The Left Ventricle in Dogs with Myxomatous Mitral Valve Disease

Remodeling and Overall Performance

Ingrid Ljungvall

Faculty of Veterinary Medicine and Animal Science Department of Clinical Sciences Uppsala

Doctoral Thesis Swedish University of Agricultural Sciences Uppsala 2011 Acta Universitatis agriculturae Sueciae 2011:53

ISSN 1652-6880 ISBN 978-91-576-7597-2 © 2011 Ingrid Ljungvall, Uppsala Cover illustration: Ingrid Ljungvall Print: SLU Service/Repro, Uppsala 2011

The Left Ventricle in Dogs with Myxomatous Mitral Valve Disease. Remodeling and Overall Performance.

Abstract

The concept of left ventricular (LV) remodeling in dogs with myxomatous mitral valve disease (MMVD) includes changes in the LV occurring in response to mitral regurgitation (MR). The general aim of this thesis was to study LV remodeling and function in dogs with different severities of naturally acquired MMVD using 1) digital signal analysis technique for murmur and heart sound assessment, 2) biomarker analyses of circulating cardiac troponin I (cTnI), C-reactive protein (CRP), and matrix metalloproteinases (MMPs), and 3) real-time three-dimensional (RT3D) echocardiography for assessment of changes in LV shape and volume.

Digital linear and nonlinear analyses (using seven different sound variables) of cardiac sounds showed that more severe MR produced a murmur of "harsher" quality, longer duration, and with more complexity in the signal. The energy of the first heart sound was not associated with MR severity (assessed by echocardiography), whereas the energy of the second heart sound decreased with increasing MR severity. Biomarker assessments showed circulating (cTnI) concentration to increase with increasing disease severity, whereas circulating Creactive protein (CRP) concentration was not associated with disease severity. Neither MMP-2 nor -9 were associated with the MMVD severity groups (which was mainly based on volume overload status), however, MMP-9 activity decreased with worsening systolic function. The RT3D echocardiographic examinations showed prominent LV volume expansions only in dogs with more severe MMVD. The mid LV segment contributed the most to the global volume increase. The LV shape changed from elliptical to more globular in response to increasing volume overload, with the basal and apical segments contributing the most to the increase in sphericity.

In conclusion, the findings in the present thesis provide data that might contribute to the understanding of the complex pathophysiology of MMVD; thereby potentially impacting both clinical management and prediction of outcome for affected dogs in the future.

Keywords: Left ventricle (LV), remodeling, myxomatous mitral valve disease (MMVD), murmur, biomarker, real-time three-dimensional (RT3D) echocardiography.

Authors' address: Ingrid Ljungvall, Department of Clinical Sciences, SLU, Box 7054, 75007 Uppsala, Sweden. *E-mail*: Ingrid.Ljungvall@slu.se

To My Family

Contents

List of Publications		
Abbr	eviations	10
1	Introduction	13
1.1	General background	13
2	The mitral valve apparatus	15
2.1	Normal anatomy and function	15
2.2	Myxomatous mitral valve degeneration	16
2.3	Mitral regurgitation	18
3	The left ventricle	21
3.1	Myocardial histology	21
3.2	Left ventricular changes in response to mitral regurgitation	22
	3.2.1 Changes in the myocardium	22
	3.2.2 Changes in left ventricular size, shape, and function.	24
3.3	Assessment of left ventricular remodeling and function	25
	3.3.1 Heart sounds and murmurs	25
	3.3.2 Circulating biomarkers	28
	3.3.3 Echocardiography	31
4	Aims of the thesis	35
5	Materials and methods	37
5.1	Dogs	37
5.2	Methods of examinations	38
	5.2.1 Blood pressure measurement (paper II-IV)	38
	5.2.2 Assessment of heart sounds and murmurs (paper I-IV)	39
	5.2.3 Analysis of circulating cardiac biomarkers (paper II-III)	40
	5.2.4 Echocardiographic examinations (paper I-IV)	42
5.3	MMVD classification systems (paper I-IV)	45
5.4	Statistical analyses	45
6	Results	47
6.1 6.2	Signal analysis of heart sounds and murmurs (paper I)	47
	Analysis of circulating biomarkers (paper II-III)	49
		7

	6.2.1 Cardiac troponin I (paper II-III).	49	
	6.2.2 C-reactive protein (paper II-III)	49	
	6.2.3 Matrix metalloproteinase 2- and -9 (paper III).	50	
6.3	Assessment of left ventricular volume and shape (paper IV).	50	
7	General discussion		
7.1	Changes in left ventricular morphology and function		
	7.1.1 Heart sounds and murmurs	53	
	7.1.2 Circulating cardiac biomarkers	54	
	7.1.3 Left ventricular volume and shape	60	
7.2	Possible clinical implications	61	
	7.2.1 Heart sounds and murmurs	61	
	7.2.2 Circulating cardiac biomarkers	62	
	7.2.3 Left ventricular volume and shape	63	
8	Conclusions		
9	Implications for future research		
10	Populärvetenskaplig sammanfattning		
11	References		
Ackno	owledgements	89	

List of Publications

This thesis is based on the work contained in the following papers, referred to by Roman numerals in the text:

- I Ljungvall I, Ahlstrom C, Höglund K, Hult P, Kvart C, Borgarelli M, Ask P, Häggström J. 2009. Use of signal analysis of heart sounds and murmurs to assess mitral valve regurgitation attributable to myxomatous mitral valve disease in dogs. *American Journal of Veterinary Research*, 70, 604-613.
- II Ljungvall I, Höglund K, Tidholm A, Olsen LH, Borgarelli M, Venge P, Häggström J. 2010. Cardiac troponin I is associated with severity of myxomatous mitral valve disease, age, and C-reactive protein in dogs. *Journal of Veterinary Internal Medicine*, 24:153–159.
- III Ljungvall I, Rajamäki MM, Crosara S, Olsen LH, Kvart C, Borgarelli M, Höglund K, Häggström J Matrix metalloproteinase-9 is associated with systolic function in dogs with myxomatous mitral valve disease. *American Journal of Veterinary Research*. In press.
- IV Ljungvall I, Höglund K, Carnabuci C, Tidholm A, Häggström J. Assessment of global and regional left ventricular volume and shape using real-time-three-dimensional echocardiography in dogs with myxomatous mitral valve disease. Revised manuscript.
- Papers I-III are reproduced with the permission of the publishers.

Abbreviations

AUC	Area under the curve
BNP	B-type natriuretic peptide
CHF	Congestive heart failure
CKCS	Cavalier King Charles spaniel
CRP	C-reactive protein
cTnI	Cardiac troponin I
ECG	Electrocardiogram
ECM	Extracellular matrix
EDV	End-diastolic volume
ESV	End-systolic volume
HR	Heart rate
LA	Left atrium
LA/Ao	Left atrial to aortic ratio
LV	Left ventricle
LVIDd	End-diastolic left ventricular internal dimension
LVIDs	End-systolic left ventricular internal dimension
LVIDd _{inc}	Percentage increase in end-diastolic left ventricular
	internal dimension
LVIDs _{inc}	Percentage increase in end-systolic left ventricular
	internal dimension
MMP	Matrix metalloproteinase
MMVD	Myxomatous mitral valve disease
MR	Mitral regurgitation
MVP	Mitral valve prolapse
PCG	Phonocardiogram
2D	Two-dimensional
3D	Three-dimensional
RT3D	Real-time three-dimensional
RA	Right atrium
RV	Right ventricle
ROC	Receiver operating characteristic curve

SAP	Systolic arterial pressure
S1	First heart sound
S2	Second heart sound

1 Introduction

1.1 General background

The concept of ventricular remodeling, which was coined in the 1980s (Pfeffer *et al.*, 1985), is applied when describing ventricular changes occurring in response to hemodynamic changes of various etiologies (Opie *et al.*, 2006). The ventricular remodeling processes include left ventricular (LV) dilation, changes in LV shape, and LV muscle mass hypertrophy (Cohn, 1995); which all might adversely affect cardiac performance.

In 1817, Delabere Blaine described abnormal heart beats to "afford a decided characteristic of the complaint", detectable as a thrill when placing the hand on the side of the chest of a dog; thus indicating the presence of a cardiac disease (Blaine, 1817). Pathological changes of the mitral valve apparatus in dogs, possibly caused by valve degeneration, was described in the literature as early as 1935 (Münich, 1935). The knowledge in veterinary cardiology has improved tremendously since then, and assessment of cardiac diseases can nowadays be performed using various diagnostic techniques. As a result, myxomatous mitral valve disease (MMVD) has been proven the most prevalent cardiac disease in dogs (Buchanan, 1977; Whitney, 1974; Das & Tashjihan, 1965; Detweiler & Patterson, 1965), and hence, the disease most commonly causing LV remodeling in dogs. The highest disease prevalence in dogs is seen in small to medium-sized breeds, such as Cavalier King Charles Spaniel (CKCS), Dachshunds, miniature Poodles, and Yorkshire Terriers (Egenvall et al., 2006; Olsen et al., 1999; Haggstrom et al., 1992; Darke, 1987; Buchanan, 1977). The prevalence increases with age and is, at a given age, higher in males (Olsen et al., 1999; Häggström et al., 1992; Buchanan, 1977; Whitney, 1974; Das & Tashjihan, 1965; Detweiler

& Patterson, 1965). Affected dogs have no signs of valve abnormalities at birth, but develop MMVD later in life. The etiology of MMVD is currently not known, but the current leading scientific hypothesis is that a genetically determined dystrophic process, rather than succession of repeated trauma of the valve leaflets, initiates the valve degeneration (Olsen et al., 1999; Swenson et al., 1996). This hypothesis is strengthened by the knowledge that some breeds, such as the CKCS and Dachshund, are predisposed to an early onset of MMVD (Lewis et al., 2011; Olsen et al., 1999; Swenson et al., 1996; Häggström et al., 1992). The disease, which has been described to strongly resemble primary mitral valve prolapse (MVP) syndrome in people (Pedersen & Häggström, 2000), is characterized by progressive degeneration of the mitral valve (Kogure, 1980; Buchanan, 1977; Whitney, 1974). The valve degeneration leads to mitral valve leakage, referred to as mitral regurgitation (MR), and subsequently chronic volume overload with dilation of the left atrium (LA) and the LV. Progression of the disease varies between individuals, but affected dogs can usually compensate the MR for years. Eventually the heart might become incapable of meeting the increased work load imposed upon it and congestive heart failure (CHF) develops. An increased cardiac mortality before the age of 10 years has been shown in dog breeds affected by an early onset of MMVD (Egenvall et al., 2006).

The mechanisms involved in the remodeling process of MMVD remain poorly characterized and understood; partly owing to the slow progression from early degenerative changes to development of CHF; which complicates studies of disease progression. Cardiac remodeling processes in experimental settings as well as in acute vascular diseases in other species have been more extensively documented (Lang et al., 2006a; Stewart et al., 2003; Dell'Italia et al., 1995). However, these cardiovascular events differ fundamentally from naturally acquired chronic cardiac disease in origin and progression, which emphasizes the importance of investigating the LV remodeling process also in dogs with naturally acquired MMVD. Studying LV remodeling in dogs with different severities of MMVD using recently developed diagnostic techniques, such as sound analysis techniques, magnetic resonance imaging, real-time three-dimensional (RT3D)echocardiography, and high sensitivity biomarker assays, as well as more established techniques, might reveal new concepts of pathophysiological changes occurring during MMVD progression. Increased knowledge of these complex pathophysiological processes might, hopefully, be of value for improved clinical management of the individual dog affected by the disease.

2 The mitral valve apparatus

2.1 Normal anatomy and function

The mitral valve apparatus includes six anatomical elements; the posterior LA wall, the mitral orifice, the leaflets (the posterior and anterior), the chordae tendineae, the papillary muscles, and the LV wall; which all work in fine concert to maintain competence (Perloff & Roberts, 1972).



Figure 1. Normal anatomy of the canine heart.

In addition to serving as a hinge for the leaflets, the mitral orifice reduces the area required for the leaflets to bridge during ventricular systole (the ventricular contraction period) by decreasing its circumferential size (Ahmed et al., 2009; Brolin, 1967; Davis & Kinmonth, 1963). A larger surface area of the leaflets compared to the annulus area, as described in people, works as a reserve for leaflet coaptation; hence preventing backflow of blood (Ahmed et al., 2009; Perloff & Roberts, 1972). Each leaflet is supported by fibrous strands (chordae tendineae), which insert on papillary muscles arising from the apical and middle sections of the LV wall. When the LA pressure exceeds the LV pressure, the mitral valve opens and the LA empties into the LV. The mitral valve closes completely during LV systole to prevent retrograde flow. The mechanisms involved in closure of the mitral valve are not fully understood, and different events are probably involved in this process (Little, 1979). The LV filling pressure stretches the LV wall to its greatest geometric dimension (preload) during ventricular diastole. The increased intraventricular pressure during early systole forces the mitral valve to close by synergistic contraction of the LV walls; resulting in an appropriate application of vertical force to the chordae tendineae and hence, prevention of leaflet-eversion into the LA (Perloff & Roberts, 1972). Likely, significant reduction in the mitral orifice area contributes to closure of the mitral valve (Tsakiris et al., 1971; Chiechi et al., 1956). The mitral valve orifice is finally sealed when the free edges of the mitral valve leaflets firmly coapt.

2.2 Myxomatous mitral valve degeneration

Myxomatous mitral valve disease in dogs is characterized by progressive myxomatous degeneration of the mitral valve apparatus (Kogure, 1980; Buchanan, 1977; Whitney, 1974). Although the myxomatous degeneration most commonly affects the mitral valve; any of the four intracardiac valves can be affected. However, the pulmonary and aortic valves (the semilunar valves) rarely develop such degenerative changes (Buchanan, 1977). Histopathological findings include myxomatous degeneration (which refers to a characteristic pathological weakening and disturbance in the organization of the connective tissue) in which the spongiosa component is unusually prominent, and the collagen fibers are disorganized in the fibrosa layer (Hadian *et al.*, 2010; Hadian *et al.*, 2007; Black *et al.*, 2005). Proteolytic enzymes, such as the matrix metalloproteinases (MMPs), might be involved in the degenerative processes leading to atypical organization of

connective tissue components (Aupperle et al., 2009b; Aupperle et al., 2009c; Dreger et al., 2002). An increased amount of mucopolysaccarides, and glycosaminoglycans are commonly seen within affected valves (Han et al., 2010; Hadian et al., 2007; Kogure, 1980). Characteristic findings of endothelial cells covering the valve surface include pleomorphism and damage to the cell-lining. The endothelial damage, which is most commonly evident near the edges of the valve leaflets, can cause regional loss of endothelial cells; hence exposing underlying basement membranes or subendothelial matrix (Corcoran et al., 2004). Endothelial damage induces the release of vasoactive peptides, such as endothelin-1, which potentially is involved in transforming subendothelial valvular interstial cells (VICs) from a predominantly fibroblast phenotype into more active myofibroblast and smooth muscle cell phenotypes (Black et al., 2005; Corcoran et al., 2004; Mow & Pedersen, 1999). The transformation of VICs has also been suggested to be initiated by serotonin (5HT) (Oyama & Levy, 2010; Arndt et al., 2009).

The mitral valve leaflets, which normally are thin, translucent and soft, become thickened and elongated with disease progression (Han *et al.*, 2010; Corcoran *et al.*, 2004; Kogure, 1980). The chordae tendineae also become affected by the myxomatous degeneration (Corcoran *et al.*, 2004; Beardow & Buchanan, 1993); resulting in elongation (Kogure, 1980), which together with distortion of the valve architecture, contributes to systolic atrial displacement of the mitral valve leaflets (i.e. mitral valve prolapse) (Pedersen *et al.*, 1996). The thickened and elongated chordae tendineae might rupture, which potentially worsens MR by causing leaflets to partially or completely prolapse (valve flail) into the LA (Beardow & Buchanan, 1993).



Figure 2. Post-mortem specimen of a dog with end-stage MMVD. The mitral valve leaflets appear irregularly thickened and contracted. There is also evidence of chordal engagement. Chordal rupture, particularly of lesser-order chordae, is a common finding, but not clearly apparent in this image. The left atrium and left ventricle are dilated and there is evidence of jet lesions on the atrial wall (which occur when the regurgitant jet of blood from the left ventricle strikes the atrial wall)

2.3 Mitral regurgitation

The degenerative changes of the mitral valve and the corresponding chordae tendineae lead to abnormal coaptation of the mitral valve leaflets during ventricular systole; why a percentage of the LV stroke volume is ejected backwards into the LA. The retrograde ejection of LV stroke volume starts already in early systole; leading to a short isovolumetric contraction period (defined as the interval between closing of the atrioventricular valves and opening of the semilunar valves) (Lord, 1974; Eckberg et al., 1973). The extra pathway (into the LA) for stroke volume ejection reduces the LV afterload (the resistance to LV emptying). Most commonly, dogs with MMVD have a laterally directed MR jet, presumably because the anterior leaflet is longer and has a greater mobility than the posterior leaflet (as described in dogs and people) (Ahmed et al., 2009; Borgarelli, 2004), and hence, is more likely to prolapse than the posterior leaflet. However, the spatial orientation of the jet is not constant, particularly not in mild cases of MR, probably due to changes in the shape and orientation of the mitral orifice area during LV contraction (Tsakiris et al., 1971).



Figure 3. Left apical four-chamber views of the heart in a dog with mitral regurgitation caused by MMVD. A schematic drawing of the anatomy of the heart is shown in the same orientation as the echocardiograms (A). The echocardiograms show the identical image frame with the color mode off (B) and on (C). There is systolic displacement of the mitral valve leaflets (B) and valve leakage (C) as indicated by the turbulent flow (mosaic appearance on the color echocardiogram) in the left atrium. On the left ventricular side, blood is accelerating towards the regurgitant orifice, after which the flow becomes turbulent and is primarily directed laterally in the left atrium. The driving force for the jet is the pressure gradient between the left ventricle and atrium, which typically leads to flow velocities between 5.5 to 6 m/sec until late stages of the disease. Because the cross-sectional area changes from being comparably small at the regurgitant orifice to large on the atrial side, the flow becomes turbulent. In the left atrium, the kinetic energy of the jet is transformed into heat and vibrations, which are audible as a heart murmur on the thoracic wall. LA-left atrium, LV-left ventricle, RA-right atrium, RV-right ventricle.

The MR volume has been described to depend on the mitral valve orifice area, and the systolic pressure gradient between the LA and the LV (Mihalatos et al., 2007; Pierpont & Talley, 1982), of which the latter is influenced by the intra-atrial pressure, the LV function, as well as the systemic arterial blood pressure. The myxomatous degeneration of the mitral valve apparatus causes an abnormal leaflet-apposition, and hence primary MR, whereas left sided cardiac dilation exaggerates the abnormal valve apposition, leading to a secondary MR (Ahmed et al., 2009; Buchanan, regurgitation Consequently, begets regurgitation. 1977). Several mechanisms have been described involved in production of secondary MR: The LV dilation prevents the orifice from fully decreasing its circumferential size during ventricular systole (Buchanan, 1977; Brolin, 1967). In addition, the altered LV shape causes the laterally displaced papillary muscles to exert a more lateral, rather than vertical, force of exertion on the leaflets; hence further separating the mitral leaflets in systole (Hung et al., 2004; Lapu-Bula et al., 2002; Otsuji et al., 1997; Kono et al., 1992; Buchanan, 1977). Left atrial dilation might also aggravate the preexisting MR due to further leaflet displacement (Levy & Edwards, 1962). Expansion of the LA buffers the increasing MR volume, thereby allowing the intra-atrial pressure to remain comparably low, and blood can easily be ejected into the LA during ventricular systole, even in dogs with severe MMVD. More than 75% of the total LV stroke volume has been reported to be ejected into the LA during systole in dogs with severe MMVD (Kittleson & Brown, 2003). The severity of left sided cardiac dilation is linked to MR severity, suggesting that MR volume is the major determinant factor for the degree of left sided cardiac dilation (Eriksson et al., 2010; Kittleson & Brown, 2003).

