA metabolism in *Arabidopsis thaliana* mitochondria. *Nucleic Acids Research* 32, 5174-5182.
- Polowick, P.L. & Sawhney, V.K. 1987. A scanning electron microscopic study on the influence of temperature on the expression of cytoplasmic male sterility in *Brassica napus*. *Canadian Journal of Botany* 65, 807-814
- Prakash, S., Ahuja, I., Upreti, C., Kumar, V.D., Bhat, S.R., Kirti, P.B. & Chopra, V.L. 2001. Expression of male sterility in alloplasmic *Brassica juncea* with *Erucastrum canariense* cytoplasm and the development of a fertility restoration system. *Plant Breeding* 120, 479-482.
- Pruitt, K.D. & Hanson, M.R. 1991. Transcription of the *Petunia* mitochondrial CMS-associated *Pcf* locus in male sterile and fertility-restored lines. *Molecular and General Genetics* 227, 348-355.
- Rathburn, H.B. & Hedgcoth, C. 1991. A chimeric open reading frame in the 5' flanking region of *cox1* mitochondrial DNA from cytoplasmic male-sterile wheat. *Plant Molecular Biology* 16, 909-912.
- Sabar, M., Gagliardi, D., Balk, J. & Leaver, C.J. 2003. ORFB is a subunit of F₁F₀-ATP synthase: Insight into the basis of cytoplasmic male sterility in sunflower. *EMBO Reports* 4, 1-6.
- Sarria, R., Janska, H., Arrieta-Montiel, M., Lyznik, A. & Mackenzie, S.A. 1999. Two nuclear-directed means of suppressing a dominant mitochondrial mutation in common bean. *The Journal of Heredity* 90, 357-361.
- Sarria, R., Lyznik, A., Vallejos, C.E. & Mackenzie, S.A. 1998. A cytoplasmic male-sterility-associated mitochondrial peptide in common bean is post-translationally regulated. *The Plant Cell* 10, 1217-1228.
- Schnable, P.S. 2002. Is *Rf2* a restorer gene of CMS-T in maize? *Trends in Plant Science* 7, 434.
- Schnable, P.S. & Wise, R.P. 1998. The molecular basis of cytoplasmic male sterility and fertility restoration. *Trends in Plant Science* 3, 175-180.
- Schuster, W. & Brennicke, A. 1989. Conserved elements at putative processing sites in plant mitochondria. *Current Genetics* 15, 187-192.
- Shull, G.H. 1908. The composition of a field of maize. *Annual Report of American Breeder's Association* 4, 296-301.
- Singh, M. & Brown, G.G. 1991. Suppression of cytoplasmic male sterility by nuclear genes alters expression of a novel mitochondrial gene region. *The Plant Cell* 3, 1349-1362.
- Small, I., Peeters, N., Legeai, F. & Lurin, C. 2004. Predotar: A tool for rapidly screening proteomes for N-terminal targeting sequences. *Proteomics* 4, 1581-1590.
- Song, J. & Hedgcoth, C. 1994. A chimeric gene (*orf256*) is expressed as protein only in cytoplasmic male-sterile lines of wheat. *Plant Molecular Biology* 26, 535-539.
- Stone, S.L. & Goring, D.R. 2001. The molecular biology of self-incompatibility systems in flowering plants. *Plant Cell, Tissue and Organ Culture* 67, 93-114.
- Tang, H.V., Pring, D.R., Shaw, L.C., Salazar, R.A., Muza, F.R., Yan, B. & Schertz, K.F. 1996. Transcript processing internal to a mitochondrial open reading frame is correlated with fertility restoration in male-sterile sorghum. *The Plant Journal* 10, 123-133.
- Teixeira, R.T., Knorrpp, C. & Glimelius, K. 2005a. Modified sucrose, starch and ATP levels in two alloplasmic male-sterile lines of *Brassica napus*. *Journal of Experimental Botany* 56, 1245-1253.