3 The left ventricle

3.1 Myocardial histology

The LV myocardium in dogs has been shown to be composed of muscle layers with a characteristic three-dimensional (3D) organization, running radially across the LV wall from subendocardium to subepicardium (LeGrice *et al.*, 1995). Most of the myocardium is occupied by cardiac myocytes. Each myocyte is composed of bundles of myofibrils, which comprise longitudinally arranged microanatomical units termed sarcomeres. The sarcomeres, which represent the basic contractile units of the myocyte, are composed of thick and thin filaments; myosin and actin, respectively. Chemical and physical interactions between actin and myosin cause myocyte contraction; effected by sliding filaments along one another (Katz, 2001; Walker & Spinale, 1999).



Figure 4. Histologic section of the myocardium showing myocytes, extracellular matrix (ECM) and a blood vessel. Haemotoxylin & eosin staining. (Courtesy of Fredrik Södersten)

Structural integrity and overall geometry of the heart is maintained by the extracellular matrix (ECM), which tethers individual myocytes together in proper alignment within the myocardium, hence working as a cardiac framework (Spinale, 2002; Weber *et al.*, 1988; Robinson *et al.*, 1983). This structural organization provides tensile strength to the myocardium; enabling transduction of contractile force generated by the myocytes in systole (Pelouch *et al.*, 1993; Weber, 1989; Robinson *et al.*, 1986). The cardiac fibroblast is the most abundant cell type within the ECM and most ECM components are produced exclusively by these cells (Pelouch *et al.*, 1993; Robinson *et al.*, 1983). The fibroblasts can also secrete enzymes that regulate ECM turnover; such as the MMPs, their tissue inhibitors (TIMPs), as well as other proteolytic enzymes (Hutchinson *et al.*, 2010). Collagen subtypes and fibronectin compose the majority of the ECM proteins responsible for maintaining structural integrity of the myocardium (Pelouch *et al.*, 1993).

3.2 Left ventricular changes in response to mitral regurgitation

Mitral regurgitation results in an increased total LV stroke volume as blood is ejected both forward into the aorta and retrograde into the LA. In order to accommodate the increase in preload, the LV undergoes various compensatory responses.

3.2.1 Changes in the myocardium

Changes in the composition and structure of both the ECM and the myocytes have been reported in dogs with MR, but the underlying cellular and molecular bases for these changes remain poorly understood. The ECM is a highly adaptive structure, which has been shown to play a fundamental role in myocardial adaptation to pathological stress; thereby facilitating remodeling (Weber et al., 1992). The remodeling process of the ECM has been suggested to occur as a mechanically mediated response to stretch caused by volume overload and/or due to selective induction of proteolytic enzymes, such as the MMPs (Woessner, 1991). Normal collagen chains are fractured and replaced by poorly cross-linked collagen, resulting in loss of normal structural support and myocyte slippage (Zheng et al., 2009; Dell'italia et al., 1997; Kato et al., 1995). The myocytes are exposed to an abnormal stress-and-strain pattern during the cardiac cycle, particularily during diastole owing to increased filling pressure (preload) (Spinale, 2002; Pelouch et al., 1993; Grossman et al., 1975). This triggers an unnatural growth response with myocyte hypertrophy and replication of sarcomeres in

series; so-called eccentric hypertrophy (Grossman et al., 1975), thereby increasing myocyte length (Katz, 1990). Ventricular muscle mass is determined by the net difference between synthesis and degradation rate, and development of myocardial hypertrophy due to volume overload has been suggested to be caused by a decrease in degradation, rather than an increase in synthesis of contractile proteins (Carabello, 2002). Myocardial hypertrophy has also been suggested stiumulated by an increased neurohormonal activation, such as by increased formation of angiotensin II (AII) (Morgan & Baker, 1991; Schelling et al., 1991). In dogs with naturally occurring MR caused by MMVD, circulating levels of AII appear comparably unchanged during progression from mild MMVD to overt CHF (Haggstrom et al., 1997). However, the canine myocardium is most likely capable of forming AII locally in the heart in response to hemodynamic wall stress: Angiotensin converting enzyme (ACE), chymase and cathepsin D have all been reported capable of promoting tissue AII formation in the volume over-loaded canine heart (Stewart et al., 2003; Barlucchi et al., 2001; Dell'Italia et al., 1995).

The structural changes in the cardiac ECM and myocytes allow for chronic dilatation without overstretching of the myocytes, thereby minimizing the effects on the myocardium from the increased volume overload (Komamura *et al.*, 1993; Anversa *et al.*, 1986). The resulting eccentric hypertrophy is characterized by chamber enlargement, but with maintained relative wall thickness; sufficient to normalize pressure in the volume overloaded LV and maintain an adequate forward stroke volume (Carabello, 2002). However, the LV mass-to volume ratio has been shown to decrease with progression of MR in dogs (both naturally acquired and experimentally induced MR) and in people (Borgarelli *et al.*, 2007; Carabello, 2000; Dell'Italia *et al.*, 1995; Katz, 1995; Urabe *et al.*, 1992; Grossman *et al.*, 1975), indicating insufficient degree of hypertrophy to accommodate severe volume overload.

The compensatory mechanism, such as LV hypertrophy, dilation, and enhanced activity of the neurohormonal system, are all initially considered beneficial in order to provide the hemodynamic support needed to maintain sufficient cardiac output despite MR. However, with progression of disease, these mechanisms themselves become factors leading to deterioration of the failing heart; such as by myocyte injury and accumulation of collagen fibers (i.e. myocardial fibrosis) (Opie, 2002; Cohn *et al.*, 2000; Sabbah *et al.*, 1995; Grossman *et al.*, 1975).



Figure 5. Histologic section of the myocardium in a dog. The center of the image shows an intramyocardial artery with arteriosclerotic changes of the vessel wall. The vessel is surrounded by demarcated myocardial fibrosis. Masson trichrome staining. (Courtesy of Lennart Jönsson).

3.2.2 Changes in left ventricular size, shape, and function.

Changes in the myocardium induced by chronic volume overload in MMVD dogs will, in turn, impact size, shape and mechanical function of the heart. The LV has been suggested to obtain a more globular rather than elliptical shape with increasing volume overload, as seen in human and canine cardiac diseases of varying etiologies (Di Donato et al., 2006; Monaghan, 2006; Lord, 1974). However, sparse information is available regarding the transition into a more globular shape in dogs with naturally occurring MMVD. Interactions between myocardial structure and function exist, and the systolic function has been shown to decline in dogs with naturally acquired MMVD and in dogs with experimentally induced MR (Borgarelli et al., 2007; Urabe et al., 1992). Mechanisms underlying this deterioration are poorly understood, but various hypotheses have been suggested. Left ventricular apical rotation and twist, which have been proven of fundamental importance for cardiac performance, might be influenced by alterations in spherical geometry; thereby changing the normal pattern of contraction (van Dalen et al., 2010; Sengupta et al., 2008). Myocardial fibrosis, perhaps in combination with inadequate myocyte hypertrophy, might also contribute to the loss of force of transmission through the LV in MR dogs (Borgarelli et al., 2007; Falk et al., 2006; Urabe et al., 1992; Carabello et al., 1989; Detweiler et al., 1968). Furthermore, an overall reduction in the volume fraction of myofibrils per myocyte unit volume, as seen in dogs with experimentally induced chronic MR and in human patients with chronic cardiac diseases, could potentially be a major

contributing factor to contractile dysfunction (Spinale *et al.*, 1993; Urabe *et al.*, 1992; Zimmer *et al.*, 1992). In addition, downregulation and desensitization of the β_1 - adrenergic receptor (which is the primary adrenergic receptor in the normal heart) due to chronic overstimulation, as seen in human CHF patients (Bristow *et al.*, 1986; Bristow *et al.*, 1982) and in dogs with dilated cardiomyopathy (DCM) (Re *et al.*, 1999), could potentially also play a role in development of systolic dysfunction in dogs with advanced MMVD.

3.3 Assessment of left ventricular remodeling and function

Various diagnostic methods can be used when investigating LV remodeling and function in dogs with MMVD:

3.3.1 Heart sounds and murmurs

Heart murmurs

A systolic heart murmur is a prominent clinical finding in dogs with MMVD (Häggström *et al.*, 1995). The cross-sectional area of the mitral orifice is much smaller than the cross-sectional area of the LA, which causes turbulence on the atrial side in systole when blood is ejected backward from the LV up into the LA (*Figure 3*). In addition, changes in valvular motion and function due to myxomatous degeneration alterations, might contribute to the production of turbulence. The kinetic energy of the regurgitant jet is known to be dependent on the mass (m) of the regurgitant volume, and the velocity (v) of the jet, according to the formula $E=mv^2/2$. The kinetic energy is transformed into heat and vibrations in the LA cavity: The vibrations set particles in motion, which propagate as wave-sequences of alternating pressure toward the chest surface, where they are accessible for interpretation as a murmur.

Heart sounds

Heart murmur assessment is one of the major objectives when performing cardiac auscultation, but valuable information can also be obtained from assessment of heart sounds. The first heart sound (S1) is concurrent with the closure of the atrioventricular valves, whereas the second heart sound (S2) is concurrent with closure of the semilunar valves. Disagreement still exists regarding the origin of heart sounds and two main hypotheses have been presented (Durand & Pibarot, 1995): According to the valvular hypothesis, the heart sounds are caused by transient vibrations arising when the valves

come to a sudden halt at the end of coaptation. Alternatively, the cardiohemic hypothesis assumes the heart sounds to be created by vibrations in the whole cardiac structure. Likely, the origin of heart sounds is best described by a combination of these hypotheses (Durand & Pibarot, 1995). Regardless which of these mechanisms that best describes the origin of heart sounds; LV stroke volume and function are likely involved in the production of heart sounds, indicating a diagnostic potential of heart sound assessment when investigating LV remodeling and function in MMVD dogs.

Assessment of heart sounds and murmurs

Sounds can be described by their frequency (unit hertz; Hz), intensity (unit decibel; dB) and duration (unit ms). Frequency is a physical entity, which is perceived by the human senses in pitch (unit mel), while intensity is perceived in loudness (unit phon). However, neither frequency and pitch, nor intensity and loudness are linearly related to each other (Ahlstrom, 2008). Because a perceived increase in "murmur intensity" can be caused by either a change in frequency or a change in absolute intensity (or both),



Figure 6. Phonocardiogram (PCG) from a dog with a moderately audible murmur caused by MMVD. The recording is displayed in two modes, which are timed with respect to each other: The upper mode shows synchronous electrocardiographic (ECG) and

phonocardiographic (PCG) traces; and the lower mode shows a time-frequency graph where different frequencies are displayed according to intensity, with high-intensity frequencies in red and low-intensity frequencies in blue. Note that the murmur is composed of sounds with frequencies up to approximately 1500 Hz. S1 – first heart sound and S2 –second heart sound.

it might be discussed if the term "murmur intensity" is adequate for describing murmur severity. "Murmur audibility" is possibly a more appropriate terminology.

Previous studies evaluating heart sounds and murmurs in dogs with MR have either been carried out by subjective assessment of auscultatory findings or by assessment on standard phonocardiographic (PCG) recordings (Häggström et al., 1995; Gould et al., 1968). Acoustic stethoscopes used for auscultation propagate sounds from a chest piece, which is either of diaphragm type (covered by a membrane) or of bell type (without a membrane), through a tubing system into two ear pieces. In the electronic stethoscope, which was introduced to avoid the resonances created in the tubing system, the bell and diaphragm are replaced by a broad-band acoustic sensor and an amplifier, and the tubing and the ear pieces are replaced by wires and head phones. These improvements lead to enhanced sound quality, potentially facilitating detection of low audibility murmurs (Höglund, 2007). Some information in the cardiac sound signal is inaccessible by standard auscultation due to physical limitations in the human auditory system (Selig, 1993): The human auditory system, which is adapted to speech, has an audible range of sounds between 20 Hz and 20 kilohertz (kHz); with sounds in the frequency range between 1000 and 5000 Hz being most easily perceived. This range is much higher compared to the range of most cardiac sounds; which often are band-limited to about 10-1000 Hz (Ahlstrom, 2008). The interpretation of auscultatory findings has, furthermore, been shown highly dependent on experience and a considerable inter-observer variation in the ability to detect and interpret hearts sounds and murmurs exists (Höglund et al., 2004; Pedersen et al., 1999; Rajakumar et al., 1999; Kinney, 1988).

Phonocardiography (PCG), which is a quantitative graphic representation of cardiac sound waveforms, does not have these limitations, and the technique allows visual interpretation of cardiac sounds. Previous studies in dogs with MMVD have shown increasing severity of MR to be associated with certain characteristic features on the PCG recording: The murmur duration increases from early or late systolic to holosystolic, the murmur intensity (amplitude) increases, and there is a shift in the amplitude ratio between the first heart sound (S1) and the second heart sound (S2) (Häggström *et al.*, 1995). However, manual interpretations of PCG recordings cannot reveal detailed information about the cardiac sounds. The more recent possibility to process the recorded PCG signals by use of signal analysis technique has the potential to provide an objective and more comprehensive characterisation of heart sounds and murmurs (Ahlstrom, 2008); thereby potentially increasing information about the hemodynamic alterations occurring in the heart during different severities of MMVD.

3.3.2 Circulating biomarkers

The cardiac remodeling process stimulates release of various circulating cardiac biomarkers, and the potential of using biomarkers for diagnosing and monitoring dogs with cardiac diseases has recently gained interest. Besides the obvious utility of biomarkers for optimizing clinical management of MMVD dogs, biomarkers can also provide information of potential value to increase our understanding of the complex cardiac remodeling process.

Cardic troponins

Cardiac troponins are myofibril proteins, which by regulating the calciummediated action between actin and myosin filaments in the myocytes are crucial for muscle contraction (Katz, 2001). The cardiac troponin complex is composed of three subunits (cardiac troponin C, cardiac troponin I, and cardiac troponin T), of which cardiac troponin I (cTnI) is the only one uniquely expressed in the myocardium (O'Brien *et al.*, 1997).



Figure 7. Schematic illustration of the cardiac troponin complex. Cardiac troponin C – cTnC, cardiac troponin I – cTnI, and cardiac troponin T – cTnT.

Most cTnI is structurally bound within the myocyte, and it is released into the circulation only after myocyte injury (O'Brien *et al.*, 2006). If the rate of cTnI release exceeds the rate of synthesis, the myocardium might become partially depleted of troponins; thereby potentially affecting the contractile function (Van der Laarse, 2002). The release of cTnI from the myocardium has been shown to correlate with the amount of myocyte injury, as shown in experimental animal studies (O'Brien et al., 2006; Feng et al., 1998; Ricchiuti et al., 1997; Smith et al., 1997), and cTnI has for this reason become established as a clinical biomarker of cardiac injury. The value of cTnI as a cardiac biomarker in dogs is supported by studies demonstrating increased circulating cTnI concentrations in dogs with a variety of etiologies of cardiac injuries (Segev et al., 2008; Linde et al., 2006; Oyama & Sisson, 2004; Schober et al., 2002; Schober et al., 1999). Although cardiac troponin assays are most commonly used in the diagnosis of acute cardiovascular events, a growing interest exists in evaluating troponin concentrations in dogs with chronic cardiac diseases. Increased circulating troponin concentrations have previously been shown in dogs with MMVD (Spratt et al., 2005; Oyama & Sisson, 2004), and improvements in the sensitivity of available cTnI assays might further increase the knowledge of the cardiac remodeling process in dogs.

C-reactive protein

Stimulation with inflammatory mediators causes a time-dependent increase in LV remodeling in experimentally induced heart failure models in animals (Bozkurt et al., 1998), and changes in inflammatory pathways occurring locally in mitral valve leaflets have been described in dogs with naturally acquired heart diseases (de Laforcade et al., 2003; Mow & Pedersen, 1999). However, it is unknown if inflammatory processes contribute to the progression of cardiac remodeling in MMVD in dogs. C-reactive protein (CRP), which is an acute-phase protein mainly produced in the liver, has been shown a valuable marker of systemic inflammatory activity in various diseases in dogs (Eckersall & Conner, 1988). C-reactive protein increases rapidly with the onset of tissue destruction or inflammatory stimuli (Eckersall & Conner, 1988). In human medicine, CRP concentration is reported to be related to severity of heart failure, and to be a strong predictor of adverse outcome in cases of acute cardiovascular diseases (Sakkinen et al., 2002; Pye et al., 1990). Less is known about CRP in chronic cardiac diseases and previous studies of CRP in dogs with MMVD have shown divergent results (Tarnow et al., 2007; Rush et al., 2006).

Matrix metalloproteinases

The matrix metalloproteinases, the MMPs, is a family of zinc-dependent proteolytic enzymes known to be responsible for degeneration and remodeling of extracellular components (Woessner, 1991). The net

proteolytic activities of MMPs are regulated by the tissue inhibitors of the MMPs, the TIMPs, which form irreversible complexes with the MMPs; hence blocking access to extracellular matrix substrates (Woessner, 1991). Various cell types within the myocardium, including myocytes and fibroblasts, can express and synthesize MMPs (Hutchinson *et al.*, 2010; Coker *et al.*, 1999; Ries & Petrides, 1995). The MMPs are secreted into the extracellular space in a latent form (pro MMP), which remains enzymatically silent until activation when an amino-terminal propeptide domain is removed; resulting in the ability to degradate extracellular matrix components.



Figure 8. Schematic illustration of the matrix metalloproteinase (MMP) showing structural domains of pro MMP and active MMP (MMP-2 and MMP-9), and the transformation of the latent pro form into the active form. Modified from Vu & Werb (2000).

The MMPs contain zink at their active site, they need calcium for stability, and they are known to be activated at neutral pH. Presence of disease can stimulate activity through a number of enzymatic pathways, resulting in excessive breakdown of extracellular components (Spinale, 2002; Thomas *et al.*, 1998). The biological activation of MMPs is, however, still incompletely understood. Mast cell secretory products, such as chymases, have been suggested capable of inducing myocardial MMP activation in volume-overload states in dogs with experimentally induced chronic MR (Stewart *et al.*, 2003; Dell'Italia *et al.*, 1995). In addition, MMP activation can be induced by the membrane-bound MMPs (MT-MMPs), the extracellular matrix metalloproteinase inducer (EMMPRIN), or by various cytokines (Visse & Nagase, 2003; Spinale *et al.*, 2000; Nagase, 1997). Of the large family of MMPs, which includes stromelysins, collagenases,

gelatinases, and membrane-type MMPs, the gelatinases MMP-2 and -9 have been frequently reported involved in cardiac remodeling processes (Spinale et al., 2002). The MMP-2 and -9 possess the capacity to degrade a number of interstitial proteins, including basement membrane components, collagenes, and laminin (Vu & Werb, 2000; Nagase & Woessner, 1999; Ries & Petrides, 1995). An increased expression of genes encoding MMP-1, -2, -9 and -13 has been shown in myxomatous mitral valves in humans (Togashi et al., 2007; Rabkin et al., 2001; Soini et al., 2001). This is in contrast to findings in myxomatous mitral valves in dogs where no up-regulation of genes encoding MMP-2 and -9 has been shown (Aupperle et al., 2009c; Oyama & Chittur, 2006). Furthermore, the immunohistochemical expression of MMP-2 in myxomatous mitral valves in dogs has been shown to decrease with increasing disease severity (Aupperle et al., 2009b). Human valve samples might have been collected earlier during disease progression; at the time of surgical valve replacement, and not in severe end-stage disease; making assessment of alterations of MMP expression during disease progression difficult in humans. Additionally, this complicates comparisons of MMP results between human and canine studies. Few studies have been published investigating MMP changes in other myocardial tissue structures than valve leaflets in people and dogs with naturally acquired mitral valve disease.

3.3.3 Echocardiography

Ultrasounds are sounds of frequencies higher than 20 kilohertz (kHz), which cannot be perceived by the human auditory system. Ultrasounds are created when piezoelectrical crystals in an ultrasonographic transducer system transform electrical oscillations (of varying voltages) into mechanical oscillations (sounds). When performing an echocardiographic examination (ultrasonographic examination of the heart); sound signals are transmitted by the piezoelectrical crystals into the thorax. Sound waves are reflected back to the transducer when they encounter acoustic interfaces; generating electrical signals which are analyzed by the ultrasonographic unit. The resultant monitor image provides information about cardiac dimensions and function, as well as valvular structure and motion. In addition, Doppler technology allows determination of the velocity and direction of blood flow, which is of value when evaluating potential valve leakages and intracardiac pressure gradients.

Left ventricular anatomical dimensions and function can be assessed subjectively or by using various echocardiographic quantitative techniques. Traditional clinical echocardiographic assessments of the LV rely on 1dimensional (M-mode) and two-dimensional (2D) images. These assessments can, however, be flawed by assumptions of LV geometry, and by LV foreshortening due to image plane positioning; potentially leading to inaccuracies in measurements. These technical limitations might be pronounced when LV morphology is changed due to presence of cardiac diseases (Lang *et al.*, 2005; Kupferwasser *et al.*, 1997).



Figure 9. Illustration of the difference between 2D and RT3D image acquisition of the heart in the four-chamber left apical view. In the 2D mode (left), the image is acquired as a two dimensional slice of the heart, whereas the RT3D modality (right) allows acquisition of the image as a three dimensional pyramid. The benefit with the RT3D modality is that the volume can be rotated and cropped to visualize specific anatomic parts in three dimensions, which means that acquisition is less angle dependent than the 2D mode. The modality also allows RT3D casting of the left ventricle.

The first real-time three-dimensional (RT3D) echocardiographic system was introduced in the early 1990ies (Sheikh et al., 1991), and further

improvements in design and engineering have led to the recent commercialization of RT3D echocardiographic systems. Modern RT3D echocardiographic systems utilize high-frequency transducers that consist of more than 3000 individual crystals, which simultaneously acquire data in a 3D pyramidal fashion in real time. The technique, which when including time can be referred to as four-dimensional, allows for superior anatomical delineation of the LV in real time. Due to the ability to manipulate the plane to align the true short- and long-axes of the LV, the problems with chamber foreshortening and oblique imaging planes can be reduced; thus providing more anatomically correct views than conventional 2Dechocardiography (Lu et al., 2008; Jacobs et al., 2006; Lang et al., 2006b). All voxels (i.e. volumetric picture elements) representing intensity of echocardiographic reflections at a particular point in space are used in the LV border detection process when creating a global LV RT3D volume dataset. The RT3D dataset can further be divided into 17 regional segments by sectioning the LV from base to apex, perpendicular to the LV long-axis; thereby allowing regional volume assessment (Cerqueira et al., 2002). Abnormal changes in LV shape, accompanying LV dilation, can be assessed by an echocardiographically derived sphericity index (Di Donato et al., 2006; Monaghan, 2006). A 3D echocardiographically derived sphericity index has been demonstrated an earlier and more accurate predictor of remodeling compared to other echocardiographic variables following acute myocardial infarction in human patients (Mannaerts et al., 2004). The sphericity index is usually not included in the routine echocardiographic protocol when assessing LV remodeling in MMVD dogs, and this index might have a potential when investigating progression of remodeling in dogs with MMVD.

4 Aims of the thesis

The general aim of this thesis was to study LV remodeling and function in dogs with different severities of naturally acquired MMVD using both recently developed and previously established diagnostic techniques in order to further explore the complex pathophysiology of MMVD. Increased knowledge of the pathophysiological processes occurring during disease progression could potentially impact both clinical management and prediction of outcome for the individual dog in the future.

The specific aims were to:

- Investigate whether linear and nonlinear signal analyses of cardiac sounds could be used to assess MR severity.
- Investigate whether plasma concentrations of cTnI and CRP were associated with disease severity.
- Investigate whether plasma activities of MMP-2 and -9 were associated with disease severity.
- Investigate how the LV changes in shape and volume in response to increasing disease severity using RT3D echocardiography.
5 Materials and methods

This section summarizes and comments on the material and methods used in the separate papers included in this thesis. More detailed descriptions of the procedures performed are presented in the separate papers.

5.1 Dogs

All studies included in this thesis were approved by the Local Ethical Committee in Uppsala, Sweden. Client-owned dogs were prospectively recruited at the cardiology unit of the Faculty of Veterinary Medicine and Animal Sciences in Uppsala, and informed owner consent was obtained. The number of dogs occurring in more than one of the included papers is summarized in Table 1.



Table 1. Number of dogs included in the different studies, and shared by two studies of the present thesis. Numbers within brackets represent dogs which were reexamined in study IV, but had been included and examined in previous studies approximately two years earlier.

As inclusion criteria for the studies, dogs had to either have evidence of MMVD or be free from physical and echocardiographic evidence of cardiac disease. Dogs of all breeds were allowed into the study provided that they fulfilled the inclusion criteria. Dogs with congenital heart disease, other acquired cardiovascular disorders or significant organ-related or systemic diseases were not included in the studies. Dogs in need of heart failure therapy in order to prevent clinical signs were allowed into the studies. Furthermore, included dogs had to have a body weight less than 15 kg: Occasionally, large-breed dogs have coexisting MMVD and dilated cardiomyopathy (DCM), and excluding larger dogs from the studies limits the risk of mixing multiple cardiac diseases in the study populations. In addition, some echocardiographic variables might be dependent on body size (Bonagura & Schober, 2009). Hence, interpretation of results was considered easier and more reliable in a more homogenous group of dogs.

A majority of dogs in paper II were also included in paper III. However, pregnant bitches, dogs treated with glucocorticoids, and dogs with detectable neoplasms were excluded from study III due to a possible influence on MMP results (Schafer-Somi *et al.*, 2005; Loukopoulos *et al.*, 2003; Dollery *et al.*, 1995); hence six dogs from study II were excluded from enrolment in study III due to ongoing glucocorticoid therapy or presence of small mammary tumors.

5.2 Methods of examinations

All examinations were performed without sedation in a quiet examination room. Dog-owners were present during all examination procedures in order to keep the dogs comfortable and calm. The procedures included (presented in the order of which they were performed in the studies) an ownerinterview (paper I-IV), blood pressure measurement (paper II-IV), physical examination including cardiac and pulmonary auscultation (paper I-IV), electrocardiographic (ECG) and PCG recordings (paper I), venous blood collection (paper II-III), and echocardiographic examinations (paper I-IV)

5.2.1 Blood pressure measurement (paper II-IV)

Blood pressure measurement was performed at the beginning of the examination protocol; after the dog had been adapted to the environment for approximately 10-15 minutes; all in order to reduce the influence of stress on the blood pressure results. Blood pressure was indirectly measured using automated oscillometric technique. In paper II-III, a standard

oscillometric device (Oscillometric Krutech VET420A, Jorgen Kruuse A/S, Marslev, Denmark) was used, whereas a high definition oscillometric (HDO) device (Vet HDO monitor, S +B medVet GmbH, Babenhausen, Germany) was used in paper IV. For all measurements, dogs were minimally restrained in a standing position, and an appropriate neonatal cuff, with a width of approximately 40% of the tail circumference, was applied to the base of the tail with the artery marker placed ventrally. Once reliable consecutive readings were obtained, the mean of five consecutive blood pressure measurements was calculated (Brown *et al.*, 2007).

5.2.2 Assessment of heart sounds and murmurs (paper I-IV)

Cardiac auscultation (paper I-IV)

Cardiac auscultation was conducted in a quiet examination room with the dog in a standing position. A Welsh Allyn Meditron sensor-based electronic stethoscope (Meditron ASA, Medi-Stim ASA, Oslo, Norway) was used for auscultation in all papers. The audibility of detectable heart murmurs (which all had to have the point of maximal audibility located over the mitral valve area; at the costochondral junctions between the fifth and sixth intercostal spaces on the left side of the chest wall) was graded on a scale from I-VI in accordance with established guidelines (Gompf, 1988): (Grade I murmur is very faint and only heard with special effort, while a grade VI is extremely load and accompanied by a palpable thrill on the thoracic surface).

Signal analysis of heart sounds and murmurs (paper I).

The electronic stethoscope was connected to a laptop computer (Dell latitude D800 laptop, Dell Computer Corp, Limerick, Ireland) with accompanying acquisition software (Meditron analyzer, version 4.0V, Welch Allyn Meditron ASA, Medi-Stim ASA, Oslo, Norway), for recording of the PCG signals. During recording, the flat acoustic sensor of the electronic stethoscope was placed firmly over the point of maximal audibility over the mitral valve area to provide the loudest and clearest heart murmur possible. An *ECG* (lead II) was recorded simultaneously with the PCG. Each recording lasted for 10 seconds. Background noise was minimized, and the mouth was gently closed during the recording in panting dogs; to reduce ventilation artifacts.

All recorded phonocardiographic signals were manually segmented using the ECG recordings as an aid for timing of the heart sounds. Four markers (beginning of S1, end of S1, beginning of S2 and end of S2) were determined for each heart cycle. Noisy or corrupted signal segments (as

determined by visual inspection of the data) were excluded from further analyses. All processing of PCG signals was performed by use of a mathematical computer program (MATLAB, version 7.3, The MathWorks Inc, Natick, Mass., USA), which was programmed to automatically derive seven sound variables from the segmented PCG signals. While regular frequency sounds can be investigated using linear techniques, non-linear techniques are required when investigating the complexity of sounds: Both linear and non-linear techniques were used in paper I. In brief: 1) The first frequency peak represents the dominant frequency component in the signal. 2) The murmur energy ratio quantifies the percentage of higher frequencies in the spectrum (Nygaard et al., 1993), and is defined as the energy between 50-500 Hz divided by the energy between 20-50 Hz. 3) The murmur duration>200 Hz measures the duration of sounds exceeding 200 Hz as a fraction of the length of systole. 4) The sample entropy is a complexity measure useful for investigating dynamics of time series (Richman & Moorman, 2000). 5) The auto mutual information function represents the predictability of a signal 6) The energy ratio of S1 represents the normalized energy within the S1 segment. Since it is impossible to measure an absolute sound audibility, the variable was normalized against the energy in diastole as outlined by Durand et al (1990). 7) The energy ratio of S2 represents the energy within the S2 segment normalized against the energy in diastole.

5.2.3 Analysis of circulating cardiac biomarkers (paper II-III)

Blood sampling (paper II and III)

Blood was collected by jugular venipuncture into 5-mL tubes containing EDTA, and the plasma was separated by centrifugation within 30 minutes of collection. Plasma was transferred to 1.5 mL plastic cryotubes, and all samples were stored at -80°C for batched analyses. The frozen plasma was thawed slowly at room temperature prior to analysis.

Cardiac troponin I (paper II and III).

Concentrations of cTnI were analyzed in duplicate using a recently refined, enzyme-linked immunosorbent assay (Access® Systems AccuTnI® Assay, Beckman Coulter, Inc., Fullerton, California, USA), according to the manufacturer's instructions. The troponin amino acid sequence is highly conserved across species, allowing the use of human immunoassays for analysis of canine samples (O'Brien *et al.*, 2006;Oyama & Sisson, 2004). In addition, an in-house validation was performed by a trained laboratory technician, and the tested dilutional parallelism of canine plasma confirmed linearity within the assay system. The lower limit of detection for the assay used in the present study was 0.001 ng/mL, which is an approximately 10-100 times higher sensitivity compared to most assays used in clinical settings today.

C-reactive protein (paper II and III)

Concentrations of CRP were analyzed in duplicate using a previously validated (Kjelgaard-Hansen *et al.*, 2003) commercially available canine CRP ELISA assay (Tridelta PhaseTM Range CRP – Canine Assay, Tridelta Development Ltd, County Wicklow, Ireland), according to the manufacturer's instructions. An in-house validation confirmed dilutional parallelism linearity within the assay system.

Matrix metalloproteinase 2- and -9 (paper III).

Plasma MMP activity was analyzed using gelatin zymography, as previously described (Rajamäki *et al.*, 2002). Zymography is an electrophoretic technique that includes a substrate copolymerized with a polyacrylamide gel for detection of enzyme activity. After preparation and incubation, the zymogram was stained and washed, and the gelatinolytic activity was revealed as clear bands against a darkly stained background (where the enzyme had degraded the substrate). For quantification of gelatin degradation, the gels were scanned, background grey values were subtracted, and the densitometric results were measured. Due to the risk of variation between each gel, the densitrometric results of each band were assessed by comparison with the activity of the pro MMP-2 band in standard lane on each gel. All zymograms were analyzed for pro- and active MMP-2 and -9 forms.



Figure 10. A zymogram from a dog showing gelatinolytic activity of the matrix metalloproteinases pro MMP-2 and -9 and active MMP-2 and -9, which are revealed as clear bands against a darkly stained background. (Courtesy of Minna Rajamäki).

5.2.4 Echocardiographic examinations (paper I-IV)

Dogs were gently restrained in both right and left lateral recumbency on an ultrasound examination table. Echocardiographic examinations were performed by use of a GE Vivid 3 ultrasonographic unit (General Electric Co, Stockholm, Sweden) equipped with a 5-MHz transducer in paper I-III, and a Philips iE33 ultrasonographic unit (Philips Ultrasound, Bothell, WA, USA) equipped with a 5-1 MHz transducer (for M-mode and 2D), and a 7-2 MHz matrix transducer (for 2D and RT3D) in paper IV. Continuous ECG (lead II) monitoring was performed during the echocardiographic examinations in all studies.

M-mode, 2D and Doppler examinations (paper I-IV)

M-mode and 2D loops of standardized views (Thomas *et al.*, 1993) were digitally stored in all studies. Screening of potential regurgitations through the mitral, tricuspid, aortic, and pulmonic valves was performed using color Doppler echocardiography. Assessment of mitral valve structures and MR severity was conducted from the right parasternal long-axis view and the left apical four-chamber view. Demonstrated MR on the echocardiogram was subjectively assessed as the area of regurgitant jet relative to the area of the

left atrium, as previously described (Olsen *et al.*, 2003; Pedersen *et al.*, 1996). The left atrial to aortic root (LA/Ao) ratio was measured as previously described (Hansson *et al.*, 2002). M-mode measurements of the LV were performed using standard techniques (Bélanger, 2005) on images obtained from the right parasternal short axis view. The LV internal dimensions were related to body size by allometric scaling of body weight; and the values for the percent increases of end-diastolic left ventricular internal dimension (LVIDd_{inc}) and end-systolic left ventricular internal dimension (LVIDs_{inc}) were calculated as follows: [observed dimension – expected normal dimensions were calculated as previously described (Cornell *et al.*, 2004): LVIDd (body weight^{0.294} × 1.53), and LVIDs (body weight^{0.315} × 0.95).

Real-time three-dimensional echocardiography (paper IV).

Real-time three-dimensional (RT3D) datasets were acquired from the left parasternal apical four-chamber view. Series of four to seven consecutive ECG R-wave triggered cycles were acquired, from which four sub-volumes were automatically derived and integrated into one pyramidal volume; thereby providing a RT3D full-volume dataset of the entire LV.

The RT3D data analyses were performed off-line using a commercial software (QLAB advanced quantification, version 7.0, Philips Ultrasound, Bothell, WA, USA), which displays the RT3D volume data in three different orthogonal planes; the two-and four-chamber views, and the shortaxis view. The orthogonal planes were all aligned interactively by use of color-coded conventions, according to the manufacturer's instructions, in order to obtain the most anatomically correct apical views for optimal border delineation of the LV. Anatomic landmark definition was performed manually at the endocardial border in the LV cavity in the end-diastolic volume (EDV) frame. Automatic endocardial border tracing created a cast of the LV cavity, and the LV cavity border detection was then verified for accuracy in each view, and manually adjusted as required. The procedures described for the EDV frame were repeated on the end-systolic volume (ESV) frame. Finally, the traced borders were processed to automatically calculate the EDV and ESV by an algorithm model in the software. The cardiac volumes were indexed to body weight (volume); EDV/kg and ESV/kg, based on the presumption of a linear relationship between cardiac volumes and body weight (Bonagura & Schober, 2009; Cornell et al., 2004). The RT3D long-axis length was measured in the four-chamber view as the distance from the endocardial apex to the mid-point of the mitral valve.



Figure 11. Example of a real-time three-dimensional (RT3D) volume dataset from a dog displayed in three orthogonal planes; the two- (upper right) and four-chamber views (upper left), and the short axis view (lower left), from which endocardial borders of the LV cast were identified, and end-diastolic and end-systolic LV volumes were obtained. The left ventricular 3D cast is automatically divided into 17 segments according to established guidelines (lower right). For the purpose of this thesis, the 17 segments were further joined into three major segments: basal, mid and apical segments.

Seventeen segments, defined by the American Society of Echocardiography (Cerqueira *et al.*, 2002), were automatically calculated from the LV cast, allowing regional RT3D LV volume assessment (*Figure 2*). The different segments were identified on the LV cast as fractions of the LV long axis length. The 17 segments were further joined into three major segments: Basal EDV and ESV included segments 1–6; mid EDV and ESV included segments 7–12; apical EDV and ESV included segments 13–17. Percentage contributions to the global EDV and ESV of each of the major regional segments, were calculated (basal, mid and apical EDV % and ESV %).

Sphericity index was calculated as the RT3D-EDV divided by the volume of a sphere, with the sphere volume calculated as $1/6 \pi \times L^3$ (where L is equal to the LV long-axis length) (Di Donato *et al.*, 2006).

5.3 MMVD classification systems (paper I-IV)

Criteria for the diagnosis of MMVD included characteristic valvular lesions of the mitral valve apparatus (thickened and/or prolapsing mitral valve leaflets) and demonstrated MR on color Doppler echocardiogram

All dogs included in the four papers were classified using an echocardiographic classification system: Estimation of disease severity was based on the obtained LA/Ao ratio and the MR jet size, and dogs were classified as follows: Healthy; LA/Ao < 1.5 and no MR jet, mild; LA/Ao \leq 1.5 and MR jet < 30%, moderate; LA/Ao > 1.5 and < 1.8 and MR jet < 50%, and severe; LA/Ao \geq 1.8 and MR jet > 50%.