- Teixeira, R.T., Farbos, I. & Glimelius, K. 2005b. Expression levels of meristem identity and homeotic genes are modified by nuclear-mitochondrial interactions in alloplasmic male-sterile lines of *Brassica napus*. *The Plant Journal* (in press).
- Theissen, G. 2001. Development of floral organ identity: stories from the MADS house. *Current Opinion in Plant Biology* 4, 75-85.
- Theissen, G. & Saedler, H. 2001. Floral quartets. *Nature* 409, 469-471.
- Touzet, P. 2002. Is *Rf2* a restorer gene of CMS-T in maize? *Trends in Plant Science* 7, 434.
- Tsarouhas, V. 2002. Genome mapping of quantitative trait loci in *Salix* with an emphasis on freezing resistance. Doctoral thesis. Swedish University of Agricultural Sciences. *Agraria* 327.
- Tu, Z.P. & Banga, S.K. 1998. Chemical hybridizing agents. In: Banga, S.S. & Banga, S.K. (eds.) *Hybrid Cultivar Development*. Narosa Publishing House, New Dehli, India.
- Unsold, M., Marienfeld, J.R., Brandt, P. & Brennicke, A. 1997. The mitochondrial genome of *Arabidopsis thaliana* contains 57 genes in 366,924 nucleotides. *Nature Genetics* 15, 57-61.
- Ward, B.L., Anderson, R.S. & Bendich, A.J. 1981. The mitochondrial genome is large and variable in a family of plants (Cucurbitaceae). *Cell* 25, 793-803.
- Wen, L. & Chase, C.D. 1999. Pleiotropic effects of a nuclear restorer-of-fertility locus on mitochondrial transcripts in male-fertile and S male-sterile maize. *Current Genetics* 35, 521-526.
- Williams, M.E. 1995. Genetic engineering for pollination control. *Trends in Biotechnology* 13, 344-349.
- Wintz, H., Chen, H.C., Sutton, C.A., Conley, C.A., Cobb, A., Ruth, D. & Hanson, M.R. 1995. Expression of the CMS-associated urfS sequence in transgenic petunia and tobacco. *Plant Molecular Biology* 28, 83-92.
- Wise, R.P., Fliss, A.E., Pring, D.R. & Gegenbach, B.G. 1987. urf13-T of T cytoplasm maize mitochondria encodes a 13 kD polypeptide. *Plant Molecular Biology* 9, 121-126.
- Wise, R.P., Gobelman-Werner, K., Pei, D., Dill, C.L. & Schnable, P.S. 1999. Mitochondrial transcript processing and restoration of male fertility in T-cytoplasm maize. *The Journal of Heredity* 90, 380-385.
- Wretblad, S. 2002. Defence responses in *Brassica nigra* and *B. napus* to the fungal pathogen *Leptosphaeria maculans*. Doctoral thesis. Swedish University of Agricultural Sciences. *Agraria* 316.
- Wright, H. 1980. Commercial hybrid seed production. In Fehr, W.R. & Hadley, H.H. (eds.) *Hybridization of crop plants*. American society of agronomy and crop science society of America, Madison, WI, USA.
- Young, E.G., & Hanson, M.R. 1987. A fused mitochondrial gene associated with cytoplasmic male sterility is developmentally regulated. *Cell* 50, 41-49.
- Yuan, S-C., Zhang, Z-G., He, H-H., Zen, H-L., Lu, K-Y., Lian, J-H. & Wang, B-X. 1993. Two photoperiodic-reactions in photoperiod-sensitive genic male-sterile rice. *Crop Science* 33, 651-660.
- Zabala, G., Gabay-Laughnan, S., & Laughnan, J.R. 1997. The nuclear gene *Rf3* affects the expression of the mitochondrial chimeric sequence R implicated in S-type male sterility in maize. *Genetics* 147, 847-860.

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