The echocardiographic classification system used in paper IV differed slightly from the above described classification system. There is a problem in interpretation of small MR jets on the color echocardiogram: Some are likely to represent early stages of MMVD, whereas others may be trivial nonpathologic jets (Nakayama et al., 1994; Perry & Bouchard, 1990). Improvements of the Doppler technique in new ultrasonographic units might increase the likelihood of detecting minimal MR signals. Hence, the use of the new ultrasonographic unit in paper IV might have increased detection of with minimal MR of which a proportion most likely was trivial nonpathologic jets. Hence, dogs with minimal MR and LA/Ao < 1.5 were allowed into the healthy classification group in paper IV. Because minimal jets could create sounds influencing the sound signal variables investigated in paper I; dogs having minimal MR jets were classified as mild in this study. Some of the dogs included in paper I were also included in paper II and III, and the same echocardiographic classification system was applied in these papers. The murmurs were analyzed by sound signal analysis techniques in paper I, and it was therefore of interest to relate the results obtained using this technique to traditional auscultation assessments performed by veterinarians experienced in veterinary cardiology. Hence, an auscultatory classification system was also applied (in addition to the echocardiographic classification system) in paper I. For the auscultatory classification, dogs were divided into the following groups: normal (no audible cardiac murmur), low audibility (grades I-II), moderate audibility (grades III-IV) and high audibility (grades V-VI) murmurs.

5.4 Statistical analyses

Data are presented as medians and interquartile ranges (IQR). A value of P < 0.05 was considered significant for the analyses, unless otherwise indicated.

The Cuzick test for ordered groups (paper I) and Kruskal-Wallis test (paper II-IV) were used to investigate overall associations between the investigated variables (sound signal variables, circulating biomarkers and RT3D echocardiographic variables) and the four MMVD severity groups. In variables in which a significant association was detected, a pair-wise comparison was also performed by use of the Mann Whitney U-test with Bonferroni adjustment: Comparing four groups to each other involves six comparisons, resulting in a significant *P*-value of < 0.008.

Unilinear (paper I-IV) and multiple regression (paper I-III) analyses were used to evaluate associations between the variables of interest (sound signal variables, circulating biomarkers and RT3D echocardiographic variables), and dog characteristics, HR obtained from the echocardiogram, systolic arterial pressure (SAP), and M-mode and 2D echocardiographic measurements. In the multiple regression model, analyses were performed in a backward stepwise manner (Bland, 1995), starting with all variables included in the model and then removing the variable with the highest *P*value until all the remaining variables had a value of P < 0.05. All variables were assessed only as main effects; no interaction terms were considered in the model. The adjusted R^2 is defined as the percentage of the total sum of squares that can be explained by the regression and it also considers the degrees of freedom for variables added.

In paper I, linear discriminant analysis (Fischer, 1936) was used to investigate whether a combination of sound variables could be used to distinguish severe MR from the other three severity groups. The diagnostic efficacy of the optimal combination of sound variables (which were obtained from the linear discriminant analysis) was further evaluated by use of receiver operating characteristic (ROC) curves. In particular, sensitivity, specificity, and area under the curve (AUC) were investigated.

6 Results

This section summarizes the results from the separate papers included in the thesis. More detailed descriptions of the results are presented in the separate papers.

6.1 Signal analysis of heart sounds and murmurs (paper I)

The box and whisker plots in *Figure 12* show the sound variables evaluated against the auscultatory and echocardiographic classification systems. In brief: The first frequency peak, and consequently the frequency spectrum, was shifted towards higher frequencies with more severe MR. More severe MR produced a murmur with "harsher" quality, as shown by higher murmur energy ratio. Analysis of murmur duration > 200 Hz showed that murmurs shifted from early or late systolic to holosystolic with increasing MR severity. More severe MR showed more irregularity in flow behavior, as indicated by higher values of sample entropy. Lower values of auto mutual information were found in dogs with severe MR, reflecting decreasing predictability of the sound signal with increasing signal complexity. The energy of S1 decreased with increasing murmur audibility, but showed no association with MR severity using the echocardiographic classification system. The energy of S2 was found to decrease with increasing MR severity.



Figure 12. Box-and-whisker plots of seven sound variables evaluated against the auscultatory (left column) and echocardiographic (right column) classification systems. The upper and lower limits of each box represent the lower quartile and upper quartile values, respectively; the horizontal line within each box represents the median. The whiskers represent the extent of the data (1.5 times the interquartile range). Outliers are indicated (plus signs). Because of the low number of dogs in the clinically normal group using the echocardiographic classification system, these dogs were excluded from the multiple group-wise comparison tests. *Values indicated by brackets differ significantly (P < 0.008).

Using the seven sound variables as dependent variables and dog characteristics (age, gender, breed and body weight), heart rate and LA/Aoratio as independent variables in a multiple regression model, confirmed a major effect of the LA/Ao on the sound variables, but showed an absence of effect of the other variables included in the model. The first frequency peak was the sound variable giving the highest model $R^2 (R^2 = 0.40, P < 0.001)$.

The linear discriminant analysis demonstrated that the optimal combination of sound signal variables (smallest variable set with the largest possible amount of correct detections) was the selection of energy ratio of S2, auto mutual information and first frequency peak, resulting in a sensitivity of 88% a specificity of 82% (using the echocardiographic classification system). The ROC curve, which summarizes the diagnostic performance of a test, had an AUC of 0.89 for this combination.

6.2 Analysis of circulating biomarkers (paper II-III)

6.2.1 Cardiac troponin I (paper II-III).

Detectable concentrations of cTnI were found in 67 % of the included dogs. Plasma cTnI showed either barely detectable concentrations, or concentrations below the lower limit of detection for the assay in the healthy dogs, but cTnI concentration increased with increasing MMVD severity. Uni- and multiple regression analyses showed a major effect of age, CRP concentration, HR, and LVIDd_{inc}% on cTnI concentration, and the final multiple regression model had an adjusted R^2 of 0.63 (P< 0.0001). Age was the variable most strongly associated with cTnI in the unilinear analyses ($R^2 = 0.50$, P < 0.0001). Because of the strong association between age and cTnI concentration, further investigations were performed within the four different MMVD groups: Concentration of cTnI increased significantly with increasing age in the mild MMVD group ($R^2 = 0.47$, P < 0.0001), which included nearly 50% of the dogs in the study population, and in the severe MMVD group ($R^2 = 0.26$, P = 0.018).

6.2.2 C-reactive protein (paper II-III)

Detectable concentrations of CRP were found in all included dogs.

No significant difference was shown between CRP concentration and the four MMVD severity groups, but CRP concentration was associated with cTnI concentration, breed, and systolic blood pressure in the multiple regression analysis. However, the regression model had a comparably low model R^2 ($R^2 = 0.24$, P < 0.0001), and slight modifications in the order the

variables were removed in the backward analysis process had a comparably large effect on the outcome, indicating an unstable model.

6.2.3 Matrix metalloproteinase 2- and -9 (paper III).

Zymography of the plasma samples revealed gelatinase expression of pro MMP-2 and pro MMP-9 activities in all included dogs, and active MMP-9 was found in 85 % of the dogs. Active MMP-2 could not be detected in the study population. No overall significant differences were found between MMP activity, and the four MMVD severity groups.

Uni- and multiple regression analyses showed that pro MMP-9 decreased with decreasing SAP, and was higher in male dogs than in female dogs. However, the final multiple regression model was weak with an adjusted R^2 of 0.14. Systolic arterial pressure (SAP) was the variable most strongly associated with pro MMP-9 in the unilinear analyses ($R^2 = 0.10$, P < 0.0037).

Uni- and multiple regression analyses showed that active MMP-9 decreased with increasing LVIDs_{ine}, and with decreasing cTnI, SAP, and age (the latter did only reach a significant level in the unilinear analysis). The final multiple regression model had an adjusted R^2 of 0.29. Left ventricular end-systolic dimension was the variable most strongly associated with active MMP-9 in the unilinear analyses ($R^2 = 0.11$, P = 0.0039).

Pro MMP-2 activity was not significantly associated with any of the investigated variables in the uni- or multiple regression analyses.

6.3 Assessment of left ventricular volume and shape (paper IV).

Dogs with severe MMVD had higher values of EDV, ESV, long-axis length, and sphericity index, compared to dogs in the other MMVD severity groups (*Figure 13*). Dogs with moderate MMVD had higher contribution of the mid EDV segment to the global EDV, compared to values in dogs with mild MMVD.

Unilinear regression analyses showed that global and regional EDV and ESV, long-axis length, and sphericity index increased with increasing disease severity, as indicated by increasing LA/Ao ratio, LVIDd_{inc}%, and LVIDs_{inc}%. In addition, SAP decreased with increasing EDV and ESV. Even though all three regional LV segments contributed to the increase in global EDV and ESV with increasing MMVD severity, the mid EDV contributed the most to the global EDV increase. Assessing regional contribution to changes in LV shape; sphericity index was associated with decreasing percentage

contribution of basal EDV, and increasing percentage contribution of apical EDV to global EDV.



Figure 13. Left ventricular end-diastolic casts obtained from the RT3D dataset from a healthy dog (left) and a dog with severe MMVD (right). The diseased heart has a more rounded left ventricular appearance (and thereby an increased sphericity index), in addition to a globally increased left ventricular volume.

7 General discussion

The cardiac remodeling process in dogs with MMVD is highly complex, with different processes occurring simultaneously, and most likely with varying degrees of contribution during disease progression. Some results from the studies included in the present thesis might primarily be important for understanding the pathophysiological changes occurring during the LV remodeling process, whereas others could potentially be more directly valuable for improving the assessment of MMVD severity; and hence, the clinical management of affected dogs.

7.1 Changes in left ventricular morphology and function

7.1.1 Heart sounds and murmurs

The murmur-audibility has previously been shown to increase with severity of MMVD (Häggström et al., 1995), but signal processing of murmurs has, to the best of the author's knowledge, never been applied for assessment of MR in any species. Quantification of MR can provide valuable information about LV remodeling status and function in MMVD dogs. In paper I, duration of systolic frequency contents exceeding 200 Hz was nearly nonexistent in normal dogs (recorded cardiac acoustic signals can be obscured by respiratory sounds, rumbling sounds from the stomach, friction rubs, and ambient sounds; explaining the existence of detectable sound signals in systole; even in the absence of a systolic murmur), but increased in duration with increasing MR severity: Dogs with moderate to severe MMVD had long systolic durations of frequencies exceeding 200 Hz. These findings indicate a maintained energy in the MR jet in late stages of the disease. In the typical MMVD dog, the maximal systolic flow velocity of the MR remains comparably constant until late stages of the disease when it might decrease (Olsen et al., 2010); likely as a consequence of altered

intracardiac pressure gradients due to LV systolic dysfunction and increased intraatrial pressure. But even though LV systolic function worsens in late stages of the disease (Borgarelli *et al.*, 2007), sufficient function remains to eject a large MR volume into the LA; thus generating a holosystolic high frequency murmur. The large retrograde stroke volume in dogs with more severe MR creates a murmur of "harsher" quality and with higher frequency content, indicated by an increase in the first frequency peak, and murmur energy ratio, in these dogs. In addition, the results from the nonlinear investigations using sample entropy and auto mutual information indicate that more severe MR has more irregularity in the flow behavior, and thus less predictability in the signal.

The intensity of S1 has commonly been regarded to increase with increasing disease severity in dogs with MMVD. Remarkably, in paper I, the energy of S1 was shown to decrease with increasing murmur audibility, and showed no association with MR severity using the echocardiographic classification system. Merging of the S1 with the forceful murmur might give an impression of increased audibility of the S1 when subjectively assessed by cardiac auscultation, possibly explaining previous reports of increased S1 audibility with increasing MR severity (Gould et al., 1968). The relative intensity between S1 and S2 has been shown to change with increasing MMVD severity when manually measured on PCG recordings (Häggström et al., 1995), but an objectively performed study of potential heart sound alterations caused by MR has, to the author's knowledge, never been published. A possible mechanism explaining the decrease in S1 energy with increasing murmur audibility is that the degenerated valve might influence the vibrations involved in the origin of S1. A reduction in LV systolic function with disease progression could potentially also influence the S1. The energy of S2 was found to decrease with increasing MR severity, which is in accordance with a previous PCG study in MMVD dogs (Häggström et al., 1995). The intensity of S2 is reported to primarily depend on the rate of change in the pressure gradient across the aortic valve at closure (Sabbah & Stein, 1976), and a diminished forward stroke volume (Kittleson & Brown, 2003) might explain the decreased energy ratio of S2 seen in dogs with increasing severity of MR.

7.1.2 Circulating cardiac biomarkers

Cardiac troponin I

To the best of the author's knowledge, this was the first study exclusively designed to investigate the association between cTnI concentration and

MMVD severity in dogs. Potential effects of CRP and dog characteristics on cTnI concentration were controlled for in the study. Plasma cTnI concentration was found to increase with increasing MMVD severity, which indicates ongoing myocyte injury in a chronic remodeling process. However, the exact cause of such processes remains unknown. Intramural myocardial arteriosclerosis; with loss of compliance in the arterial vascular walls due to luminal narrowing, has been described in dogs in CHF due to MMVD (Falk et al., 2010; Falk et al., 2006). These changes might affect the vascular supply to the myocardium; ultimately promoting regional hypoxia and myocyte death (Sabbah et al., 1995; Detweiler et al., 1968). The arteriosclerotic changes have furthermore been shown associated with myocardial fibrosis in MMVD dogs (Falk et al., 2010; Falk et al., 2006; Detweiler, 1989; Jönsson, 1972; Detweiler et al., 1968) supporting the hypothesis of arteriosclerosis-induced myocyte death as a causative factor in development of myocardial fibrosis in MMVD dogs. Yet, a reversed scenario is possible: Remodeling of the ECM might increase the amount of myocardial fibrosis, which by reducing the capillary density in the myocardium; and thereby, the oxygen diffusion distance, damage myocyte integrity (as seen in dogs with experimentally induced CHF) (Sabbah et al., 1995). The oxygen demand might furthermore be increased in the hypertrophied myocardium; potentially lowering the threshold for myocardial hypoxia (Piano et al., 1998). Such a scenario, if present, is likely to occur intermittently or be of subclinical character in the clinical situation because most dogs with MMVD do not present typical changes indicative of hypoxia on the ECG. Myocyte loss can also be a result of programmed cell death (apoptosis) in the hypertrophied myocardium in dogs (Sharov et al., 1996), potentially due to increased levels of aldosterone, norepinenephrine, AII, and various inflammatory mediators (Ferrari et al., 1998; Sabbah et al., 1995): However, it is not established if their concentrations are sufficiently high to promote direct myocyte necrosis in the failing heart in dogs with MMVD.

Paper II described that the cardiac release of cTnI started in the early stages of MMVD in dogs. The true biological half-life of cTnI has been reported to be approximately 70 min in dogs (Jaffe *et al.*, 1996). Thus, the increase in cTnI concentration was most likely due to an ongoing release of cTnI caused by a continuous remodeling process, rather than an acute process. Minimal myocardial cTnI loss might be of minor importance for contractile function in the short term. However, chronic myocardial degradation and long-term loss of myocardial troponins has been suggested to affect LV contractile function (Van der Laarse, 2002).

A strong association between age and cTnI concentration was found, indicating that age causes myocardial changes leading to cTnI leakage. In fact, age was the variable most strongly associated with cTnI concentration. An association between age and cTnI has previously been reported in healthy dogs (Oyama & Sisson, 2004), people (Venge et al., 2003), and rats (O'Brien et al., 2006). Intramural arteriosclerotic changes have been shown associated with normal aging in dogs, potentially causing defects in the oxygenation potential in the myocardium in elderly dogs; and thereby myocyte injury or loss (Whitney, 1976; Jönsson, 1972; Detweiler & Patterson, 1965). A marked myocyte loss has been reported to occur with age in humans (more evident in males) potentially explained by ischemic injuries (Olivetti et al., 1995; Olivetti et al., 1991). A certain degree of cardiac fibrosis has also been seen in the aging heart in various species (Lakhan & Harle, 2008; Thomas et al., 2000; Villari et al., 1997; Klima et al., 1990; Mukherjee & Sen, 1990), but the actual prevalence of myocardial fibrosis in elderly dogs is not known. Because changes associated with normal aging might be difficult to separate from those caused by cardiac diseases, the effect of age on cTnI needs consideration when evaluating this biomarker in dogs. Hypothetically, age-related changes in cardiovascular structure and function might reduce the ability to compensate for cardiac diseases in elderly individuals.

C-reactive protein

Concentration of CRP was not associated with MMVD severity in the examined dogs. All dogs had detectable circulating concentrations of CRP, but based on previous reported normal variations in healthy dogs (Mischke et al., 2007; Kjelgaard-Hansen et al., 2003) dogs with clinically important systemic inflammation were not represented in the study population; as all dogs had CRP concentrations within the normal range. A multiple regression model showed significant associations between CRP concentration and cTnI concentration, breed, and SAP. However, a comparably low model R^2 , in combination with the knowledge that CRP can increase due to tissue destruction or inflammatory stimuli in other organs, suggest that CRP is not a sensitive biomarker for evaluation of MMVD remodeling in dogs. Yet, in order to further evaluate the role of inflammation in the pathogenesis of MMVD, further investigations, including other markers of inflammation, are needed.

Matrix metalloproteinase 2- and -9

Changes in the myocardial structure with loss of the fine collagen weave surrounding the myocytes might occur due to selective induction of MMPs (Spinale, 2002; Woessner, 1991). Previous reports on MMPs in dogs with naturally acquired MMVD have focused on changes in the mitral valve leaflets (Aupperle et al., 2009a; Aupperle et al., 2009c; Oyama & Chittur, 2006). To the best of the authors's knowledge, paper III is the first study exclusively designed to investigate circulating activity of MMPs in different severities of MMVD in any species. Circulating activity of pro- and active MMP-2 and -9 were investigated, and activity of MMP-9 was shown to decrease with increasing LVIDs, % and decreasing SAP. Hence, activity of MMP-9 was, in contrast to cTnI, linked to variables reflecting systolic function, but not to variables reflecting LV dilation; indicating that MMP-9 and cTnI might reflect different aspects of the complex cardiac remodeling process in MMVD dogs. The MMVD severity classification system used in this thesis did primarily reflect degree of left-sided cardiac dilation; possibly explaining the lack of significant differences between MMP activity and MMVD severity groups. Changes in activity could, moreover, possibly be intermittently up- or down-regulated during the disease progression (potentially counterbalanced by the TIMPs), which could create a unique MMP profile in the single dog at a given time, not reflected in group-wise comparisons including many dogs.

Down-regulation of MMP-9 has previously been suggested involved in the pathogenesis of naturally occurring MMVD in dogs (Aupperle et al., 2009c). A normalization or down-regulation of MMP activity has furthermore been described to occur in dogs with sustained volume overload after experimentally induced cardiac failure (Khan et al., 2004; Nagatomo et al., 2000). Such a down-regulation might protect against uncontrolled progressive dilation, by reducing extracellular matrix breakdown and enhance fibrosis development (Gill et al., 2006; Anne et al., 2005; Peterson et al., 2001; Blaustein et al., 1995; Dollery et al., 1995); a hypothesis strengthened by the finding of decreased activity of MMP-9 with increasing amounts of fibrosis in atrial appendages in human patients with mitral valve disease (Anne et al., 2005). An increased expression of genes encoding MMP-1 and -9, in combination with decreased expression of genes controlling synthesis of ECM components, was suggested responsible for the loss of collagen within the ECM seen in a dog model four months after MR induction (Zheng et al., 2009). As stated by the authors of that study; a time point of four months post MR induction might have been early in the time course, and fibrosis (as commonly seen in dogs with severe

MMVD of naturally acquired origin (Falk *et al.*, 2010; Falk *et al.*, 2006)) and a changed MMP profile might have ensued at a later stage of MR (Zheng *et al.*, 2009).

Active MMP-9 was in paper III shown most strongly associated with the echocardiographic variable LVIDs,, which (based on the finding in the present thesis and in previous published articles) has been shown to increase in dogs with more severe MMVD; indicating systolic dysfunction (Borgarelli et al., 2007). An increased amount of myocardial fibrosis might influence the Frank-Starling mechanism (Komamura et al., 1993), and disable an optimal transduction of contractile force generated by the myocytes in systole. This could possibly explain the association between systolic dysfunction and MMP activity. However, identification of systolic dysfunction is challenging in dogs with MMVD: The retrograde ejection of LV stroke volume, which starts already in early systole (Lord, 1974; Eckberg et al., 1973); reduces afterload, whereas the increased volume load leads to an increased preload (O'Gara et al., 2008). Depending on severity of MR, these changes lead to normal to hyperdynamic LV contraction, even in the presence of intrinsic myocardial dysfunction. Many of the commonly used echocardiographic indices for evaluating systolic function, such as the ejection phase indices (ejection fraction and shortening fraction) are, besides being dependent on intrinsic contractility, also known to be influenced by hemodynamic load and sympathetic tone, which potentially mask significant myocardial dysfunction in dogs with MR. Assessment of LV end-systolic dimension has been suggested to better reflect systolic dysfunction in the presence of MR (Borgarelli et al., 2007; Borow et al., 1980). The endsystolic dimension increases as the systolic function declines; despite increasing retrograde LV stroke volume into the low resistance LA (O'Gara et al., 2008). But if the LV contractile function is preserved, the fully compensated LV will shorten to an almost normal end-systolic dimension (Bonagura & Schober, 2009). The more recently introduced tissue doppler imaging (TDI) technique has been considered comparably independent of loading conditions, but recent studies suggest that this technique is affected by loading condition and sympathetic tone activity to a greater extent than previously expected, which might limit the additional informative value obtained from this technique, compared to when using LVIDs_{inc}(Tidholm et al., 2009). Although myocardial systolic function declines with progression of disease, the remodeling process allows the LV to retain a relatively well preserved forward cardiac pump function even in advanced MMVD (Kittleson et al., 1984): Increased pulmonary blood volume, and not decreased forward stroke volume, has been shown to be the main cause of abnormal cardiopulmonary function in dogs with MMVD (Eriksson *et al.*, 2010). This finding corresponds with the clinical observation that dogs with severe MR suffer more commonly from pulmonary congestion and edema (which cause respiratory signs) than signs caused by reduced forward cardiac output (lethargy, weakness, exercise intolerance) (Olsen *et al.*, 2010).

Activity of MMP-9 decreased with decreasing SAP in paper III. Reduced forward stroke volume due to MR (Kittleson & Brown, 2003) and LV systolic dysfunction (Borgarelli et al., 2007), might contribute in lowering SAP in dogs with more severe MMVD (as SAP is determined by cardiac output, which is the product of stroke volume and systemic vascular resistance). However, all dogs in paper II-IV had SAP within, or close to, normal reference ranges, indicating that the LV forward systolic function does not decline dramatically, and that regulatory mechanisms contribute in maintaining acceptable SAP even in the severe stage of the disease. The SAP did not differ significantly between dogs in the different MMVD severity groups in paper III, although dogs with more severe disease tended to have lower SAP than dogs with less severe disease. Such a tendency has previously been described in dogs with MMVD (Moonarmart, 2008). Significantly lower SAP (using a conservative P value; P < 0.008) in dogs with severe MMVD compared to dogs in the other MMVD severity groups were shown in paper IV. Data obtained by the HDO device used in paper IV might be more accurate and precise than data obtained by a standard oscillometry device (Schmelting et al., 2009). In addition, a faster acquisition time of the HDO device could have reduced the influence of stress on the results in paper IV. All in all; these improvements in technology might have uncovered differences in SAP between MMVD severity groups in paper IV, which could not be shown in paper III using standard oscillometry.

Circulating activities of MMP-2 and -9 have been reported to increase in human patients with acute cardiovascular diseases (Hojo *et al.*, 2001; Inokubo *et al.*, 2001), which contrasts the findings in the MMVD dogs in paper III. Hence, type, degree, and duration of extracellular stimuli in different cardiac diseases likely affect the MMP profile within the failing myocardium, as previously suggested (Spinale, 2002). Pro MMP-2 was not significantly associated with any of the investigated variables in paper III, and active MMP-2 could not be detected in the study population. Consequently, based on these results, and previously published results (Aupperle *et al.*, 2009c; Oyama & Chittur, 2006), MMP-2 is possibly not playing a central role in the progression of naturally acquired MMVD in dogs.

7.1.3 Left ventricular volume and shape

To the best of the authors' knowledge, this is the first RT3D echocardiographic study designed to investigate how the LV changes in global and regional volume and shape in response to different severities of naturally acquired mitral valve disease in any species. The cardiac remodeling process progresses slowly, but inexorably, over years as the degree of MR worsens (Lord et al., 2010). However, the results from paper IV clearly suggest that a large LV volume expansion does not occur before dogs have reached the more severe stage of the disease: Group-wise comparisons of global and regional EDV and ESV (using a conservative P value; P < 0.008) could only separate dogs with severe MMVD from other MMVD severity groups. The regurgitant fraction (the percentage of stroke volume ejected into the LA), which is regarded the major determinant factor of cardiac size and disease severity, is higher in dogs with severe disease compared to in dogs with mild to moderate disease (Kittleson & Brown, 2003); likely explaining the more pronounced LV dilation in dogs with severe MMVD in paper IV. This result is in accordance with previous studies using radiography, or circulating B-type natriuretic peptide (BNP) (which is released from the myocytes in response to increased LV myocardial stress) to investigate the progression of volume overload before onset of CHF in dogs with MMVD (Lord et al., 2010; Tarnow et al., 2009). The results reflect a more drastic increase in volume overload in dogs with close proximity to CHF. The rate of change of LV volumes in individual dogs could not be assessed in paper IV because the dogs were not followed over time.

The LV mid segment was the regional segment increasing the most in EDV and ESV with increasing disease severity in dogs included in paper IV. Potentially, LV anatomy allows more pronounced myocardial stretch in the mid segment, while the apical and basal segments are more restricted to LV expansion, due to supporting structures such as the AV annular ring in the basal segment. In addition, the LV has a smaller diameter in the apical region than in the mid-segment, and because the pressure within the LV at a given moment is the same, regardless location, wall stress is smaller in the apical than in the mid segment, according to the law of LaPlace. This, in turn, could lead to lesser tendency for dilatation of the apex. The percentage contribution of the mid segment to global EDV was only significantly higher in dogs with moderate MMVD compared to values in dogs with mild MMVD. The lack of significant differences between severe MMVD dogs and other severity groups suggests a change in LV shape in the

moderate stage; leading to increased percentage contribution of the mid segment.

Results from paper IV on LV sphericity index indicate that the LV shape changes from elliptical to more globular in response to chronic volume overload. An increase in global LV sphericity might allow myocardial adaptation to abnormal regional wall stress. However, an increase in LV sphericity might also disrupt normal mitral annular geometry; and as a result; increase the production of secondary MR (Hung *et al.*, 2004; Lapu-Bula *et al.*, 2002; Otsuji *et al.*, 1997; Kono *et al.*, 1992). Accordingly, a more globular LV shape might, to some degree, protect the myocardium from the abnormal wall stress caused by the LV volume overload, but at the same time contribute to a further increase in volume overload by stimulating secondary MR.

Assessing regional contribution to changes in LV shape, sphericity index was shown associated with decreasing percentage contribution of basal EDV to global EDV and increasing percentage of apical EDV. Rounding of the LV base with increasing MMVD severity, results in a better fit of the basal EDV into the sphere; and this change in LV shape leads to a reduced percentage contribution of the basal EDV to the global EDV. The apical EDV segment in the elliptical LV has less contact with the theoretical sphere, compared to in the spherical LV; resulting in increased percentage contribution of the apical EDV to the global EDV with increasing sphericity index. Although the mid segment was the segment increasing the most with increasing LV dilation, no association was shown between sphericity index and percentage contribution of mid EDV to the global EDV; most likely reflecting that only minor changes occur in the LV mid segmental outlining with increasing MMVD severity. These findings, in combination with the result that the long-axis length increased with increasing disease severity, suggest that although the LV changes into a more globular shape with increasing MMVD severity, the shape does not end up as an absolute sphere.

7.2 Possible clinical implications

7.2.1 Heart sounds and murmurs

The sound signal content of the murmur changes with severity of MR, which leads to increased audibility of the murmur. Hence, a typical dog (a small-breed dog, which at mature age has developed a systolic murmur of maximal audibility over the mitral valve area) previously diagnosed with MMVD by echocardiography, might be monitored by murmur assessment

until a high-audibility murmur has been diagnosed or overt signs of CHF have developed. Correct identification of severe MR would be of particular interest for clinicians when performing risk assessment or when deciding if special care and more extensive examinations are required. Because characterization of acoustic findings has been shown highly dependent on the experience of the examiner, and a considerable inter-observer variation exists (Höglund *et al.*, 2004; Pedersen *et al.*, 1999; Rajakumar *et al.*, 1999; Kinney, 1988); improvement in sound analysis tools is desirable. A majority of the sound variables investigated in paper I was dependent on MR severity, which demonstrates a diagnostic potential. The different investigated sound variables reflect different fractions of the heart sounds and murmurs; and a combination of variables was shown to optimize detection of severe MR.

The traditional auscultation technique will remain important in the near future because clinically practical tools for mathematical sound analyses procedures do not yet exist. Advances in the technical devices, making signal analysis techniques clinically applicable, are warranted: A future introduction of an innovative "intelligent stethoscope" with decision support abilities (Ahlstrom *et al.*, 2006), could hopefully offer a simple and cost-effective method for monitoring dogs with MMVD.

7.2.2 Circulating cardiac biomarkers

Both cardiac and extracardiac diseases have the potential to induce myocyte injury, and due to the non-specific origin of myocardial damage; assessment of circulating concentrations of cTnI is unlikely to perform well when establishing the diagnosis of MMVD. Although the circulating cTnI concentration increased with increasing MMVD severity, a considerable overlap existed between the different severity groups; hence, cTnI concentration alone cannot effectively be used for MMVD severity assessment. Establishing prognosis in a single dog with MMVD is difficult, and myocardial changes reflected by high sensitivity cTnI analyses might provide prognostic information. High sensitivity assays have shown previously immeasurable troponin concentrations in humans with cardiovascular diseases, to be highly predictive of future cardiac events (Eggers et al., 2009; Schulz et al., 2007). Increased cTnI concentrations have furthermore been shown linked to a worse outcome in dogs with cardiomyopathy (Oyama & Sisson, 2004). The studies in papers II and III were not designed to evaluate the prognostic value of the circulating biomarkers, and future longitudinal studies are needed for that purpose. Different biomarkers can provide information about different aspects of cardiovascular diseases, and a combination of troponin analyses with analyses of other biomarkers, such as BNP, has the potential to further improve the prognostic assessment of the individual cardiac patient (Latini *et al.*, 2007; Jernberg *et al.*, 2002). However, effect of age on cTnI needs consideration when assessing cTnI in canine cardiac patients.

Concentration of CRP has previously been reported associated with severity of CHF (Pye *et al.*, 1990), and shown capable of predicting acute cardiovascular events in human patients (Sakkinen *et al.*, 2002). The lack of association between CRP concentration and MMVD severity in the examined dogs in paper II indicates a lack of diagnostic value of CRP analyses in MMVD dogs. However, a potential value of CRP (preferably when used in combination with other biomarkers) for predicting outcome in MMVD dogs, cannot be excluded.

Whereas established cardiac biomarkers; such as the natriuretic peptides, primarily reflect ventricular wall stretch (and thereby indirectly volume overload), MMP-9 was shown associated with variables reflecting systolic function in paper III. Hence, MMP-9 might provide additional information to other biomarkers used when trying to explore the complex cardiac remodeling process in MMVD dogs. However, the significant associations found between MMP-9 activity and some of the investigated variables were rather weak and circulating activities of MMP-2 and -9 cannot be regarded as valuable diagnostic biomarkers in clinical practice based on the results from paper III. Identification of the specific portfolio of MMPs and TIMPs expressed within the failing myocardium could potentially improve future treatment strategies for individuals with chronic cardiac diseases; aiming to prevent or slow down the disease progression, instead of only improving clinical signs at the end of the remodeling process.

7.2.3 Left ventricular volume and shape

The results from the RT3D LV volume assessment of dogs in paper IV showed prominent LV volume expansions only in dogs with more severe MMVD. Left heart chamber enlargement has been characterized by a slow phase of steadily progressing MMVD until about 6 to 12 months before onset of CHF, when rate of change of enlargement is fast (Lord *et al.*, 2010). Routinely performed volume assessment could potentially improve prediction of outcome for individual dogs; allowing risk-detection of forthcoming CHF at an earlier stage. The study design of paper IV did not include longitudinal follow-up studies; hence, the value of RT3D measurements in predicting outcome could not be evaluated. In order to use RT3D LV volumes in clinical practice, normal reference values need to

be established. However, although clinical use of modern 3D systems is boosted by advances in computer technology; reducing acquisition and post processing time, a shortcoming of the RT3D technique still lies in the fact that it is more time consuming compared to traditional echocardiographic techniques.

8 Conclusions

- Linear and nonlinear analyses of cardiac sounds can be used to assess MR severity, as shown by the associations found between many of the investigated sound variables and MR severity: More severe MR produces a murmur of "harsher" quality, longer duration, and with more complexity in the signal. The energy of S1 was not associated with MR severity (assessed by echocardiography), whereas the energy of S2 decreased with increasing MR severity.
- Circulating cTnI concentration increased with increasing disease severity, however, the effect of age on cTnI needs consideration when evaluating chronic cardiac remodeling in dogs using a high sensitivity cTnI assay. Circulating CRP concentration was not associated with disease severity.
- The MMP-2 and -9 activities were not associated with MMVD severity groups (which were mainly based on severity of volume overload). However, MMP-9 activity decreased with worsening systolic function. Based on the findings from this thesis, MMP-2 might play a minor role in MMVD in dogs.
- Chronic volume overload in dogs with MMVD changes LV geometry. The RT3D examinations showed prominent LV volume expansions only in dogs with more severe MMVD. The mid LV segment contributed the most to the global volume increase. The LV shape changed from elliptical to more globular in response to increasing volume overload, with the basal and apical segments contributing the most to the increase in sphericity.

9 Implications for future research

- Further studies, investigating the potential of signal analysis technique for analysis of cardiac sounds in dogs with various cardiac diseases, are warranted.
- High sensitivity cTnI assays, capable of detecting subtle myocardial changes, could be valuable for future prospective studies. Preferably, cTnI could be combined with other biomarkers in order to better predict long-term outcome.
- Further investigations, including other markers of inflammation than CRP, could be conducted in order to further evaluate a potential role of inflammation in the pathogenesis of MMVD and CHF development.
- An important future direction is to characterize the specific portfolio of MMPs and TIMPs in the myocardium, and furthermore, investigate their interactions in dogs with MMVD.
- Assessment of LV volume and shape could potentially allow early detection of dogs at risk for rapid progression into congestive heart failure. Longitudinal follow-up studies are needed to investigate whether RT3D echocardiography could improve prediction of outcome in dogs with MMVD compared to conventional echocardiography.

10 Populärvetenskaplig sammanfattning

Kronisk hjärtklaffsdegeneration är den vanligaste hjärtsjukdomen hos hund, och den drabbar framför allt hundar av små till medelstora raser från medelåldern och uppåt i ålder. Sjukdomen leder till att hjärtklaffen mellan vänster förmak och kammare (mitralisklaffen) förändras i struktur och får ett onormalt rörelsemönster. Sjukdomen liknar mitralisprolaps-syndromet hos människor. Eftersom klaffen inte längre kan sluta tätt, börjar blod läcka tillbaka upp i förmaket när hjärtat kontraheras, istället för att enbart pumpas ut i stora kroppspulsådern. Turbulens i blodet uppstår i samband med klaffläckaget, vilket ger upphov till ett blåsljud som kan höras när man lyssnar på hjärtat med stetoskop. Graden av turbulens styrs både av graden av klaffläckage och av vänster kammares funktionsduglighet. För att kompensera klaffläckaget ökar volymen i vänster förmak och kammare, muskelmassa. Hundar liksom vänster kammares med kronisk hjärtklaffsdegeneration kan ha sjukdomen under flera år utan att visa sjukdomstecken. Hos vissa hundar blir dock återflödet till förmaket så kraftigt att hjärtat inte längre klarar att kompensera för otätheten, och hundarna drabbas av hjärtsvikt.

Syftet med avhandlingen var att beskriva hur vänster kammare förändras hos hundar med olika långt framskriden hjärtklaffsdegeneration genom att använda digital analys av blåsljud och hjärtljud, specifika markörer som kan mätas i blodet och modern ultraljudsteknik.

I avhandlingen användes för första gången hos något djurslag digital förändringar i blåsljudets och signalanalysteknik för att undersöka hjärttonernas karaktär vid olika långt framskriden kronisk hjärtklaffsdegeneration. Tekniken möjliggör även analys av ljudsignaler som inte kan uppfattas av människans hörsel. Resultaten visade att hundar med kraftigt klaffläckage hade blåsljud med högre frekvensinnehåll, duration och komplexitet, jämfört med hundar med mildare klaffläckage. Även hjärttonernas energiinnehåll påverkades av graden av klaffläckage.

Förändringar i hjärtats volym och vikt bidrar till att olika substanser frisätts ut i blodet. Dessa substanser kan användas som biologiska markörer (biomarkörer) för hjärtsjukdom och fungerar som ett komplement till mera traditionella undersökningsmetoder. Troponiner är proteiner som reglerar muskelcellernas kontraktionsförmåga. Cardiac troponin-I (cTnI), som endast finns i hjärtmuskulatur, läcker i samband med skador på hjärtmuskelceller ut i blodet. En ny, känslig analysmetod, med kapacitet att upptäcka mycket små cTnI nivåer i blodet användes för att undersöka hjärtmuskelskada hos hundar med olika grad av kronisk hjärtklaffsdegeneration. Resultaten visade att cTnI ökade med ökad sjukdomsgrad och en ökning förelåg redan hos hundar med lindrig sjukdom. Sambandet mellan inflammationsmarkören Creaktivt protein (CRP) och kronisk klaffdegenerationsgrad undersöktes också, och resultaten tyder på att CRP inte är kopplad till grad av kronisk hjärtklaffsdegeneration hos hund. Vid kronisk hjärtklaffsdegeneration uppstår förändringar i hjärtats stödjevävnad, vilket kan påverka hjärtats funktion. Matrix metalloproteinaser (MMP) är enzymer som anses spela en viktig roll stödjevävnad hos reglera hjärtats hundar med för att kronisk hjärtklaffsdegeneration. Resultat från avhandlingen tyder på att det finns ett samband mellan MMP nivå i blodet och hjärtats sammandragningsförmåga hos hundar med kronisk hjärtklaffsdegeneration.

Modern tredimensionell (3D) ultraljudsteknik kan förbättra utvärderingen av vänster kammares volym och form. För första gång har modern "real-time" tredimensionell (RT3D) ultraljudsteknik använts för att utvärdera kammarens geometri hos hundar med olika långt framskriden hjärtklaffsdegeneration. Redan på ett tidigt stadium av kronisk sjukdomsutvecklingen kunde en liten ökning av vänster kammares volym ses, men först vid långt gången sjukdom kunde en påtaglig volymsökning påvisas. Vänster kammare uppvisade dessutom en mera rundad form hos hundar med långt gången sjukdom.

I avhandlingen presenteras nya resultat som ökar kunskapen om de komplexa förändringar som sker i hjärtat hos hundar med kronisk hjärtklaffsdegeneration; kunskap som förhoppningsvis kommer att bidra till förbättrat omhändertagande av hjärtsjuka hundar i framtiden.

11 References

- Ahlstrom, C., Hult, P., Rask, P., Karlsson, J., Nylander, E., Dahlstrom, U. & Ask, P. (2006). Feature extraction for systolic heart murmur classification. *Ann Biomed Eng* 34(11), 1666-77.
- Ahlstrom, C. (2008). Nonlinear Phonocardiographic Signal Processing. PhD thesis. Linköping University. Linköping.
- Ahmed, M.I., McGiffin, D.C., O'Rourke, R.A. & Dell'Italia, L.J. (2009). Mitral regurgitation. *Curr Probl Cardiol* 34(3), 93-136.
- Anne, W., Willems, R., Roskams, T., Sergeant, P., Herijgers, P., Holemans, P., Ector, H. & Heidbuchel, H. (2005). Matrix metalloproteinases and atrial remodeling in patients with mitral valve disease and atrial fibrillation. *Cardiovasc Res* 67(4), 655-66.
- Anversa, P., Ricci, R. & Olivetti, G. (1986). Quantitative structural analysis of the myocardium during physiologic growth and induced cardiac hypertrophy: a review. *J Am Coll Cardiol* 7(5), 1140-9.
- Arndt, J.W., Reynolds, C.A., Singletary, G.E., Connolly, J.M., Levy, R.J. & Oyama, M.A. (2009). Serum serotonin concentrations in dogs with degenerative mitral valve disease. J Vet Intern Med 23(6), 1208-13.
- Aupperle, H., März, I., Thielebein, J., Kiefer, B., Kappe, A. & Schoon, H.A. (2009a). Immunohistochemical characterization of the extracellular matrix in normal mitral valves and in chronic valve disease (endocardiosis) in dogs. *Res Vet Sci* 87(2), 277-83.
- Aupperle, H., Thielebein, J., Kiefer, B., März, I., Dinges, G. & Schoon, H.A. (2009b). An immunohistochemical study of the role of matrix metalloproteinases and their tissue inhibitors in chronic mitral valvular disease (valvular endocardiosis) in dogs. *Vet J* 180(1), 88-94.
- Aupperle, H., Thielebein, J., Kiefer, B., März, I., Dinges, G., Schoon, H.A. & Schubert, A. (2009c). Expression of genes encoding matrix metalloproteinases (MMPs) and their tissue inhibitors (TIMPs) in normal and diseased canine mitral valves. J Comp Pathol 140(4), 271-7.

- Barlucchi, L., Leri, A., Dostal, D.E., Fiordaliso, F., Tada, H., Hintze, T.H., Kajstura, J., Nadal-Ginard, B. & Anversa, P. (2001). Canine ventricular myocytes possess a renin-angiotensin system that is upregulated with heart failure. *Circ Res* 88(3), 298-304.
- Beardow, A. & Buchanan, J. (1993). Chronic mitral valve disease in Cavalier King Charles spaniels: 95 cases (1987-1991). J Am Vet Med Assoc 203(7), 1023-9.
- Bélanger, M.C. (2005). Echocardiography. In: Ettinger, S.J., et al. (Eds.) Textbook of veterinary internal medicine. St Louis, Missouri: Elsevier Saunders.1. pp. 311-326.
- Black, A., French, A.T., Dukes-McEwan, J. & Corcoran, B.M. (2005). Ultrastructural morphologic evaluation of the phenotype of valvular interstitial cells in dogs with myxomatous degeneration of the mitral valve. *Am J Vet Res* 66(8), 1408-14.
- Blaine, D. (1817). Canine pathology or a full description of the diseases of dogs with their causes, symptoms, and mode of cure. London: T Boosey
- Bland, M. (1995). An introduction to medical statistics. Oxford: Oxford University Press. pp. 322-323.
- Blaustein, A.S., Hoit, B.D., Wexler, L.F., Ashraf, M., Ramrakhyani, K., Matoba, R., Gabel, M. & Millard, R.W. (1995). Characteristics of chronic left ventricular dysfunction induced by coronary embolization in a canine model. *Am J Cardiovasc Pathol* 5(1), 32-48.
- Bonagura, J.D. & Schober, K.E. (2009). Can ventricular function be assessed by echocardiography in chronic canine mitral valve disease? *J Small Anim Pract* 50 Suppl 1, 12-24.
- Borgarelli, M. (2004). *Mitral valve insufficiency in large breed dogs*. PhD Thesis. Universita' degli studi di Torino. Torino.
- Borgarelli, M., Tarducci, A., Zanatta, R. & Häggström, J. (2007). Decreased systolic function and inadequate hypertrophy in large and small breed dogs with chronic mitral valve insufficiency. *J Vet Intern Med* 21(1), 61-7.
- Borow, K.M., Green, L.H., Mann, T., Sloss, L.J., Braunwald, E., Collins, J.J., Cohn, L. & Grossman, W. (1980). End-systolic volume as a predictor of postoperative left ventricular performance in volume overload from valvular regurgitation. *Am J Med* 68(5), 655-63.
- Bozkurt, B., Kribbs, S.B., Clubb, F.J., Jr., Michael, L.H., Didenko, V.V., Hornsby, P.J., Seta, Y., Oral, H., Spinale, F.G. & Mann, D.L. (1998). Pathophysiologically relevant concentrations of tumor necrosis factor-alpha promote progressive left ventricular dysfunction and remodeling in rats. *Circulation* 97(14), 1382-91.
- Bristow, M.R., Ginsburg, R., Minobe, W., Cubicciotti, R.S., Sageman, W.S., Lurie, K., Billingham, M.E., Harrison, D.C. & Stinson, E.B. (1982). Decreased catecholamine sensitivity and beta-adrenergicreceptor density in failing human hearts. *New Engl J Med* 307(4), 205-11.
- Bristow, M.R., Ginsburg, R., Umans, V., Fowler, M., Minobe, W., Rasmussen, R., Zera, P., Menlove, R., Shah, P., Jamieson, S. & et al. (1986). Beta 1- and beta 2-adrenergic-receptor subpopulations in nonfailing and failing human ventricular myocardium: coupling of both receptor subtypes to muscle contraction and selective beta 1receptor down-regulation in heart failure. *Circ Res* 59(3), 297-309.
- Brolin, I. (1967). The mitral orifice. Acta Radiol Diagn (Stockh) 6(3), 273-95.
- Brown, S., Atkins, C., Bagley, R., Carr, A., Cowgill, L., Davidson, M., Egner, B., Elliott, J., Henik, R., Labato, M., Littman, M., Polzin, D., Ross, L., Snyder, P. & Stepien, R. (2007). Guidelines for the identification, evaluation, and management of systemic hypertension in dogs and cats. *J Vet Intern Med* 21(3), 542-58.
- Buchanan, J. (1977). Chronic valvular disease (endocardiosis) in dogs. Adv Vet Sci Comp Med 21, 75-106.
- Carabello, B.A. (2000). The pathophysiology of mitral regurgitation. J Heart Valve Dis 9(5), 600-8.
- Carabello, B.A. (2002). Concentric versus eccentric remodeling. J Card Fail 8(6 Suppl), S258-63.
- Carabello, B.A., Nakano, K., Corin, W., Biederman, R. & Spann, J.F., Jr. (1989). Left ventricular function in experimental volume overload hypertrophy. *Am J Physiol* 256(4 Pt 2), H974-81.
- Cerqueira, M.D., Weissman, N.J., Dilsizian, V., Jacobs, A.K., Kaul, S., Laskey, W.K., Pennell, D.J., Rumberger, J.A., Ryan, T. & Verani, M.S. (2002). Standardized myocardial segmentation and nomenclature for tomographic imaging of the heart: a statement for healthcare professionals from the Cardiac Imaging Committee of the Council on Clinical Cardiology of the American Heart Association. *Circulation* 105(4), 539-42.
- Chiechi, M.A., Lees, W.M. & Thompson, R. (1956). Functional anatomy of the normal mitral valve. *J Thorac Surg* 32(3), 378-98.
- Cohn, J.N. (1995). Critical review of heart failure: the role of left ventricular remodeling in the therapeutic response. *Clin Cardiol* 18(9 Suppl 4), IV4-12.
- Cohn, J.N., Ferrari, R. & Sharpe, N. (2000). Cardiac remodeling--concepts and clinical implications: a consensus paper from an international forum on cardiac remodeling. Behalf of an International Forum on Cardiac Remodeling. J Am Coll Cardiol 35(3), 569-82.
- Coker, M.L., Doscher, M.A., Thomas, C.V., Galis, Z.S. & Spinale, F.G. (1999). Matrix metalloproteinase synthesis and expression in isolated LV myocyte preparations. *Am J Physiol* 277(2 Pt 2), H777-87.
- Corcoran, B.M., Black, A., Anderson, H., McEwan, J.D., French, A., Smith, P. & Devine, C. (2004). Identification of surface morphologic changes in the mitral valve leaflets and chordae tendineae of dogs with myxomatous degeneration. *Am J Vet Res* 65(2), 198-206.

- Cornell, C., Kittleson, M., Della Torre, P., Haggstrom, J., Lombard, C., Pedersen, H., Vollmar, A. & Wey, A. (2004). Allometric scaling of M-mode cardiac measurements in normal adult dogs. *J Vet Intern Med* 18(3), 311-21.
- Darke, P. (1987). Valvular incompetence in Cavalier King Charles spaniels. *Vet Rec* 120(15), 365-6.
- Das, K. & Tashjihan, R. (1965). Chronic mitral valve disease in the dog. Vet Med / Small Anim Clin 60, 1209-1215.
- Davis, P.K. & Kinmonth, J.B. (1963). The Movements of the Annulus of the Mitral Valve. J Cardiovasc Surg (Torino) 4, 427-31.
- de Laforcade, A.M., Freeman, L.M. & Rush, J.E. (2003). Serum nitrate and nitrite in dogs with spontaneous cardiac disease. *J Vet Intern Med* 17(3), 315-8.
- Dell'italia, L.J., Balcells, E., Meng, Q.C., Su, X., Schultz, D., Bishop, S.P., Machida, N., Straeter-Knowlen, I.M., Hankes, G.H., Dillon, R., Cartee, R.E. & Oparil, S. (1997). Volume-overload cardiac hypertrophy is unaffected by ACE inhibitor treatment in dogs. *Am J Physiol* 273(2 Pt 2), H961-70.
- Dell'Italia, L.J., Meng, Q.C., Balcells, E., Straeter-Knowlen, I.M., Hankes, G.H., Dillon, R., Cartee, R.E., Orr, R., Bishop, S.P., Oparil, S. & et al. (1995). Increased ACE and chymase-like activity in cardiac tissue of dogs with chronic mitral regurgitation. *Am J Physiol* 269(6 Pt 2), H2065-73.
- Detweiler, D. & Patterson, D. (1965). The prevalence and types of cardiovascular disease in dogs. *Ann N Y Acad Sci* 127(1), 481-516.
- Detweiler, D.K. (1989). Spontaneous and induced arterial disease in the dog: pathology and pathogenesis. *Toxicol Pathol* 17(1 Pt 2), 94-108.
- Detweiler, D.K., Luginbuhl, H., Buchanan, J.W. & Patterson, D.F. (1968). The natural history of acquired cardiac disability of the dog. *Ann N Y Acad Sci* 147(8), 318-29.
- Di Donato, M., Dabic, P., Castelvecchio, S., Santambrogio, C., Brankovic, J., Collarini, L., Joussef, T., Frigiola, A., Buckberg, G. & Menicanti, L. (2006). Left ventricular geometry in normal and post-anterior myocardial infarction patients: sphericity index and 'new' conicity index comparisons. *Eur J Cardiothorac Surg* 29 Suppl 1, S225-30.
- Dollery, C.M., McEwan, J.R. & Henney, A.M. (1995). Matrix metalloproteinases and cardiovascular disease. *Circ Res* 77(5), 863-8.
- Dreger, S.A., Taylor, P.M., Allen, S.P. & Yacoub, M.H. (2002). Profile and localization of matrix metalloproteinases (MMPs) and their tissue inhibitors (TIMPs) in human heart valves. *J Heart Valve Dis* 11(6), 875-80; discussion 880.
- Durand, L. & Pibarot, P. (1995). Digital signal processing of the phonocardiogram: review of the most recent advancements. *Crit Rev Biomed Eng* 23(3-4), 163-219.

- Eckberg, D., Gault, J., Bouchard, R., Karliner, J. & Ross, J.J. (1973). Mechanics of left ventricular contraction in chronic severe mitral regurgitation. *Circulation* 47(6), 1252-9.
- Eckersall, P.D. & Conner, J.G. (1988). Bovine and canine acute phase proteins. *Vet Res Commun* 12(2-3), 169-78.
- Egenvall, A., Bonnett, B. & Haggstrom, J. (2006). Heart disease as a cause of death in insured Swedish dogs younger than 10 years of age. *J Vet Intern Med* 20(4), 894–903.
- Eggers, K.M., Jaffe, A.S., Lind, L., Venge, P. & Lindahl, B. (2009). Value of cardiac troponin I cutoff concentrations below the 99th percentile for clinical decision-making. *Clin Chem* 55(1), 85-92.
- Eriksson, A., Hansson, K., Häggström, J., Jarvinen, A.K. & Lord, P. (2010). Pulmonary blood volume in mitral regurgitation in cavalier king charles spaniels. J Vet Intern Med 24(6), 1393-9.
- Falk, T., Jönsson, L., Olsen, L.H. & Pedersen, H.D. (2006). Arteriosclerotic changes in the myocardium, lung, and kidney in dogs with chronic congestive heart failure and myxomatous mitral valve disease. *Cardiovasc Pathol* 15(4), 185-93.
- Falk, T., Jönsson, L., Olsen, L.H., Tarnow, I. & Pedersen, H.D. (2010). Associations between cardiac pathology and clinical, echocardiographic and electrocardiographic findings in dogs with chronic congestive heart failure. *Vet J* 185(1), 68-74.
- Feng, Y.J., Chen, C., Fallon, J.T., Lai, T., Chen, L., Knibbs, D.R., Waters, D.D. & Wu, A.H. (1998). Comparison of cardiac troponin I, creatine kinase-MB, and myoglobin for detection of acute ischemic myocardial injury in a swine model. *Am J Clin Pathol* 110(1), 70-7.
- Ferrari, R., Agnoletti, L., Comini, L., Gaia, G., Bachetti, T., Cargnoni, A., Ceconi, C., Curello, S. & Visioli, O. (1998). Oxidative stress during myocardial ischaemia and heart failure. *Eur Heart J* 19 Suppl B, B2-11.
- Fischer, R.A. (1936). The use of multiple measurements in taxonomic problems. *Ann Eugenics* 7(179-188), 179.
- Gill, R.M., Jones, B.D., Corbly, A.K., Wang, J., Braz, J.C., Sandusky, G.E. & Shen, W. (2006). Cardiac diastolic dysfunction in conscious dogs with heart failure induced by chronic coronary microembolization. *Am J Physiol Heart Circ Physiol* 291(6), H3154-8.
- Gompf, R. (1988). The clinical approach to heart disease: history and physical examination. In: Fox, P.R. (Ed.) *Canine and feline cardiology*. New York, NY: Churchill Livingstone Inc,. pp. 29-42.
- Gould, L., Ettinger, S. & Lyon, A. (1968). Intensity of the first heart sound and arterial pulse in mitral insufficiency. *Dis Chest* 53(5), 545-50.
- Grossman, W., Jones, D. & McLaurin, L.P. (1975). Wall stress and patterns of hypertrophy in the human left ventricle. *J Clin Invest* 56(1), 56-64.

- Hadian, M., Corcoran, B.M. & Bradshaw, J.P. (2010). Molecular changes in fibrillar collagen in myxomatous mitral valve disease. *Cardiovasc Pathol* 19(5), e141-8.
- Hadian, M., Corcoran, B.M., Han, R.I., Grossmann, J.G. & Bradshaw, J.P. (2007). Collagen organization in canine myxomatous mitral valve disease: an x-ray diffraction study. *Biophys J* 93(7), 2472-6.
- Häggström, J., Hansson, K., Kvart, C., Karlberg, B., Vuolteenaho, O. & Olsson, K. (1997). Effects of naturally acquired decompensated mitral valve regurgitation on the renin-angiotensin-aldosterone system and atrial natriuretic peptide concentration in dogs. *Am J Vet Res* 58(1), 77-82.
- Häggström, J., Hansson, K., Kvart, C. & Swenson, L. (1992). Chronic valvular disease in the Cavalier King Charles spaniel in Sweden. *Vet record* 131(24), 549-53.
- Häggström, J., Kvart, C. & Hansson, K. (1995). Heart sounds and murmurs: changes related to severity of chronic valvular disease in the Cavalier King Charles spaniel. *J Vet Intern Med* 9(2), 75-85.
- Han, R.I., Black, A., Culshaw, G., French, A.T. & Corcoran, B.M. (2010). Structural and cellular changes in canine myxomatous mitral valve disease: an image analysis study. *J Heart Valve Dis* 19(1), 60-70.
- Hansson, K., Häggström, J., Kvart, C. & Lord, P. (2002). Left atrial to aortic root indices using two-dimensional and M-mode echocardiography in Cavalier King Charles spaniels with and without left atrial enlargement. *Vet Radiol Ultrasound* 43(6), 568-75.
- Höglund, K. (2007). Systolic ejection murmurs and the left ventricular outflow tract in boxer dogs; Physiology and clinical evaluation. PhD Thesis. Swedish university of agricultural sciences. Uppsala.
- Höglund, K., French, A., Dukes-McEwan, J., Haggstrom, J., Smith, P., Corcoran, B. & Kvart, C. (2004). Low intensity heart murmurs in Boxer dogs: inter-observer variation and effects of stress testing. J Small Anim Pract 45(4), 178-85.
- Hojo, Y., Ikeda, U., Ueno, S., Arakawa, H. & Shimada, K. (2001). Expression of matrix metalloproteinases in patients with acute myocardial infarction. *Jap Circ J* 65(2), 71-5.
- Hung, J., Papakostas, L., Tahta, S.A., Hardy, B.G., Bollen, B.A., Duran, C.M. & Levine, R.A. (2004). Mechanism of recurrent ischemic mitral regurgitation after annuloplasty: continued LV remodeling as a moving target. *Circulation* 110(11 Suppl 1), II85-90.
- Hutchinson, K.R., Stewart, J.A., Jr. & Lucchesi, P.A. (2010). Extracellular matrix remodeling during the progression of volume overload-induced heart failure. *J Mol Cell Cardiol* 48(3), 564-9.
- Inokubo, Y., Hanada, H., Ishizaka, H., Fukushi, T., Kamada, T. & Okumura, K. (2001). Plasma levels of matrix metalloproteinase-9 and tissue inhibitor of metalloproteinase-1 are increased in the

coronary circulation in patients with acute coronary syndrome. *Am Heart J* 141(2), 211-7.

- Jacobs, L.D., Salgo, I.S., Goonewardena, S., Weinert, L., Coon, P., Bardo, D., Gerard, O., Allain, P., Zamorano, J.L., de Isla, L.P., Mor-Avi, V. & Lang, R.M. (2006). Rapid online quantification of left ventricular volume from real-time three-dimensional echocardiographic data. *Eur Heart J* 27(4), 460-8.
- Jaffe, A.S., Landt, Y., Parvin, C.A., Abendschein, D.R., Geltman, E.M. & Ladenson, J.H. (1996). Comparative sensitivity of cardiac troponin I and lactate dehydrogenase isoenzymes for diagnosing acute myocardial infarction. *Clinical chemistry* 42(11), 1770-6.
- Jernberg, T., Stridsberg, M., Venge, P. & Lindahl, B. (2002). N-terminal pro brain natriuretic peptide on admission for early risk stratification of patients with chest pain and no ST-segment elevation. *J Am Coll Cardiol* 40(3), 437-45.
- Jönsson, L. (1972). Coronary arterial lesions and myocardial infarcts in the dog. A pathologic and microangiographic study. *Acta Vet Scand Suppl* 38, 1-80.
- Kato, S., Spinale, F.G., Tanaka, R., Johnson, W., Cooper, G.t. & Zile, M.R. (1995). Inhibition of collagen cross-linking: effects on fibrillar collagen and ventricular diastolic function. *Am J Physiol* 269(3 Pt 2), H863-8.
- Katz, A.M. (1990). Cardiomyopathy of overload. A major determinant of prognosis in congestive heart failure. N Engl J Med 322(2), 100-10.
- Katz, A.M. (1995). The cardiomyopathy of overload: an unnatural growth response. *Eur Heart J* 16 Suppl O, 110-4.
- Katz, A.M. (2001). Muscle contraction. In: Katz A. M. ed, *Physiology of the heart* 3rd ed, 135-137.
- Khan, A., Moe, G.W., Nili, N., Rezaei, E., Eskandarian, M., Butany, J. & Strauss, B.H. (2004). The cardiac atria are chambers of active remodeling and dynamic collagen turnover during evolving heart failure. *J Am Coll Cardiol* 43(1), 68-76.
- Kinney, E. (1988). Causes of false-negative auscultation of regurgitant lesions: a Doppler echocardiographic study of 294 patients. J Gen Intern Med 3(5), 429-34.
- Kittleson, M. & Brown, W. (2003). Regurgitant fraction measured by using the proximal isovelocity surface area method in dogs with chronic myxomatous mitral valve disease. *J Vet Intern Med* 17(1), 84-8.
- Kittleson, M., Eyster, G., Knowlen, G., Bari Olivier, N. & Anderson, L. (1984). Myocardial function in small dogs with chronic mitral regurgitation and severe congestive heart failure. J Am Vet Med Assoc 184(4), 455-9.
- Kjelgaard-Hansen, M., Kristensen, A.T. & Jensen, A.L. (2003). Evaluation of a commercially available enzyme-linked immunosorbent assay

(ELISA) for the determination of C-reactive protein in canine serum. J Vet Med A Physiol Pathol Clin Med 50(3), 164-8.

- Klima, M., Burns, T.R. & Chopra, A. (1990). Myocardial fibrosis in the elderly. *Arch Pathol Lab Med* 114(9), 938-42.
- Kogure, K. (1980). Pathology of chronic mitral valvular disease in the dog. Nippon Juigaku Zasshi 42(3), 323-35.
- Komamura, K., Shannon, R.P., Ihara, T., Shen, Y.T., Mirsky, I., Bishop, S.P. & Vatner, S.F. (1993). Exhaustion of Frank-Starling mechanism in conscious dogs with heart failure. *Am J Physiol* 265(4 Pt 2), H1119-31.
- Kono, T., Sabbah, H.N., Rosman, H., Alam, M., Jafri, S. & Goldstein, S. (1992). Left ventricular shape is the primary determinant of functional mitral regurgitation in heart failure. J Am Coll Cardiol 20(7), 1594-8.
- Kupferwasser, I., Mohr-Kahaly, S., Stahr, P., Rupprecht, H.J., Nixdorff, U., Fenster, M., Voigtlander, T., Erbel, R. & Meyer, J. (1997). Transthoracic three-dimensional echocardiographic volumetry of distorted left ventricles using rotational scanning. J Am Soc Echocardiogr 10(8), 840-52.
- Lakhan, S.E. & Harle, L. (2008). Cardiac fibrosis in the elderly, normotensive athlete: case report and review of the literature. *Diagn Path* 3, 12.
- Lang, R.M., Bierig, M., Devereux, R.B., Flachskampf, F.A., Foster, E., Pellikka, P.A., Picard, M.H., Roman, M.J., Seward, J., Shanewise, J., Solomon, S., Spencer, K.T., St John Sutton, M. & Stewart, W. (2006a). Recommendations for chamber quantification. *Eur J Echocardiogr* 7(2), 79-108.
- Lang, R.M., Bierig, M., Devereux, R.B., Flachskampf, F.A., Foster, E., Pellikka, P.A., Picard, M.H., Roman, M.J., Seward, J., Shanewise, J.S., Solomon, S.D., Spencer, K.T., Sutton, M.S. & Stewart, W.J. (2005). Recommendations for chamber quantification: a report from the American Society of Echocardiography's Guidelines and Standards Committee and the Chamber Quantification Writing Group, developed in conjunction with the European Association of Echocardiography, a branch of the European Society of Cardiology. J Am Soc Echocardiogr 18(12), 1440-63.
- Lang, R.M., Mor-Avi, V., Sugeng, L., Nieman, P.S. & Sahn, D.J. (2006b). Three-dimensional echocardiography: the benefits of the additional dimension. J Am Coll Cardiol 48(10), 2053-69.
- Lapu-Bula, R., Robert, A., Van Craeynest, D., D'Hondt, A.M., Gerber, B.L., Pasquet, A., Melin, J.A., De Kock, M. & Vanoverschelde, J.L. (2002). Contribution of exercise-induced mitral regurgitation to exercise stroke volume and exercise capacity in patients with left ventricular systolic dysfunction. *Circulation* 106(11), 1342-8.

- Latini, R., Masson, S., Anand, I.S., Missov, E., Carlson, M., Vago, T., Angelici, L., Barlera, S., Parrinello, G., Maggioni, A.P., Tognoni, G. & Cohn, J.N. (2007). Prognostic value of very low plasma concentrations of troponin T in patients with stable chronic heart failure. *Circulation* 116(11), 1242-9.
- LeGrice, I.J., Smaill, B.H., Chai, L.Z., Edgar, S.G., Gavin, J.B. & Hunter, P.J. (1995). Laminar structure of the heart: ventricular myocyte arrangement and connective tissue architecture in the dog. *Am J Physiol* 269(2 Pt 2), H571-82.
- Lewis, T., Swift, S., Woolliams, J.A. & Blott, S. (2011). Heritability of premature mitral valve disease in Cavalier King Charles spaniels. *Vet* J 188(1), 73-6.
- Levy, M.J. & Edwards, J.E. (1962). Anatomy of mitral insufficiency. Prog Cardiovasc Dis 5, 119-44.
- Linde, A., Summerfield, N.J., Sleeper, M.M., Wright, F.B., Clifford, C.A., Melgarejo, T. & Knight, D.H. (2006). Pilot study on cardiac troponin I levels in dogs with pericardial effusion. J Vet Cardiol 8(1), 19-23.
- Little, R.C. (1979). The mechanism of closure of the mitral valve: a continuing controversy. *Circulation* 59(4), 615-8.
- Lord, P., Hansson, K., Kvart, C. & Häggström, J. (2010). Rate of change of heart size before congestive heart failure in dogs with mitral regurgitation. J Small Anim Pract 51(4), 210-8.
- Lord, P.F. (1974). Left ventricular volumes of diseased canine heart: congestive cardiomyopathy and volume overload (patent ductus arteriosus and primary mitral valvular insufficiency). *Am J Vet Res* 35, 493-501.
- Loukopoulos, P., Mungall, B.A., Straw, R.C., Thornton, J.R. & Robinson, W.F. (2003). Matrix metalloproteinase-2 and -9 involvement in canine tumors. *Vet Path* 40(4), 382-94.
- Lu, X., Xie, M., Tomberlin, D., Klas, B., Nadvoretskiy, V., Ayres, N., Towbin, J. & Ge, S. (2008). How accurately, reproducibly, and efficiently can we measure left ventricular indices using M-mode, 2dimensional, and 3-dimensional echocardiography in children? Am Heart J 155(5), 946-53.
- Mannaerts, H.F., van der Heide, J.A., Kamp, O., Stoel, M.G., Twisk, J. & Visser, C.A. (2004). Early identification of left ventricular remodelling after myocardial infarction, assessed by transthoracic 3D echocardiography. *Eur Heart J* 25(8), 680-7.
- Mihalatos, D.G., Joseph, S., Gopal, A., Bercow, N., Toole, R., Passick, M., Grimson, R., Norales, A. & Reichek, N. (2007). Mitral annular remodeling with varying degrees and mechanisms of chronic mitral regurgitation. J Am Soc Echocard 20(4), 397-404.

- Mischke, R., Waterston, M. & Eckersall, P.D. (2007). Changes in Creactive protein and haptoglobin in dogs with lymphatic neoplasia. *Vet J* 174(1), 188-92.
- Monaghan, M.J. (2006). Role of real time 3D echocardiography in evaluating the left ventricle. *Heart* 92(1), 131-6.
- Moonarmart, W. (2008). Studies on the Natural History and progression of Aquired Mitral Insufficiency in the Dog. PhD Thesis. University of London. London.
- Morgan, H.E. & Baker, K.M. (1991). Cardiac hypertrophy. Mechanical, neural, and endocrine dependence. *Circulation* 83(1), 13-25.
- Mow, T. & Pedersen, H.D. (1999). Increased endothelin-receptor density in myxomatous canine mitral valve leaflets. *J Cardiovasc Pharmacol* 34(2), 254-60.
- Mukherjee, D. & Sen, S. (1990). Collagen phenotypes during development and regression of myocardial hypertrophy in spontaneously hypertensive rats. *Circ Res* 67(6), 1474-80.
- Münich, J. (1935). Anatomische untersuchungen über endokarditis valvularis beim Hunde. München, Germany: Ludwig-Maximilians-Universität.
- Nagase, H. (1997). Activation mechanisms of matrix metalloproteinases. *Biol Chem* 378(3-4), 151-60.
- Nagase, H. & Woessner, J.F., Jr. (1999). Matrix metalloproteinases. J Biol Chem 274(31), 21491-4.
- Nagatomo, Y., Carabello, B.A., Coker, M.L., McDermott, P.J., Nemoto, S., Hamawaki, M. & Spinale, F.G. (2000). Differential effects of pressure or volume overload on myocardial MMP levels and inhibitory control. *Am J Physiol Heart Circ Physiol* 278(1), H151-61.
- Nakayama, T., Wakao, Y., Takiguchi, S., Uechi, M., Tanaka, K. & Takahashi, M. (1994). Prevalence of valvular regurgitation in normal Beagle dogs detected by color Doppler echocardiography. J Vet Med Sci 56(5), 973-5.
- Nygaard, H., Thuesen, L., Hasenkam, J., Pedersen, E. & Paulsen, P. (1993). Assessing the severity of aortic valve stenosis by spectral analysis of cardiac murmurs (spectral vibrocardiography). Part I: Technical aspects. J Heart Valve Dis 2(4), 454-67.
- O'Brien, P.J., Landt, Y. & Ladenson, J.H. (1997). Differential reactivity of cardiac and skeletal muscle from various species in a cardiac troponin I immunoassay. *Clin Chem* 43(12), 2333-8.
- O'Brien, P.J., Smith, D.E., Knechtel, T.J., Marchak, M.A., Pruimboom-Brees, I., Brees, D.J., Spratt, D.P., Archer, F.J., Butler, P., Potter, A.N., Provost, J.P., Richard, J., Snyder, P.A. & Reagan, W.J. (2006). Cardiac troponin I is a sensitive, specific biomarker of cardiac injury in laboratory animals. Lab Anim 40(2), 153-71.
- O'Gara, P., Sugeng, L., Lang, R., Sarano, M., Hung, J., Raman, S., Fischer, G., Carabello, B., Adams, D. & Vannan, M. (2008). The role of

imaging in chronic degenerative mitral regurgitation. JACC Cardiovasc Imaging 1(2), 221-37.

- Olivetti, G., Giordano, G., Corradi, D., Melissari, M., Lagrasta, C., Gambert, S.R. & Anversa, P. (1995). Gender differences and aging: effects on the human heart. *J Am Coll Cardiol* 26(4), 1068-79.
- Olivetti, G., Melissari, M., Capasso, J.M. & Anversa, P. (1991). Cardiomyopathy of the aging human heart. Myocyte loss and reactive cellular hypertrophy. *Circ Res* 68(6), 1560-8.
- Olsen, L., Martinussen, T. & Pedersen, H. (2003). Early echocardiographic predictors of myxomatous mitral valve disease in Dachshunds. *Vet Rec* 152(10), 293-7.
- Olsen, L.H., Fredholm, M. & Pedersen, H.D. (1999). Epidemiology and inheritance of mitral valve prolapse in Dachshunds. *J Vet Intern Med* 13(5), 448-56.
- Olsen, L.H., Häggström, J. & Pedersen, H.D. (2010). Acquired valvular heart disease. In: Ettinger, S.J., et al. (Eds.) *Textbook of veterinary internal medicine*. St.Louis, Missouri: Elsevier Saunders. 7th ed. Vol 2. pp. 1299-1319.
- Opie, L.H. (2002). The neuroendocrinology of congestive heart failure. *Cardiovasc J S Afr* 13(4), 171-8.
- Opie, L.H., Commerford, P.J., Gersh, B.J. & Pfeffer, M.A. (2006). Controversies in ventricular remodelling. *Lancet* 367(9507), 356-67.
- Otsuji, Y., Handschumacher, M.D., Schwammenthal, E., Jiang, L., Song, J.K., Guerrero, J.L., Vlahakes, G.J. & Levine, R.A. (1997). Insights from three-dimensional echocardiography into the mechanism of functional mitral regurgitation: direct in vivo demonstration of altered leaflet tethering geometry. *Circulation* 96(6), 1999-2008.
- Oyama, M.A. & Chittur, S.V. (2006). Genomic expression patterns of mitral valve tissues from dogs with degenerative mitral valve disease. *Am J Vet Res* 67(8), 1307–18.
- Oyama, M.A. & Levy, R.J. (2010). Insights into serotonin signaling mechanisms associated with canine degenerative mitral valve disease. *J Vet Intern Med* 24(1), 27-36.
- Oyama, M.A. & Sisson, D.D. (2004). Cardiac troponin-I concentration in dogs with cardiac disease. J Vet Intern Med 18(6), 831-9.
- Pedersen, H.D., & Häggström, J. (2000). Mitral valve prolapse in the dog: a model of mitral valve prolapse in man. *Cardiovasc Res* 47(2), 234-43.
- Pedersen, H.D., Kristensen, B., Norby, B. & Lorentzen, K. (1996). Echocardiographic study of mitral valve prolapse in Dachshunds. Zentralbl Veterinarmed A 43(2), 103-10.
- Pedersen, H.D., Häggström, J., Falk, T., Mow, T., Olsen, L.H., Iversen, L. & Jensen, A.L. (1999). Auscultation in mild mitral regurgitation in dogs: observer variation, effects of physical maneuvers, and agreement with color Doppler echocardiography and phonocardiography. J Vet Intern Med 13(1), 56-64.

- Pelouch, V., Dixon, I.M., Golfman, L., Beamish, R.E. & Dhalla, N.S. (1993). Role of extracellular matrix proteins in heart function. *Mol Cell Biochem* 129(2), 101–20.
- Perloff, J.K. & Roberts, W.C. (1972). The mitral apparatus. Functional anatomy of mitral regurgitation. *Circulation* 46(2), 227-39.
- Perry, G. & Bouchard, A. (1990). Doppler echocardiographic evaluation of mitral regurgitation. *Cardiol Clin* 8(2), 265-75.
- Peterson, J.T., Hallak, H., Johnson, L., Li, H., O'Brien, P.M., Sliskovic, D.R., Bocan, T.M., Coker, M.L., Etoh, T. & Spinale, F.G. (2001). Matrix metalloproteinase inhibition attenuates left ventricular remodeling and dysfunction in a rat model of progressive heart failure. *Circulation* 103(18), 2303-9.
- Pfeffer, J.M., Pfeffer, M.A. & Braunwald, E. (1985). Influence of chronic captopril therapy on the infarcted left ventricle of the rat. *Circ Res* 57(1), 84-95.
- Piano, M.R., Bondmass, M. & Schwertz, D.W. (1998). The molecular and cellular pathophysiology of heart failure. *Heart Lung* 27(1), 3-19; quiz 20-1.
- Pierpont, G.L. & Talley, R.C. (1982). Pathophysiology of valvar heart disease. The dynamic nature of mitral valve regurgitation. Arch Intern Med 142(5), 998-1001.
- Pye, M., Rae, A.P. & Cobbe, S.M. (1990). Study of serum C-reactive protein concentration in cardiac failure. *Br Heart J* 63(4), 228-30.
- Rabkin, E., Aikawa, M., Stone, J.R., Fukumoto, Y., Libby, P. & Schoen, F.J. (2001). Activated interstitial myofibroblasts express catabolic enzymes and mediate matrix remodeling in myxomatous heart valves. *Circulation* 104(21), 2525-32.
- Rajakumar, K., Weisse, M., Rosas, A., Gunel, E., Pyles, L., Neal, W., Balian, A. & Einzig, S. (1999). Comparative study of clinical evaluation of heart murmurs by general pediatricians and pediatric cardiologists. *Clin Pediatr (Phila)* 38(9), 511-8.
- Rajamäki, M.M., Järvinen, A.K., Sorsa, T. & Maisi, P. (2002). Clinical findings, bronchoalveolar lavage fluid cytology and matrix metalloproteinase-2 and -9 in canine pulmonary eosinophilia. *Vet J* 163(2), 168-81.
- Re, G., Bergamasco, L., Badino, P., Borgarelli, M., Odore, R., Tarducci, A., Zanatta, R. & Girardi, C. (1999). Canine dilated cardiomyopathy: lymphocyte and cardiac alpha(1)- and betaadrenoceptor concentrations in normal and affected great danes. *Vet* J 158(2), 120-7.
- Ricchiuti, V., Zhang, J. & Apple, F.S. (1997). Cardiac troponin I and T alterations in hearts with severe left ventricular remodeling. *Clinical chemistry* 43(6 Pt 1), 990-5.

- Richman, J. & Moorman, J. (2000). Physiological time-series analysis using approximate entropy and sample entropy. *Am J Physiol Heart Circ Physiol* 278(6), H2039-49.
- Ries, C. & Petrides, P.E. (1995). Cytokine regulation of matrix metalloproteinase activity and its regulatory dysfunction in disease. *Biol Chem Hoppe Seyler* 376(6), 345-55.
- Robinson, T.F., Cohen-Gould, L. & Factor, S.M. (1983). Skeletal framework of mammalian heart muscle. Arrangement of inter- and pericellular connective tissue structures. *Lab Invest* 49(4), 482-98.
- Robinson, T.F., Factor, S.M. & Sonnenblick, E.H. (1986). The heart as a suction pump. *Scientific American* 254(6), 84-91.
- Rush, J.E., Lee, N.D., Freeman, L.M. & Brewer, B. (2006). C-reactive protein concentration in dogs with chronic valvular disease. *J Vet Intern Med* 20(3), 635-9.
- Sabbah, H. & Stein, P. (1976). Investigation of the theory and mechanism of the origin of the second heart sound. *Circ Res* 39(6), 874-82.
- Sabbah, H.N., Sharov, V.G., Lesch, M. & Goldstein, S. (1995). Progression of heart failure: a role for interstitial fibrosis. *Mol Cell Biochem* 147(1-2), 29-34.
- Sakkinen, P., Abbott, R.D., Curb, J.D., Rodriguez, B.L., Yano, K. & Tracy, R.P. (2002). C-reactive protein and myocardial infarction. *J Clin Epidemiol* 55(5), 445-51.
- Schafer-Somi, S., Ali Aksoy, O., Patzl, M., Findik, M., Erunal-Maral, N., Beceriklisoy, H.B., Polat, B. & Aslan, S. (2005). The activity of matrix metalloproteinase-2 and -9 in serum of pregnant and nonpregnant bitches. *Reprod Dom Anim* 40(1), 46-50.
- Schelling, P., Fischer, H. & Ganten, D. (1991). Angiotensin and cell growth: a link to cardiovascular hypertrophy? J Hypertens 9(1), 3-15.
- Schmelting, B., Niehoff, M., Egner, B., Korte, S.H. & Weinbauer, G.F. (2009). High Definition Oscillometry: a novel technique for noninvasive blood pressure monitoring in the cynomolgus monkey (Macaca fascicularis). J Med Primatol 38(5), 293-301.
- Schober, K.E., Cornand, C., Kirbach, B., Aupperle, H. & Oechtering, G. (2002). Serum cardiac troponin I and cardiac troponin T concentrations in dogs with gastric dilatation-volvulus. J Am Vet Med Assoc 221(3), 381-8.
- Schober, K.E., Kirbach, B. & Oechtering, G. (1999). Noninvasive assessment of myocardial cell injury in dogs with suspected cardiac contusion. J Vet Cardiol 1(2), 17-25.
- Schulz, O., Paul-Walter, C., Lehmann, M., Abraham, K., Berghofer, G., Schimke, I. & Jaffe, A.S. (2007). Usefulness of detectable levels of troponin, below the 99th percentile of the normal range, as a clue to the presence of underlying coronary artery disease. *Am J Cardiol* 100(5), 764-9.

- Segev, G., Ohad, D.G., Shipov, A., Kass, P.H. & Aroch, I. (2008). Cardiac arrhythmias and serum cardiac troponins in Vipera palaestinae envenomation in dogs. J Vet Intern Med 22(1), 106-13.
- Selig, M. (1993). Stethoscopic and phonoaudio devices: historical and future perspectives. *Am Heart J* 126(1), 262–268.
- Sengupta, P.P., Tajik, A.J., Chandrasekaran, K. & Khandheria, B.K. (2008). Twist mechanics of the left ventricle: principles and application. *JACC Cardiovasc Imaging* 1(3), 366-76.
- Sharov, V.G., Sabbah, H.N., Shimoyama, H., Goussev, A.V., Lesch, M. & Goldstein, S. (1996). Evidence of cardiocyte apoptosis in myocardium of dogs with chronic heart failure. *American J Pathol* 148(1), 141-9.
- Sheikh, K., Smith, S.W., von Ramm, O. & Kisslo, J. (1991). Real-time, three-dimensional echocardiography: feasibility and initial use. *Echocardiography* 8(1), 119-25.
- Smith, S.C., Ladenson, J.H., Mason, J.W. & Jaffe, A.S. (1997). Elevations of cardiac troponin I associated with myocarditis. Experimental and clinical correlates. *Circulation* 95(1), 163-8.
- Soini, Y., Satta, J., Maatta, M. & Autio-Harmainen, H. (2001). Expression of MMP2, MMP9, MT1-MMP, TIMP1, and TIMP2 mRNA in valvular lesions of the heart. *J Pathol* 194(2), 225-31.
- Spinale, F.G. (2002). Matrix metalloproteinases: regulation and dysregulation in the failing heart. *Circ Res* 90(5), 520–30.
- Spinale, F.G., Coker, M.L., Heung, L.J., Bond, B.R., Gunasinghe, H.R., Etoh, T., Goldberg, A.T., Zellner, J.L. & Crumbley, A.J. (2000). A matrix metalloproteinase induction/activation system exists in the human left ventricular myocardium and is upregulated in heart failure. *Circulation* 102(16), 1944-9.
- Spinale, F.G., Gunasinghe, H., Sprunger, P.D., Baskin, J.M. & Bradham, W.C. (2002). Extracellular degradative pathways in myocardial remodeling and progression to heart failure. J Card Fail 8(6 Suppl), S332-8.
- Spinale, F.G., Ishihra, K., Zile, M., DeFryte, G., Crawford, F.A. & Carabello, B.A. (1993). Structural basis for changes in left ventricular function and geometry because of chronic mitral regurgitation and after correction of volume overload. J Thorac Cardiovasc Surg 106(6), 1147-57.
- Spratt, D.P., Mellanby, R.J., Drury, N. & Archer, J. (2005). Cardiac troponin I: evaluation I of a biomarker for the diagnosis of heart disease in the dog. J Small Anim Pract 46(3), 139-45.
- Stewart, J.A., Jr., Wei, C.C., Brower, G.L., Rynders, P.E., Hankes, G.H., Dillon, A.R., Lucchesi, P.A., Janicki, J.S. & Dell'Italia, L.J. (2003). Cardiac mast cell- and chymase-mediated matrix metalloproteinase activity and left ventricular remodeling in mitral regurgitation in the dog. J Mol Cell Cardiol 35(3), 311-9.

- Swenson, L., Häggström, J., Kvart, C. & Juneja, R.K. (1996). Relationship between parental cardiac status in Cavalier King Charles spaniels and prevalence and severity of chronic valvular disease in offspring. *J Am Vet Med Assoc* 208(12), 2009-12.
- Tarnow, I., Falk, T., Tidholm, A., Martinussen, T., Jensen, A.L., Olsen, L.H., Pedersen, H.D. & Kristensen, A.T. (2007). Hemostatic biomarkers in dogs with chronic congestive heart failure. J Vet Intern Med 21(3), 451-7.
- Tarnow, I., Olsen, L.H., Kvart, C., Höglund, K., Moesgaard, S.G., Kamstrup, T.S., Pedersen, H.D. & Häggström, J. (2009). Predictive value of natriuretic peptides in dogs with mitral valve disease. *Vet J* 180(2), 195-201.
- Thomas, C.V., Coker, M.L., Zellner, J.L., Handy, J.R., Crumbley, A.J., 3rd & Spinale, F.G. (1998). Increased matrix metalloproteinase activity and selective upregulation in LV myocardium from patients with end-stage dilated cardiomyopathy. *Circulation* 97(17), 1708-15.
- Thomas, D.P., Zimmerman, S.D., Hansen, T.R., Martin, D.T. & McCormick, R.J. (2000). Collagen gene expression in rat left ventricle: interactive effect of age and exercise training. *J Appl Physiol* 89(4), 1462-8.
- Thomas, W., Gaber, C., Jacobs, G., Kaplan, P., Lombard, C., Moise, N. & Moses, B. (1993). Recommendations for standards in transthoracic two-dimensional echocardiography in the dog and cat. Echocardiography Committee of the Specialty of Cardiology, American College of Veterinary Internal Medicine. *J Vet Intern Med* 7(4), 247-52.
- Tidholm, A., Ljungvall, I., Höglund, K., Westling, A.B. & Häggström, J. (2009). Tissue Doppler and strain imaging in dogs with myxomatous mitral valve disease in different stages of congestive heart failure. *J Vet Intern Med* 23(6), 1197-207.
- Togashi, M., Tamura, K., Nitta, T., Ishizaki, M., Sugisaki, Y. & Fukuda, Y. (2007). Role of matrix metalloproteinases and their tissue inhibitor of metalloproteinases in myxomatous change of cardiac floppy valves. *Pathol Internat* 57(5), 251-9.
- Tsakiris, A.G., Von Bernuth, G., Rastelli, G.C., Bourgeois, M.J., Titus, J.L. & Wood, E.H. (1971). Size and motion of the mitral valve annulus in anesthetized intact dogs. *J Appl Physiol* 30(5), 611-8.
- Urabe, Y., Mann, D.L., Kent, R.L., Nakano, K., Tomanek, R.J., Carabello, B.A. & Cooper, G.t. (1992). Cellular and ventricular contractile dysfunction in experimental canine mitral regurgitation. *Circ Res* 70(1), 131-47.
- Walker, C.A. & Spinale, F.G. (1999). The structure and function of the cardiac myocyte: a review of fundamental concepts. J Thorac Cardiovasc Surg 118(2), 375-82.

- van Dalen, B.M., Kauer, F., Vletter, W.B., Soliman, O.I., van der Zwaan, H.B., Ten Cate, F.J. & Geleijnse, M.L. (2010). Influence of cardiac shape on left ventricular twist. J Appl Physiol 108(1), 146-51.
- Van der Laarse, A. (2002). Hypothesis: troponin degradation is one of the factors responsible for deterioration of left ventricular function in heart failure. *Cardiovasc Res* 56(1), 8-14.
- Weber, K.T. (1989). Cardiac interstitium in health and disease: the fibrillar collagen network. J Am Coll Cardiol 13(7), 1637-52.
- Weber, K.T., Anversa, P., Armstrong, P.W., Brilla, C.G., Burnett, J.C., Jr., Cruickshank, J.M., Devereux, R.B., Giles, T.D., Korsgaard, N., Leier, C.V. & et al. (1992). Remodeling and reparation of the cardiovascular system. J Am Coll Cardiol 20(1), 3-16.
- Weber, K.T., Pick, R., Janicki, J.S., Gadodia, G. & Lakier, J.B. (1988). Inadequate collagen tethers in dilated cardiopathy. *Am Heart J* 116(6 Pt 1), 1641-6.
- Venge, P., Johnston, N., Lagerqvist, B., Wallentin, L. & Lindahl, B. (2003). Clinical and analytical performance of the liaison cardiac troponin I assay in unstable coronary artery disease, and the impact of age on the definition of reference limits. A FRISC-II substudy. *Clin Chem* 49(6 Pt 1), 880-6.
- Whitney, J. (1974). Observations on the effect of age on the severity of heart valve lesions in the dog. J Small Anim Pract 15(8), 511-22.
- Whitney, J.C. (1976). Some aspects of the pathogenesis of canine arteriosclerosis. J Small Anim Pract 17(2), 87-97.
- Villari, B., Vassalli, G., Schneider, J., Chiariello, M. & Hess, O.M. (1997). Age dependency of left ventricular diastolic function in pressure overload hypertrophy. J Am Coll Cardiol 29(1), 181-6.
- Visse, R. & Nagase, H. (2003). Matrix metalloproteinases and tissue inhibitors of metalloproteinases: structure, function, and biochemistry. *Circ Res* 92(8), 827-39.
- Woessner, J.F., Jr. (1991). Matrix metalloproteinases and their inhibitors in connective tissue remodeling. *Faseb J* 5(8), 2145-54.
- Vu, T.H. & Werb, Z. (2000). Matrix metalloproteinases: effectors of development and normal physiology. *Genes Dev* 14(17), 2123-33.
- Zheng, J., Chen, Y., Pat, B., Dell'italia, L.A., Tillson, M., Dillon, A.R., Powell, P.C., Shi, K., Shah, N., Denney, T., Husain, A. & Dell'Italia, L.J. (2009). Microarray identifies extensive downregulation of noncollagen extracellular matrix and profibrotic growth factor genes in chronic isolated mitral regurgitation in the dog. *Circulation* 119(15), 2086-95.
- Zimmer, G., Zimmermann, R., Hess, O.M., Schneider, J., Kubler, W., Krayenbuehl, H.P., Hagl, S. & Mall, G. (1992). Decreased concentration of myofibrils and myofiber hypertrophy are structural determinants of impaired left ventricular function in patients with

chronic heart diseases: a multiple logistic regression analysis. J Am Coll Cardiol 20(5), 1135-42.

Acknowledgements

The present thesis was carried out at the Department of Clinical Sciences, Division of Small Animal Clinical Sciences, at the Swedish University of Agricultural Sciences, Uppsala, Sweden. Financial support was generously provided by the Department of Clinical Sciences and the Agria Research Foundation.

This thesis would not have been possible without the help and support from many people. In particular, I wish to express my sincere gratitude to:

Professor **Jens Häggström**, my main supervisor, for introducing me to the world of cardiology and scientific research. Your enormous expertise in these fields has been invaluable for me. Thank you for quickly responding all my answers, for always encouraging me to see the big picture and for stimulating me to go further with my thoughts and ideas. I appreciate all the scientifically challenging discussions we have had, as well as all the discussions on other subjects of life. I am grateful and honored for being supervised by you, and I highly appreciate your friendship.

Associate Professor **Katja Höglund**, my co-supervisor, for professionally and genuinely supporting my scientific and clinical work, for the valuable and thoroughly performed manuscript feed-back, and for always being available when I have needed to discuss any issue that might have arisen. I highly appreciate you as a colleague and friend.

Associate Professor **Anna Tidholm**, my co-supervisor, for generously sharing your knowledge in cardiology, and for introducing me to world of RT3D echocardiography. Your energy is admirable and I appreciate your positive attitude to life.

Associate Professor **Michele Borgarelli**, my co-supervisor, for all our interesting discussions about cardiology and science; for your quick manuscript-feedback, for inviting me to do research at Kansas State University, and for the great hospitality you and your family showed me during my stay there.

Professor Lisbeth Høier Olsen, my co-supervisor, for all our interesting discussions about cardiology and science, for always being friendly and inclusive and for showing me great hospitality during my stay at the University of Copenhagen.

Professor **Anne-Sofie Lagerstedt**, for placing the facilities of the Division of Small Animal Clinical Sciences at my disposal, and for being very supportive during these years.

Former and current Heads of the Department **Björn Ekesten and Torkel Ekman**, for placing the facilities of the Department of Clinical Sciences at my disposal.

Hospital Director of the University Animal Hospital **Mia Runnérus**, for placing the facilities at the hospital at my disposal.

Professor **Clarence Kvart**, for kindly introducing me to the art of cardiac auscultation, and for interesting discussions about cardiology.

The Linköping research group; Professor Per Ask, Associate Professor Peter Hult, and especially Christer Ahlström; for excellent collaboration on sound signal analysis. The scientific meeting between veterinary medicine and biomedical engineering was sometimes demanding, but you made it easier by patiently answering all my questions. I learned very much from working with you.

Professor **Per Venge** at the University of Uppsala, for guiding me in the world of cTnI analysis. I admire your knowledge within this field, and I appreciate your valuable comments on my work.

Dr **Minna Rajamäki,** for professionally running the MMP analyses at the University of Helsinki and for being a great co-author. I appreciate your enormous knowledge on the MMPs. I am happy that you could help me out when the MMP-ELISA assays failed to do so.

Serena Crosara, for the valuable collaboration on our PhD projects. I knew from the start that I had found a true friend in you.

Cristina Carnabuci, for being a highly appreciated co-worker at the cardiology clinic and a very supportive friend.

Professor **Lennart Jönson**, Dr **Fredrik Södersten**, and Dr T**orkel Falk**, for providing me with the best of answers in the topic of cardiac pathology.

Dr Harold Tvedten, Dr Inger Lilliehöök and, Josefine Öberg, for great collaborations. I am hoping for future exciting projects together with you.

Heikki Säteri, for kindly sharing your knowledge in cardiology with me. I appreciate your weekly presence at the cardiology specialist clinic.

Professor **Peter Lord** and Associate Professor **Kerstin Hanson**, for taking time to answer my questions on diagnostic imaging, and for giving me valuable scientific feedback.

All present and former colleagues and friends at the Department of Small Animal Clinical Sciences. Professor Åke Hedhammar; for caring about my scientific work and my future career, Bitten Andren; for teaching me a lot about internal medicine and for always being so helpful, Associate Professor Ragnvi Hagman; for all valuable advice and interesting discussion we have had, Associate Professor Helene Hamlin; for being so thoughtful, Associate Professor Henrik von Euler; for cheering up my days with wonderful café latte and music, and all my other highly valued colleagues in the corridor who are making a great atmosphere for laughter, interesting discussions, nice dog-walks, and so much more. I am privileged to have you as my colleagues.

Lena Pelander, for being such a fantastic room-mate, travel-companion, and friend, for your endless support, and for always saying the right things.

Kia Nostell, for all your support and for cheering up my work-days with laughter and interesting discussions on many different aspects of life.

Former and present PhD students at the department of clinical sciences, especially Odd Höglund, (for bringing laughter and fun to many PhD courses), Patricio Riviera, Tove Fall, Niklas Bergknut, and Annika Bergström. It was great sharing the post-graduate time with you. Teachers and co-participators at the PhD courses I have been taking part of during these years; for all interesting discussions about many different aspects of scientific work and personal development.

Maria Dimopolou, Malin Öhlund, Sara Larsdotter and Mia Norell, for all good times, both at work as well as outside. Ane Nødtvedt and Anna Djupsjöbacka; we truly miss you here: Please come back to the University and Uppsala!

Kjell-Åke Ahlin, for quickly helping me out tremendously many times when I have had computer problems.

Lena Moberg, Gunilla Drugge, Åsa Karlsson, Kirsi Laukkanen, Marta Kot, and Robert Kruse for your skilful technical assistance on biomarker analysis.

Dragica Ojurovic-Sutic and **Petter Olofsson** at Philips Healthcare, for your valuable advice and help on RT3D echocardiography.

The administration at the Department of Clinical Sciences, for all the great help I have received.

The KC library staff (in particular Michael Eklund), for excellent service.

Dogs and their owners; for so kindly providing me the data I needed for all the studies. This thesis would definitively not have been possible without you.

All co-workers at the University Animal Hospital (UDS), for contributing to a nice and friendly atmosphere at work.

Scientific colleagues around the world, for all inspiration you have given me from your published work.

Kicki and Örjan, for all your support during the years.

My parents, **Synnøve and Per**, my sister **Heidi** and my brother **Sverre**; for your constant support and love.

To all good friends and family outside my "work-zone"; although not mentioned specifically by name in this acknowledgement, you are very important to me!

Finally, to those who are closest to my heart: **Kalle**, for all your support, for your valuable scientific ideas, and for encouraging me when I have needed it. **Axel**, you are my everyday sunshine.